

# Validation Philosophy

- Visualizations > statistics
- Local conformations > structure-level averages
- “Outlier” thresholds are set statistically
  - Expect to see experimentally justified statistical outliers sometimes, especially at functional sites
  - Cherish these! You found something cool!

# Outline



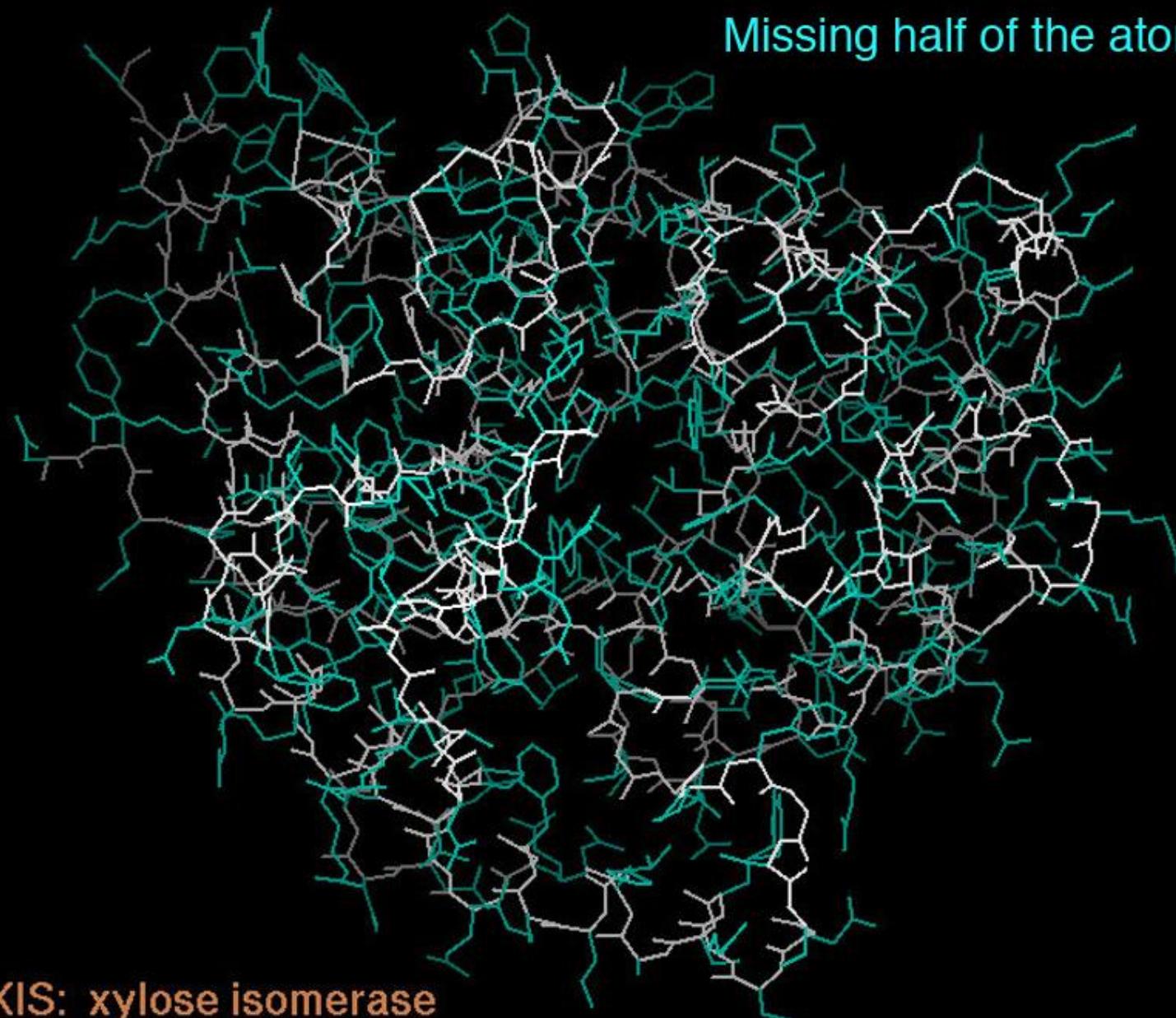
For each validation

- Method
  - Briefly, how the underlying idea or math works
- Visualization
  - How outliers are visually represented
- Probable causes
  - Example of a common or interesting type of error
  - Not comprehensive!

