





#### Phenix User Workshop, Missoula, Montana



# Phenix Tools for crystallography

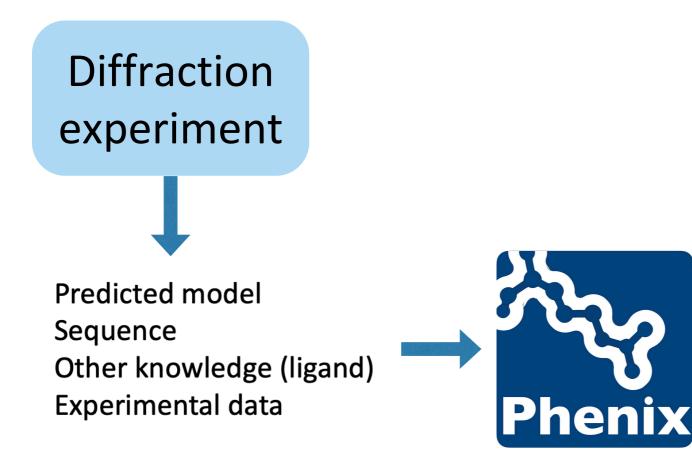
Dorothee Liebschner Lawrence Berkeley Laboratory

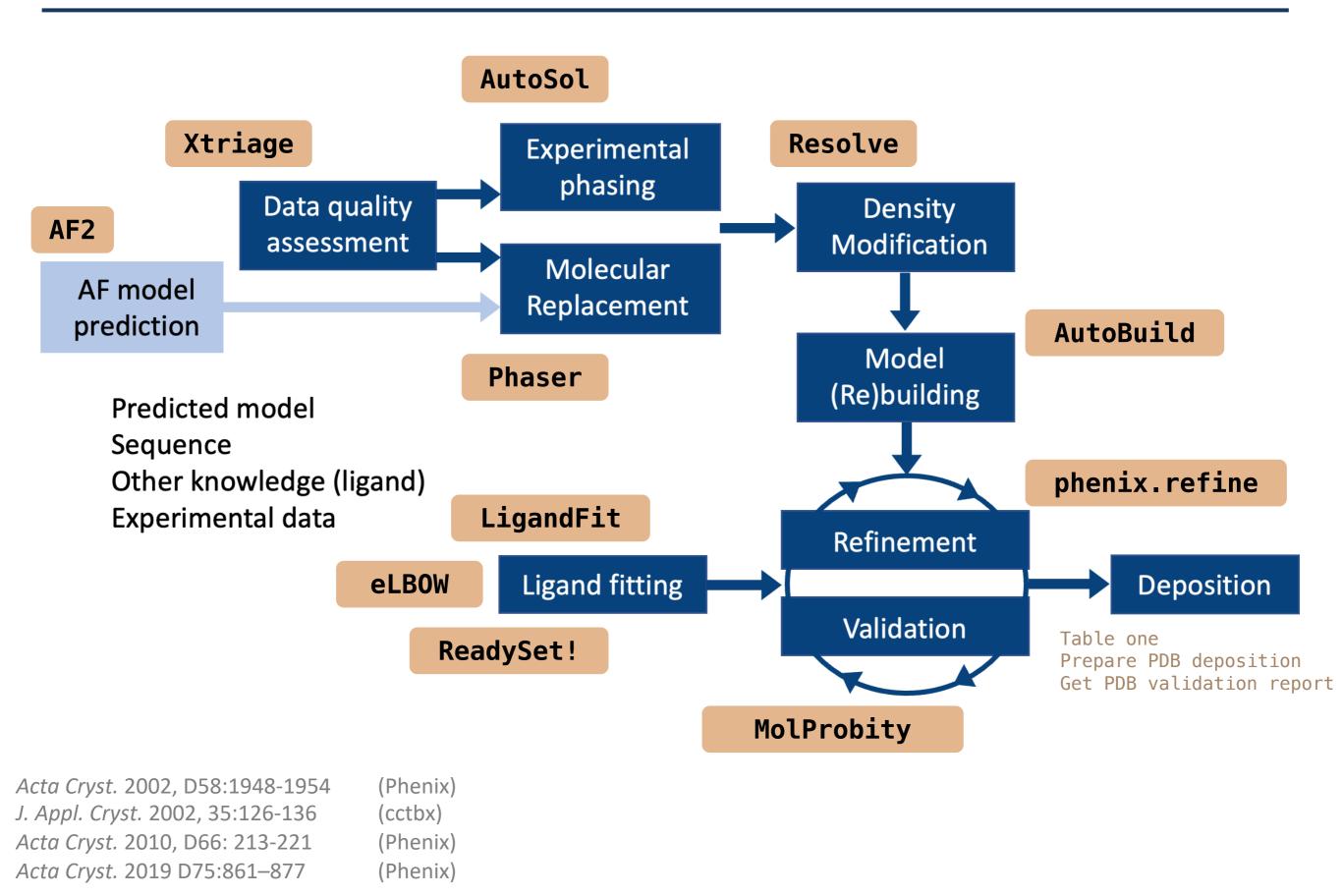


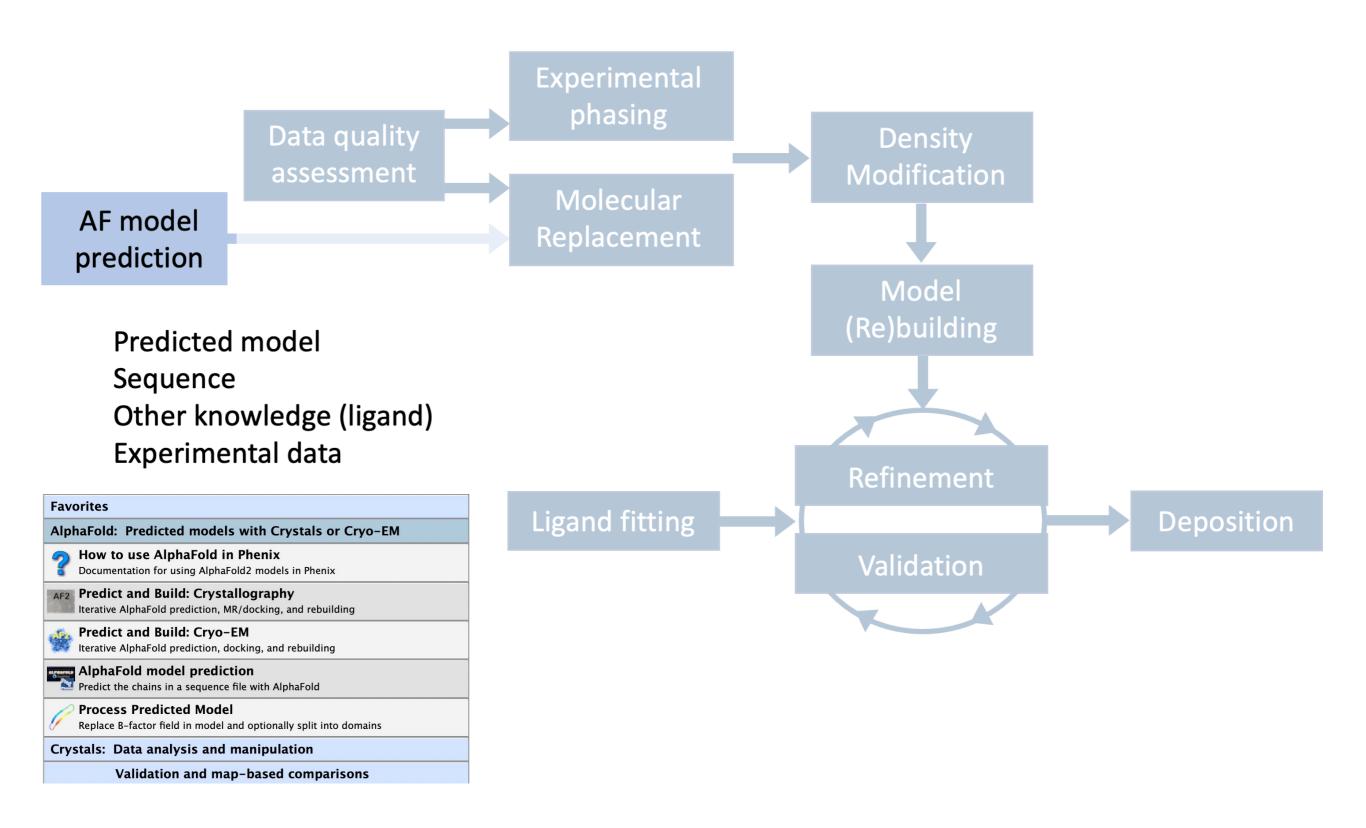




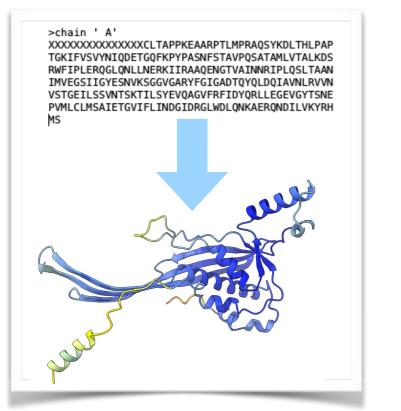








# Tools for predicted models in Phenix



# AlphaFold model prediction

Phenix server, no need to have AF2 installed locally

# Process predicted model

200

10

5

300

Scored residue

Expected position error (Ångströms)