# All-Atom Clashes and Contacts

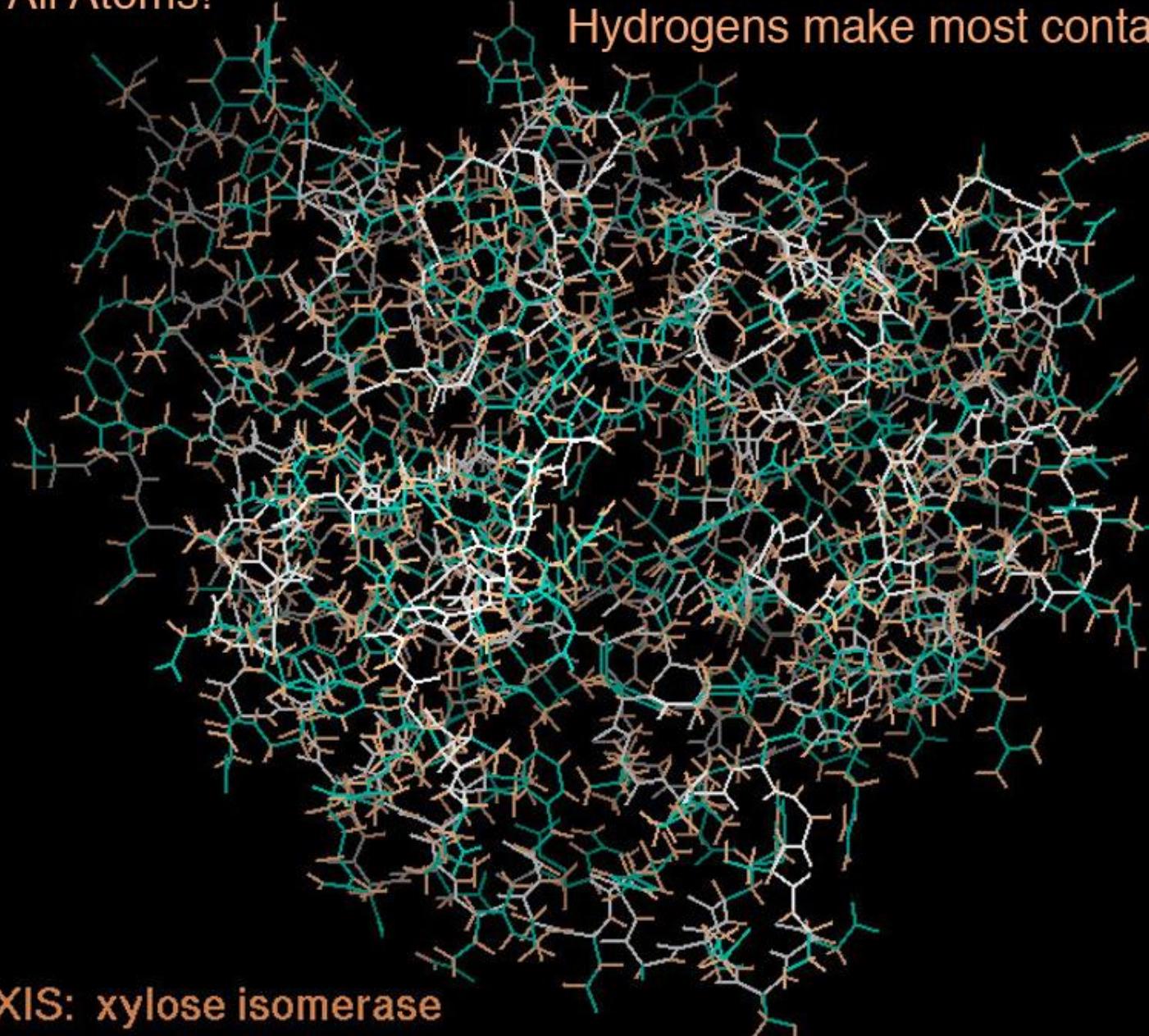
Add hydrogens  
with phenix.reduce

Missing half of the atoms!