15

400

20

25

500

30

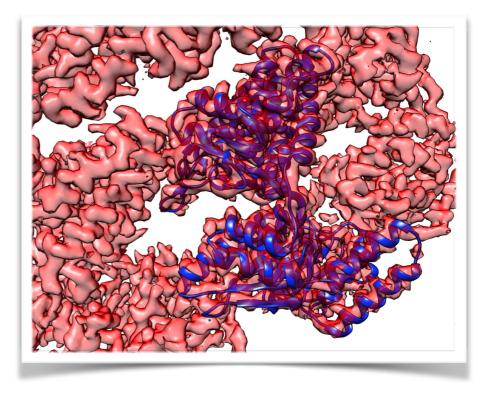
100

Aligned residue 00 00

400

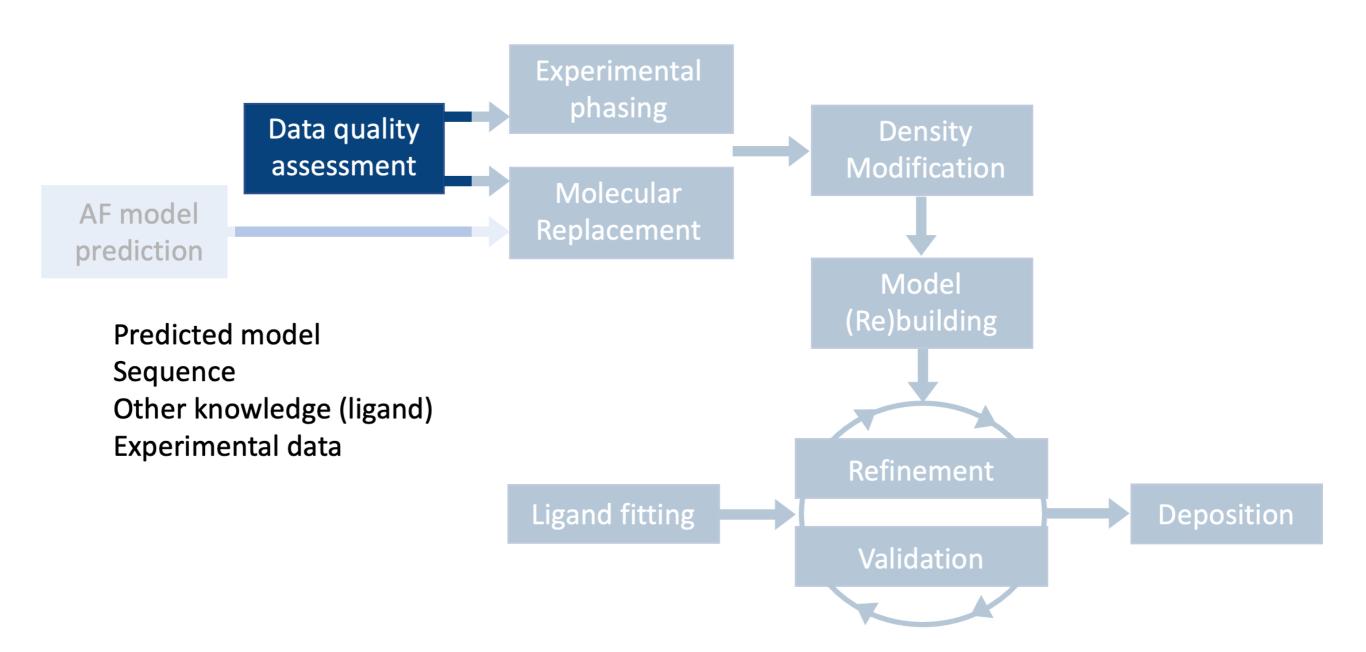
500

0

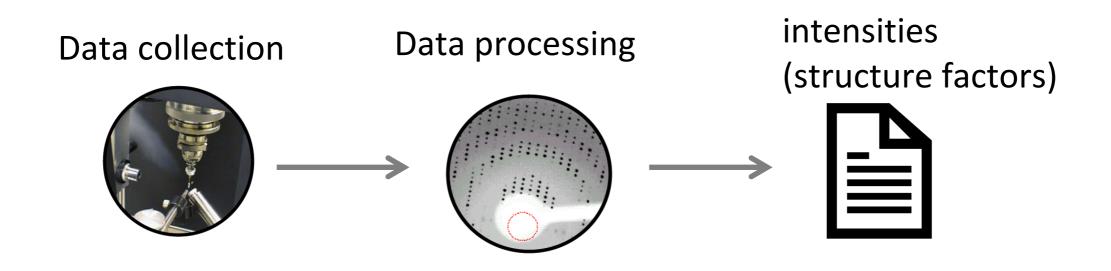


Predict and Build Iterative AlphaFold prediction, MR/docking, and rebuilding

#### Fully automatic!



#### Data Quality Assessment



#### Macromolecular crystals are prone to pathologies:

- Twinning: two or more crystals are intergrown (orientations are related by twin operation)
- tNCS: more than one copy of a molecule is in a similar orientation in the asymmetric unit

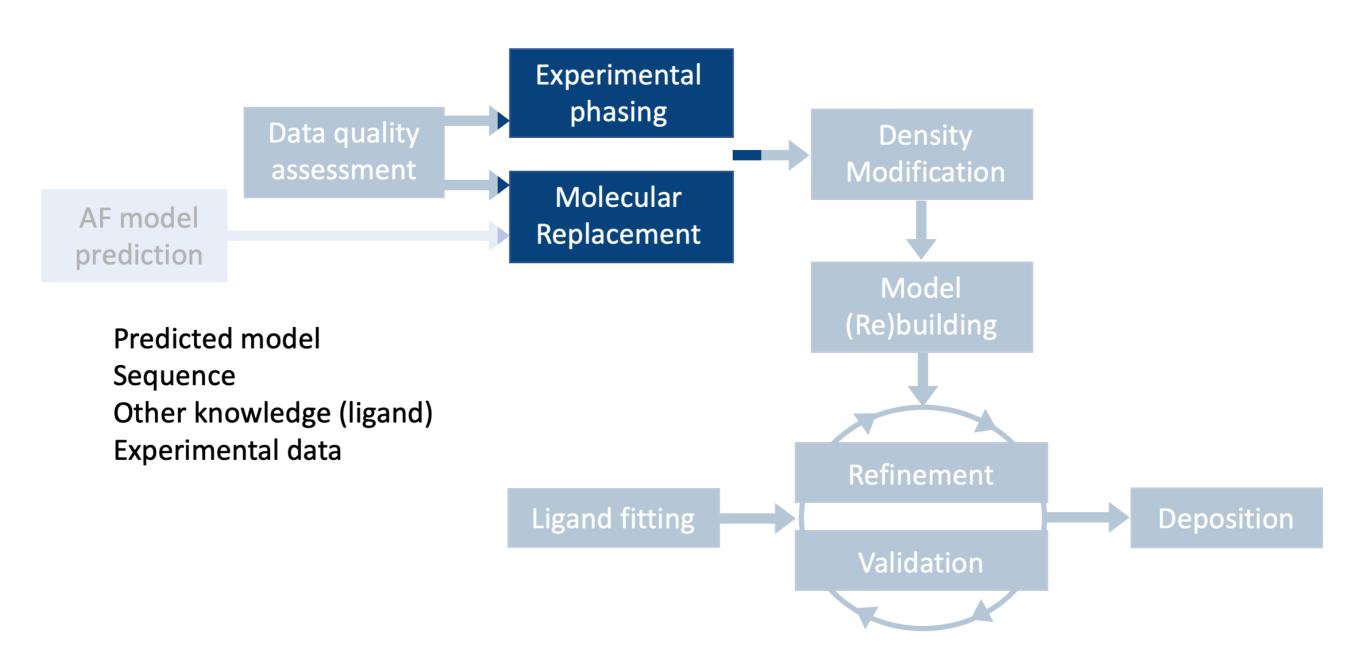
#### Data anomalies can prevent structure solution!

→ It is important to check your data before phasing, model building and refinement.

Xtriage does diagnostics for major pathologies and data properties (Wilson plot, completeness, symmetry).

Configure	Xtriage_1	4 Þ ×		
Run status	Results	4 Þ		
	Xtriage summary	Contraction (Contraction)		
	ensity statistics suggest twinning (intensities are n operators show a significant twin fraction.	e significantly different from expected for normal data) and one or more		
Trar	nslational NCS does not appear to be present.			
lce r	rings do not appear to be present.			
The	The fraction of outliers in the data is less than 0.1%. Click on panels to explore			
The	data are not significantly anisotropic.	the results and investigate		
The	resolution cutoff appears to be similar in all di	problems		
The	overall completeness in low-resolution shells	is at least some		
The	completeness is 99.76%.			

Please inspect all individual results closely, as it is difficult to automatically detect all issues.



Typically, the goal is to determine the **structure**. (arrangement of atoms in space)

The electron density in the unit cell is related to the Fourier transform of the **amplitude and phase of the scattered X-rays**.

$$\rho(\vec{r}) = FT\left(\vec{F}(\vec{H})\right) = \frac{1}{V}\int \vec{F} \cdot e^{-2i\pi\vec{H}\cdot\vec{r}}$$

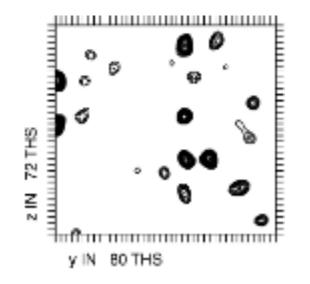
Typically, the goal is to determine the **structure**. (arrangement of atoms in space)

The electron density in the unit cell is related to the Fourier transform of the **amplitude and phase of the scattered X-rays**.