All Atoms!

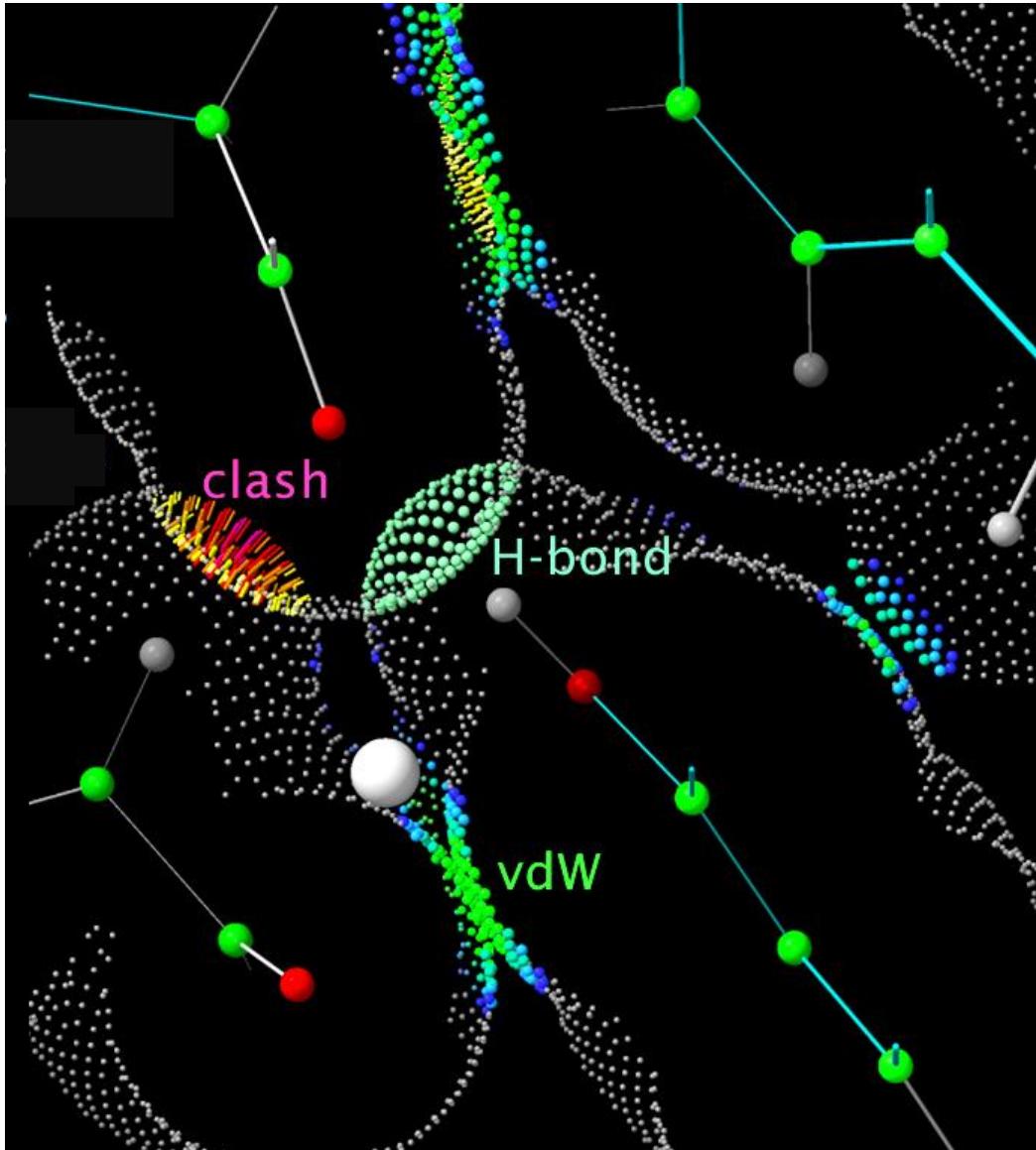
Hydrogens make most contacts



Hydrogens:  
"twigs  
on the  
tree"