Unfortunately:  

$$\rho(\vec{r}) = FT\left(\vec{F}(\vec{H})\right) = \frac{1}{V} \int |F| e^{i\phi} \cdot e^{-2i\pi \vec{H} \cdot \vec{r}}$$
obtained from the experiment:  $I \propto |F|$ 

# How to recover phases



#### Experimentally

Exploit the properties of a few special atoms:

- anomalous scattering
- a large number of electrons

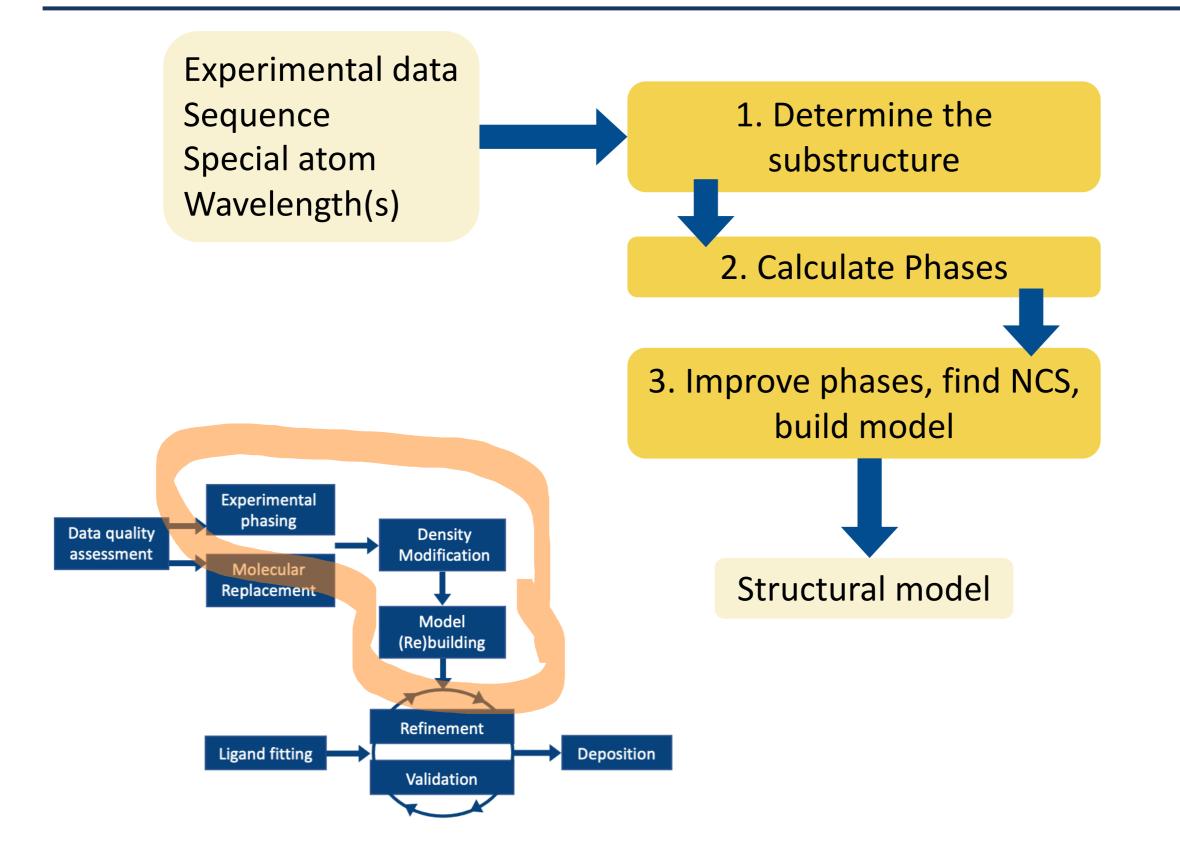
#### Computationally

• Molecular Replacement (MR)

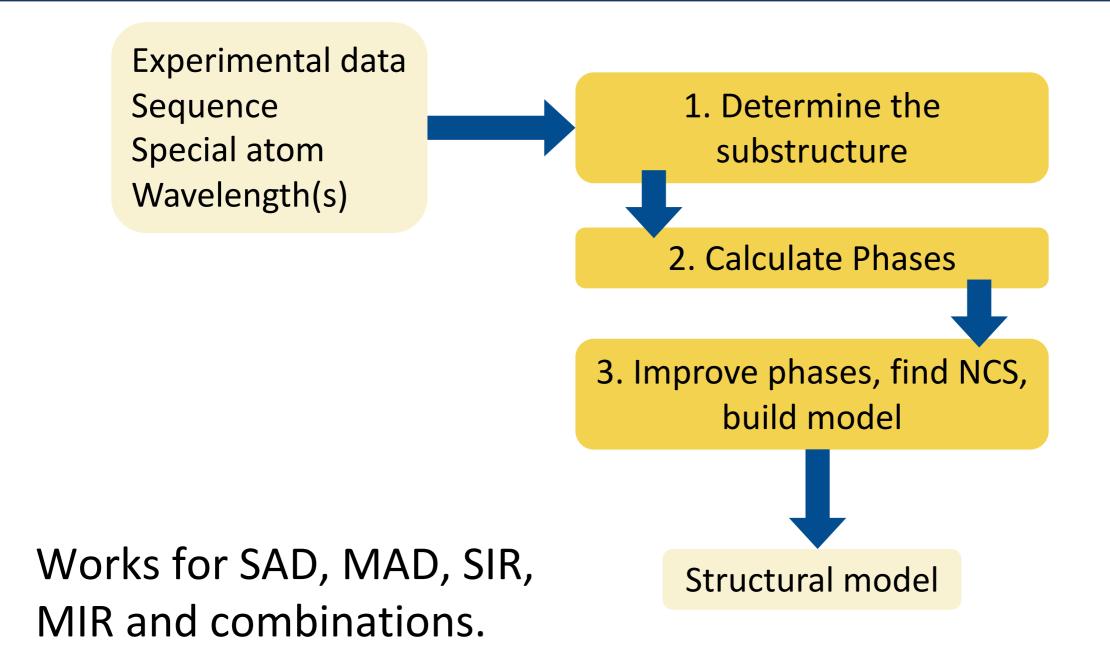
A previously known structure can provide initial phase estimates for a new structure

Direct Methods
 Phase relationships can be formulated by assuming the positivity and atomicity of the electron density

#### Experimental Phasing with AutoSol

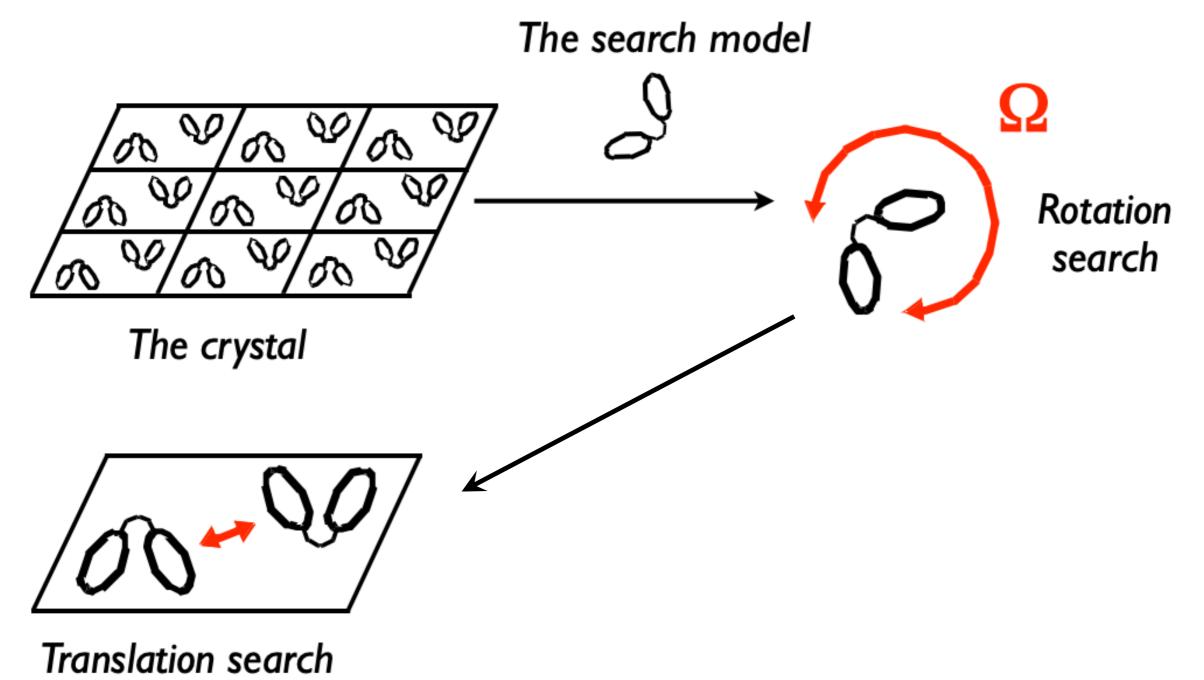


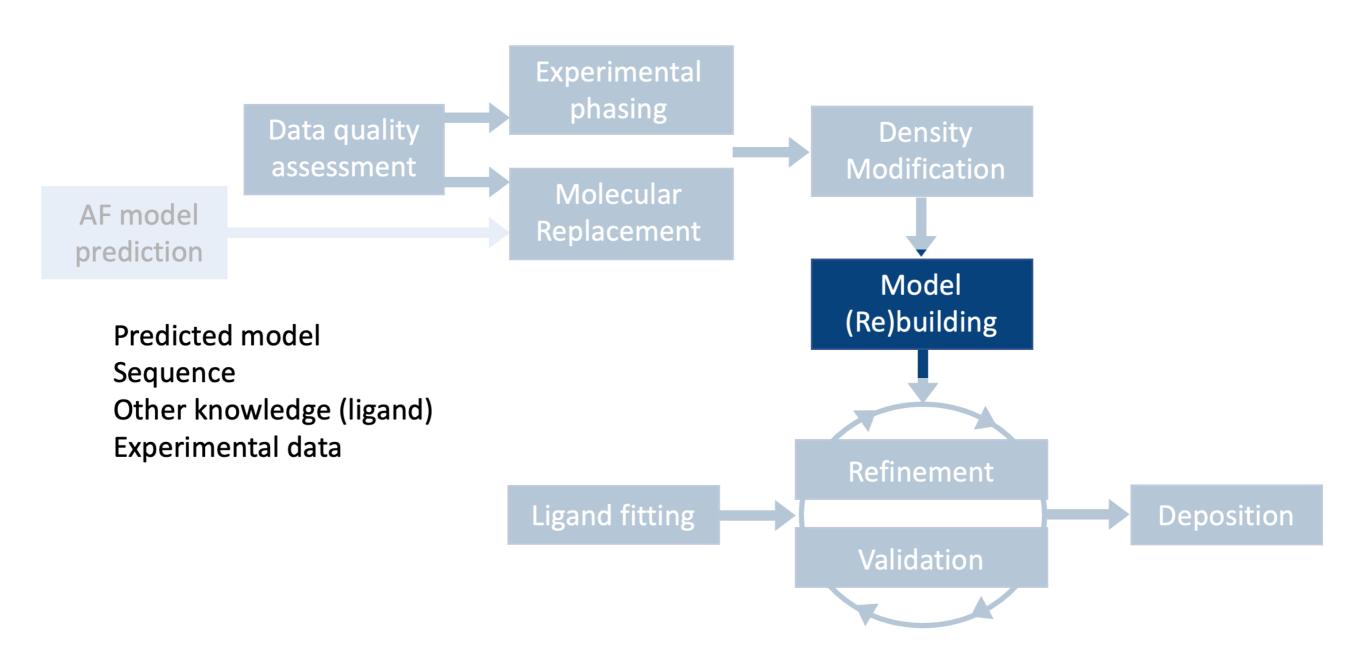
#### Experimental Phasing with AutoSol



This procedure is fully automatic!

Use a previously known structure to get phase estimates





# Model building

After phasing, we have an initial model. But sometimes it is not very good yet.