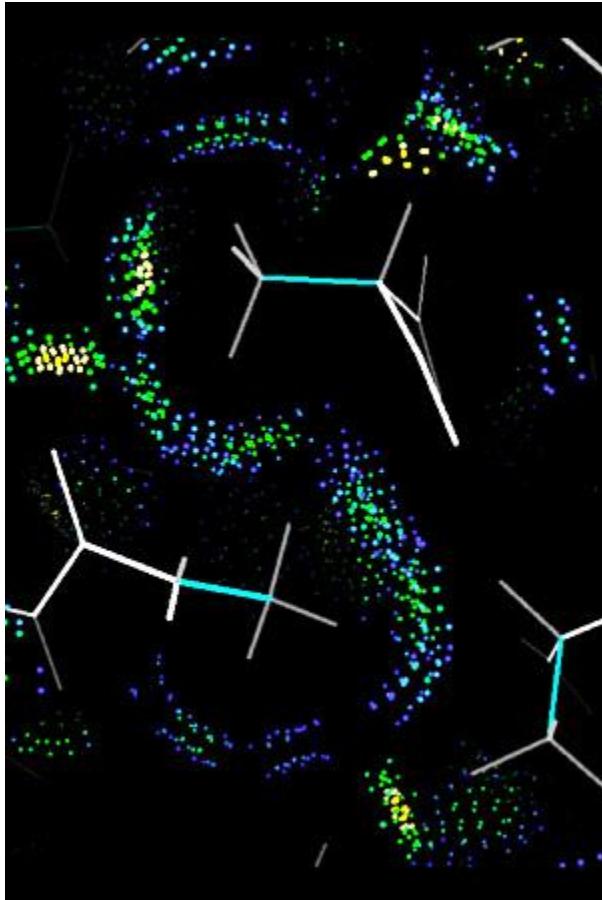
4XIS: xylose isomerase

# All-Atom Contacts and Clashes: Method

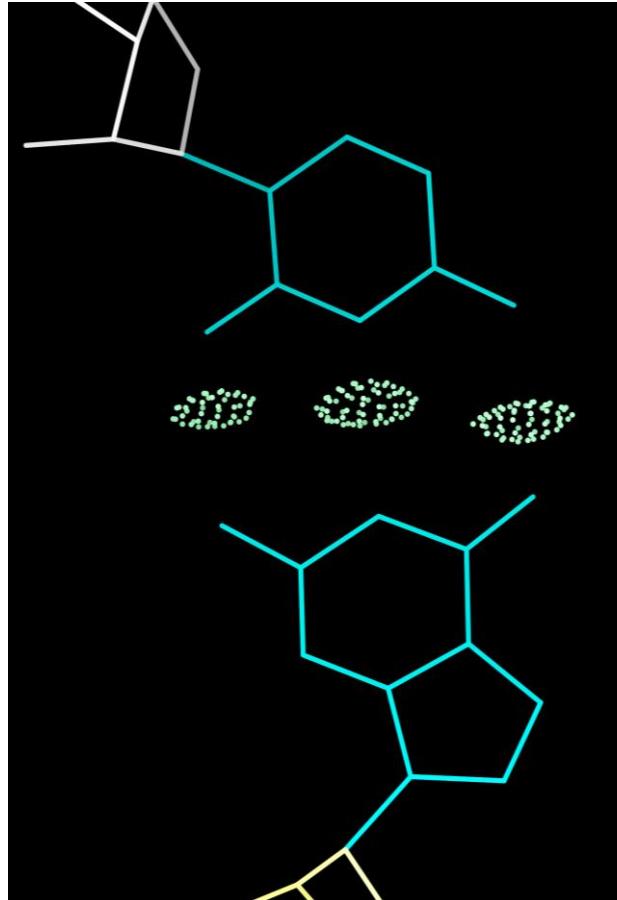


- Roll a  $0.25\text{\AA}$  radius “Probe” sphere over the van der Waals surface of each atom
- Mark where the probe touches or overlaps with another van der Waals surface
- Note that hydrogen atom surfaces can shield heavy atom surfaces