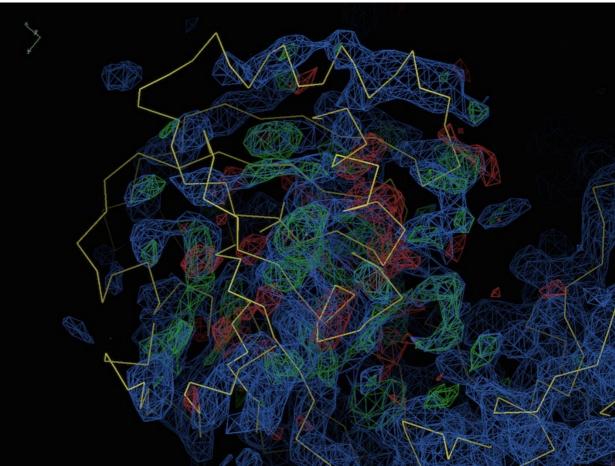
#### **Experimental phasing:**

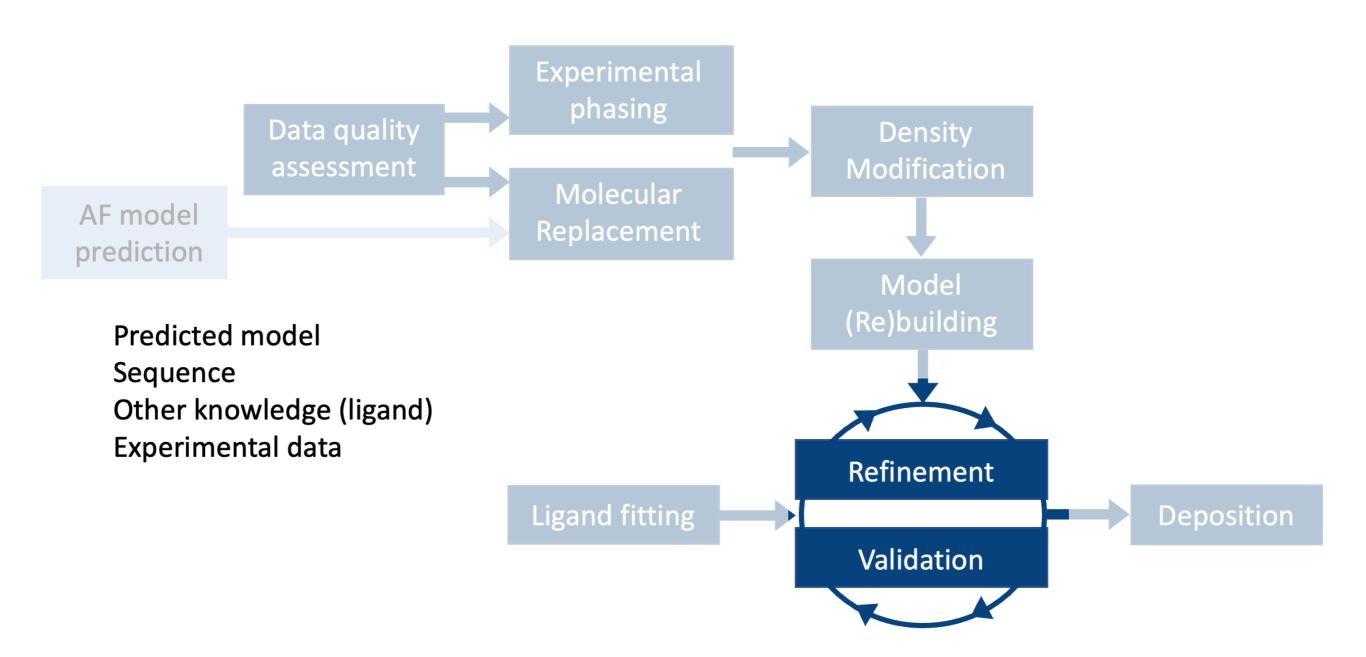
The AutoSol model can be most likely improved by doing a more thorough approach with AutoBuild.

#### **Molecular replacement:**

If the search model is of a significantly different protein and/or if there are large conformational changes, run AutoBuild.

#### → Run **AutoBuild**





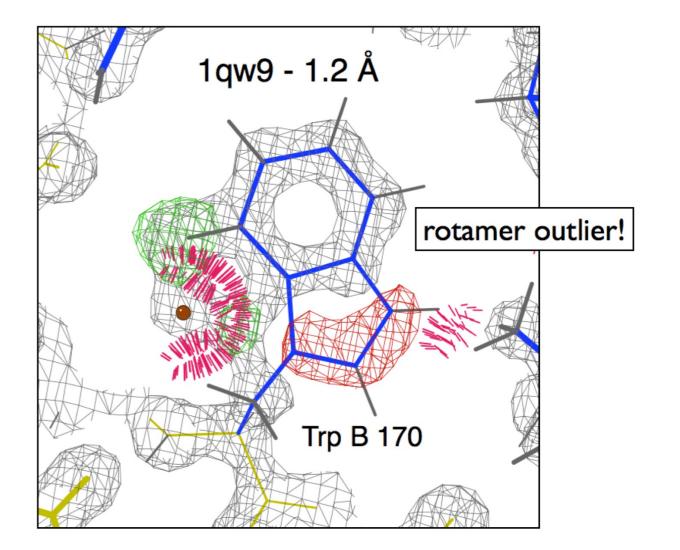
# **Refinement/Validation**

Refinement = Use an *optimization* algorithm to minimize a *target function* by changing the *parameters* of the model

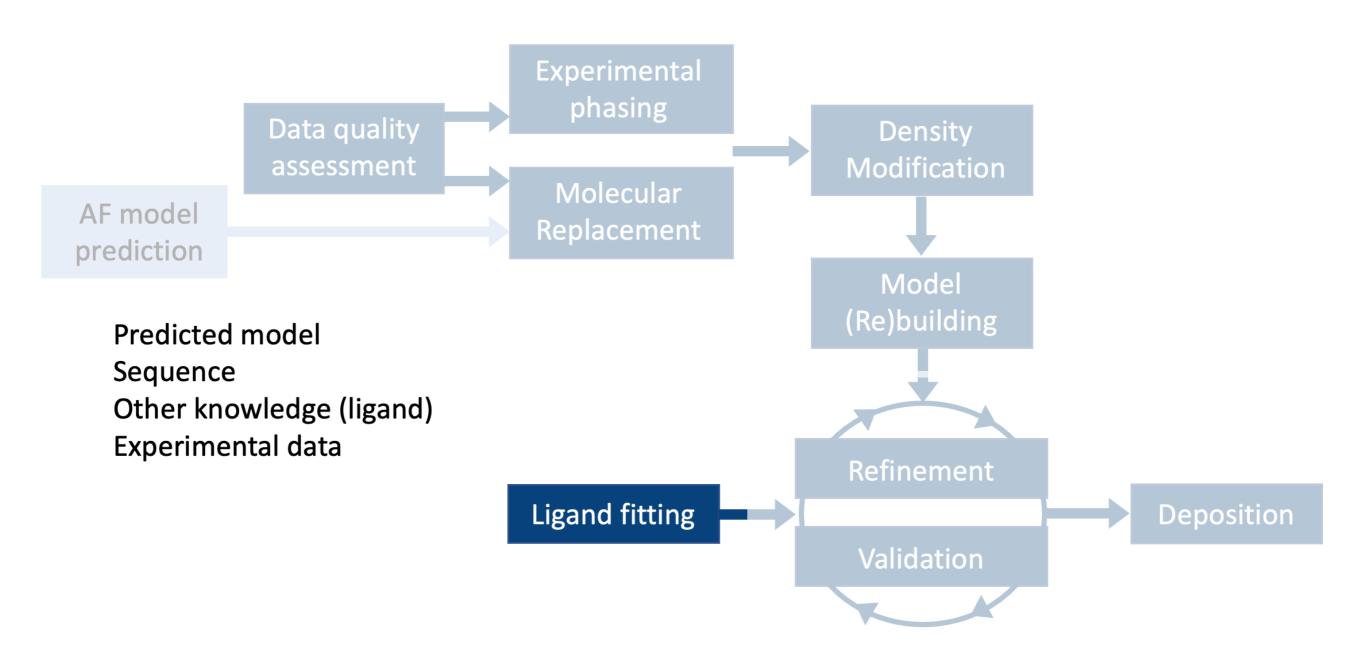
Optimize a model so that it optimally explains the data.

The model should be already quite "good".

After each refinement run, check the model for outliers/oddities.



\*Tronrud, Dale. E. (2004). Acta Cryst. D. 60, 2156–2168.

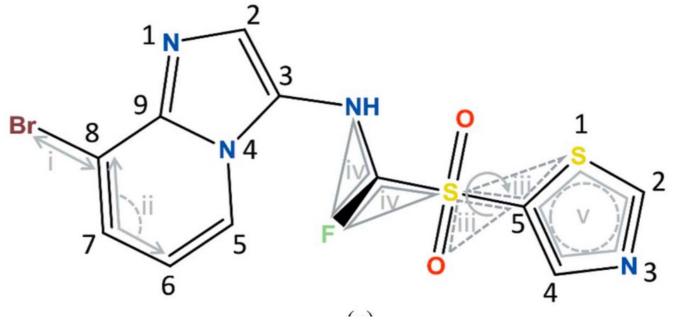


Models (usually) cannot be refined using diffraction data alone: high-resolution information is typically missing.

Use restraints to obtain chemically plausible structures.

Restraints = prior knowledge

Stereochemical information (e.g. bond distances, angles) about the ligand.



#### Ligands need restraints

If you try to run phenix.refine and a ligand has no restraints...

phenix.refine (Project:	presentations)				
Preferences Help Run Abort Save Graphics ReadySet TLS Restraints	Xtriage Ask for help				
Configure	↓ ▷ ×				
Input data Refinement settings Output Amber	4 Þ				
Job title :					
Input files	PHENIX	orror			
File path	PHENIA	enor			
Add file       Remove file       Modify file data type       Use symmetry from s         Space group :       Vinit cell :         X-ray data and experimental phases       R-free label :	the model file to resolve the p file with matching restraint de	Inknown nonbonded energy type symbols: 31 Please edit odel file to resolve the problems and/or supply a CIF th matching restraint definitions, along with			
Data labels :    High resolution :      Low resolution :	apply_cif_modification and a	pply_cif_link parameter			
Wavelength : Options	definitions if necessary.				
Neutron data		OK			
Data labels : 🕞 R-free label :		✓ OK			
High resolution : Low resolution :	options				
	Project: presentations				

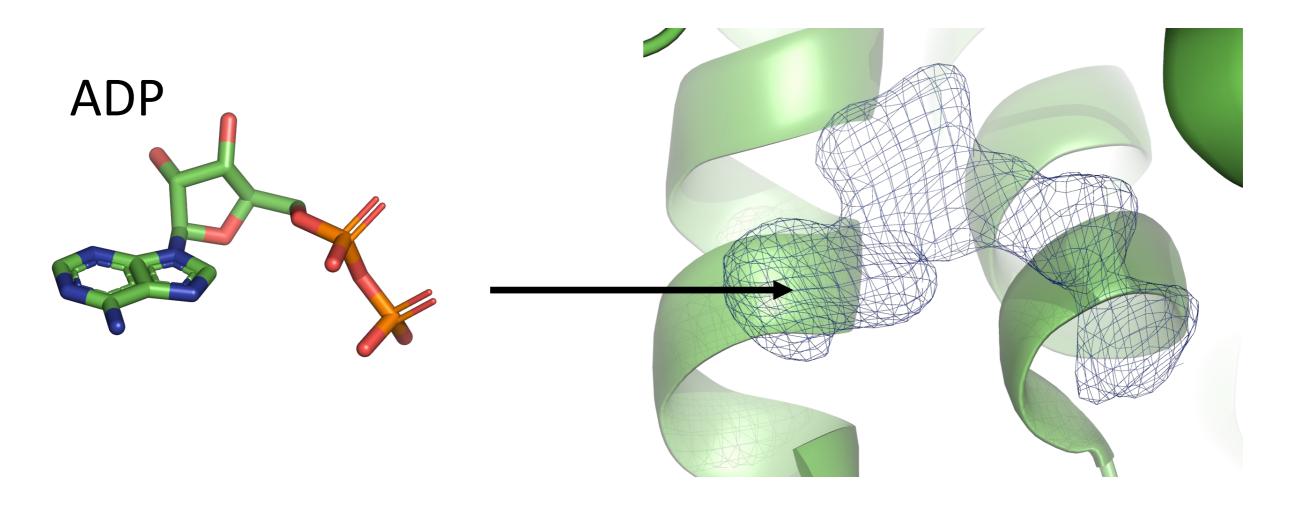
#### (This message also occurs if atom names don't match!)

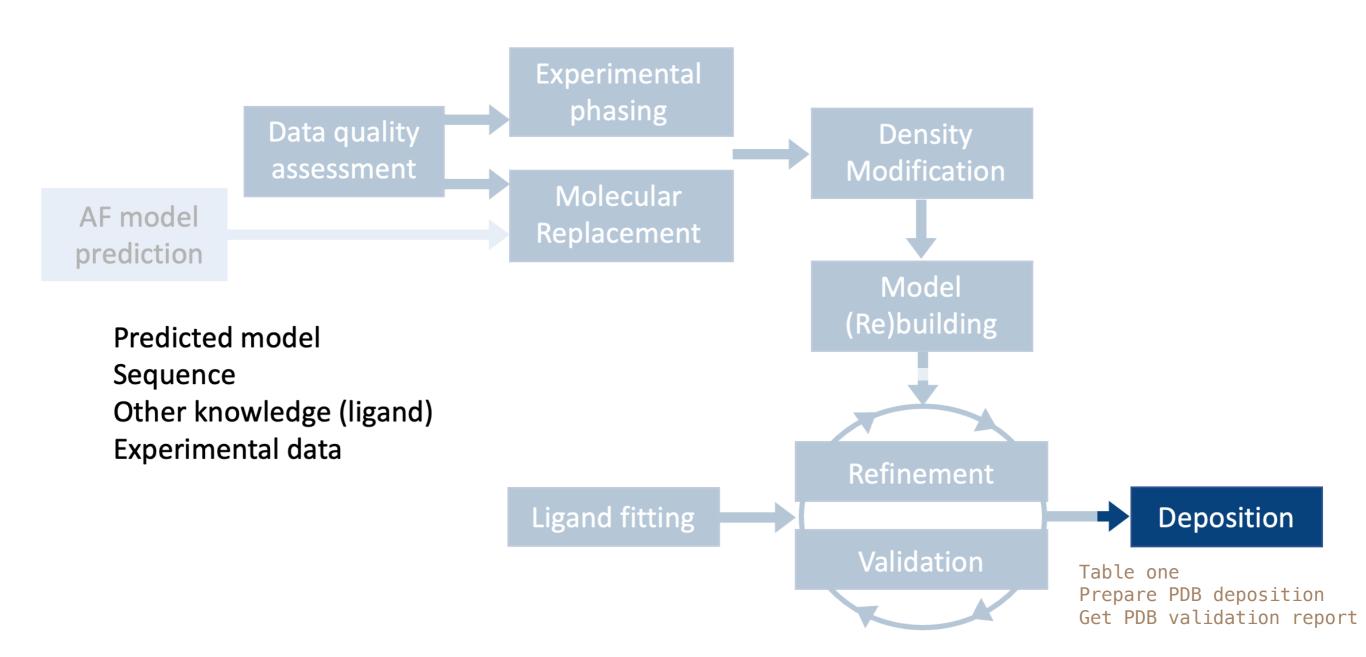
#### Other tools for ligands in Phenix

#### LigandFit: Flexible ligand fitting into difference maps.

Acta Crystallographica Section D Biological Crystallography ISSN 0907-4449 Automated ligand fitting by core-fragment fitting and extension into density