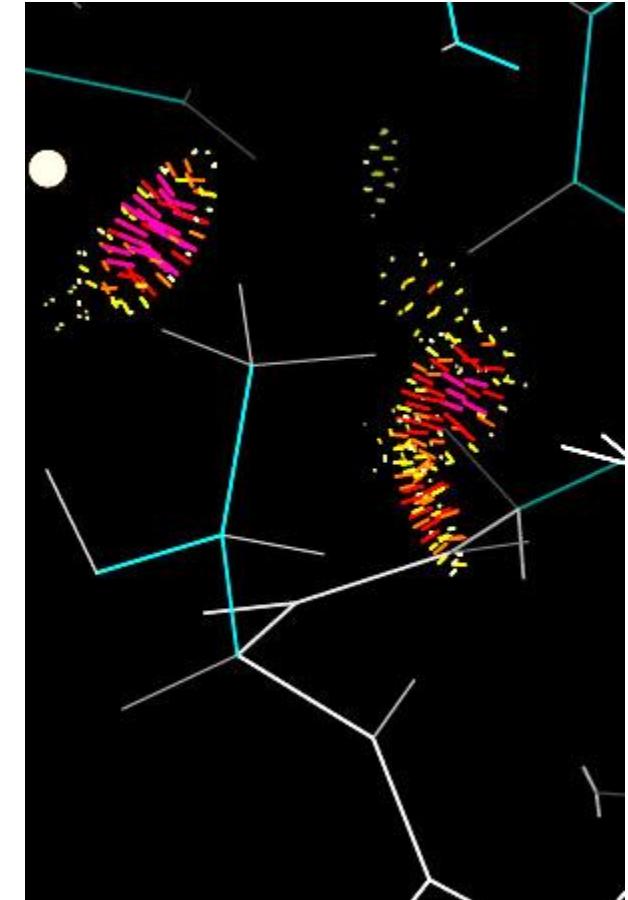
# All-Atom Contacts and Clashes: Visualization



Favorable vdW packing in  
greens and blues



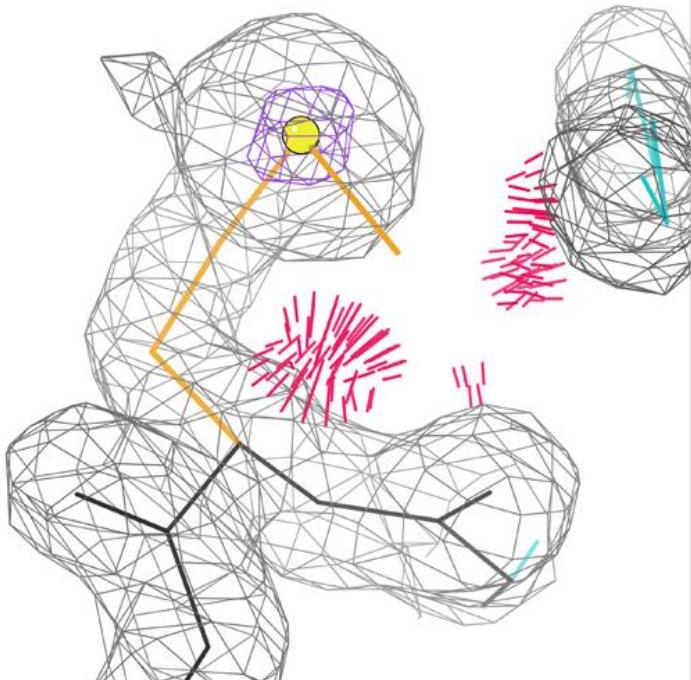
Favorable hydrogen bonding  
as light green pillows