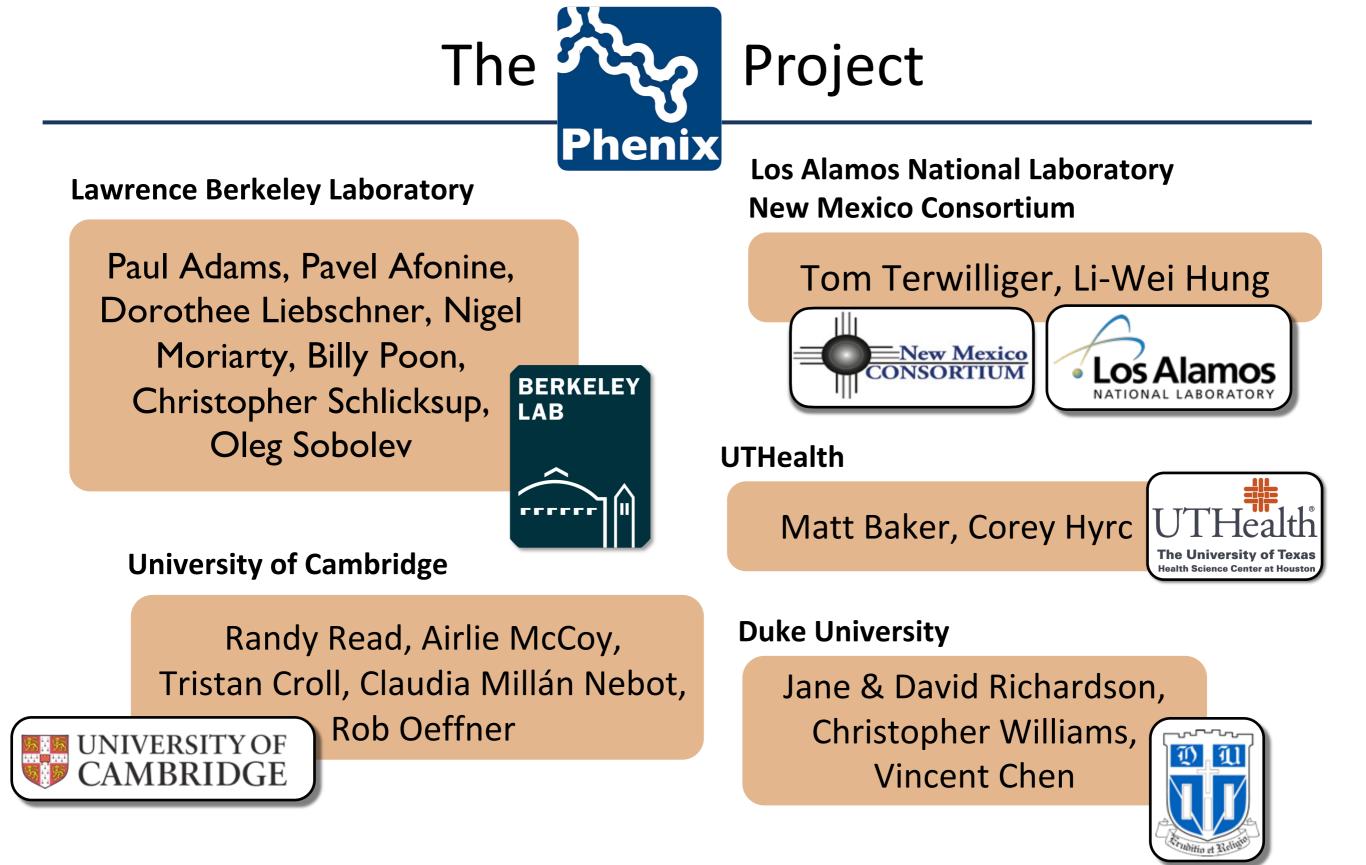
Thomas C. Terwilliger,<sup>a</sup>\* Herbert Klei,<sup>b</sup> Paul D. Adams,<sup>c</sup> Nigel W. Moriarty<sup>c</sup> and Judith D. Cohn<sup>a</sup> A procedure for fitting of ligands to electron-density maps by first fitting a core fragment of the ligand to density and then extending the remainder of the ligand into density is presented. The approach was tested by fitting 9327 ligands Received 26 January 2006 Accepted 9 May 2006





# PDB deposition

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Actions Job history	1			
Projects				maps (create, manipulate, compare)
				Enhanced maps (Polder, FEM, density-modified)
Show group: All groups 💿 Manage				Model building
Select 🖉 Dele	New project	🚽 Import project	🐼 Settings	Refinement
ID	Last modified	# of jobs R-free	•	Ligands
AF_POMGNT2_1	Jun 05 2024 11:46	3		Cryo-EM: Map analysis, symmetry, manipulation
bugs	May 30 2024 02:38.			
02_test_comma	May 24 2024 01:20			Validation and map-based comparisons
tests	May 22 2024 11:15		0	Map improvement
_	May 16 2024 10:37			
	Mar 19 2024 09:54			Docking, model building and rebuilding
	Mar 19 2024 09:28			Refinement
bugs_playground				Models: Superpose, search, compare, analyze symmetry
fmodel	Feb 28 2024 02:44			
SEACOAST	Feb 13 2024 01:09			Modification, minimization and dynamics
	Jan 03 2024 10:19			PDB Deposition
joint_XN	Nov 02 2023 03:49		9	Prepare model for PDB deposition
	Apr 13 2023 02:18			Finalize mmCIF files for deposition to the PDB
	Apr 13 2023 09:35			•
AF_POMGNT2_0				Get PDB validation report Retrieve a validation report from the PDB
AF_POMGNT2	Mar 30 2023 09:07			
7brm Zmia waabw	Mar 17 2023 11:39			Generate "Table 1" for journal
7mjs_wcsbw	Mar 17 2023 09:31 Mar 15 2023 02:00			Extraction of final model statistics for publication
presentation	Mar 15 2023 02:00 Mar 06 2023 03:23			Program search
bughaton	Mar 06 2023 03-23.	. 8	-	
	/Users/dcliebschner/D	ocuments/AF_POMGI	NT2_1	Browse Q
Phenix version 1.21.1-5	286-000			Project: AF_POMGNT2_1





Liebschner D, *et al.*, Macromolecular structure determination using X-rays, neutrons and electrons: recent developments in *Phenix*. Acta Cryst. 2019 **D75**:861–877