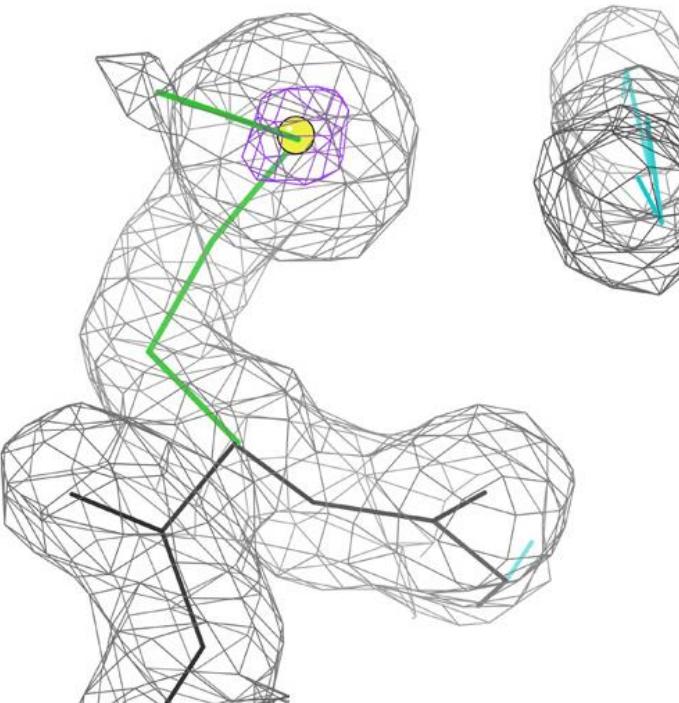
Steric overlaps, aka  
“clashes”, as hot pink spikes

# All-Atom Contacts and Clashes: Probable causes

original: !!



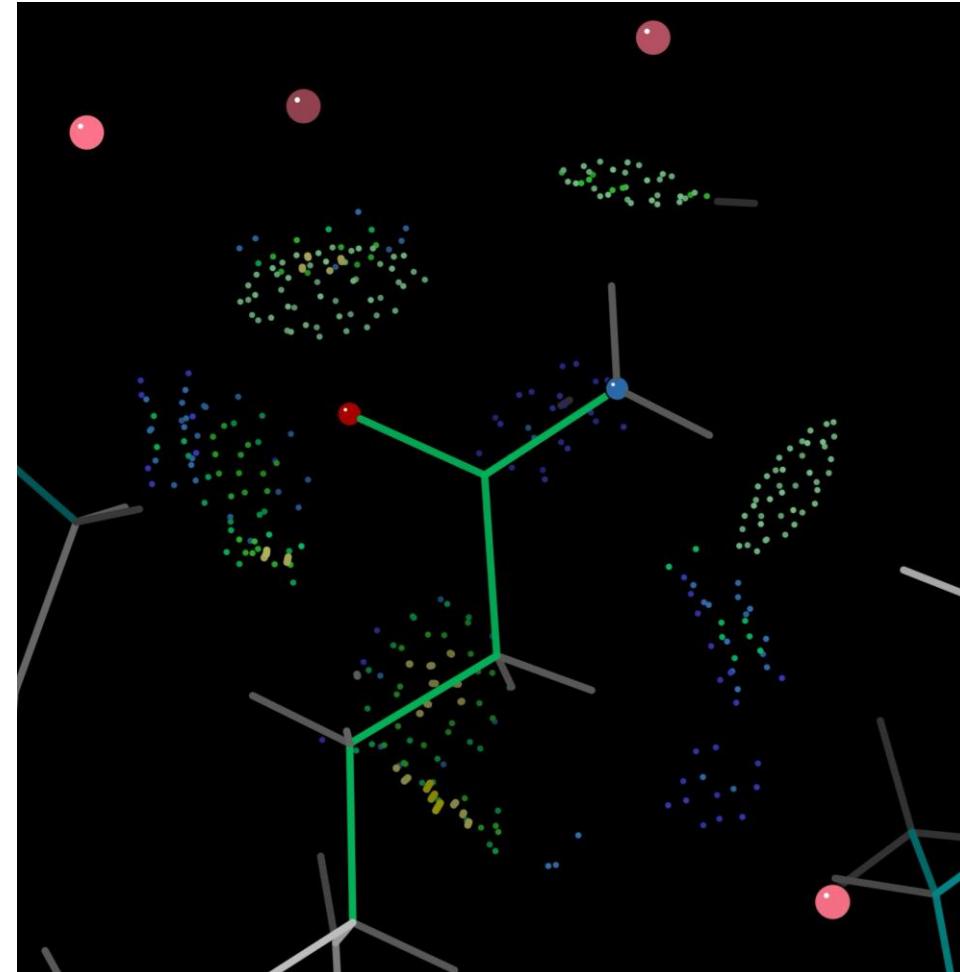
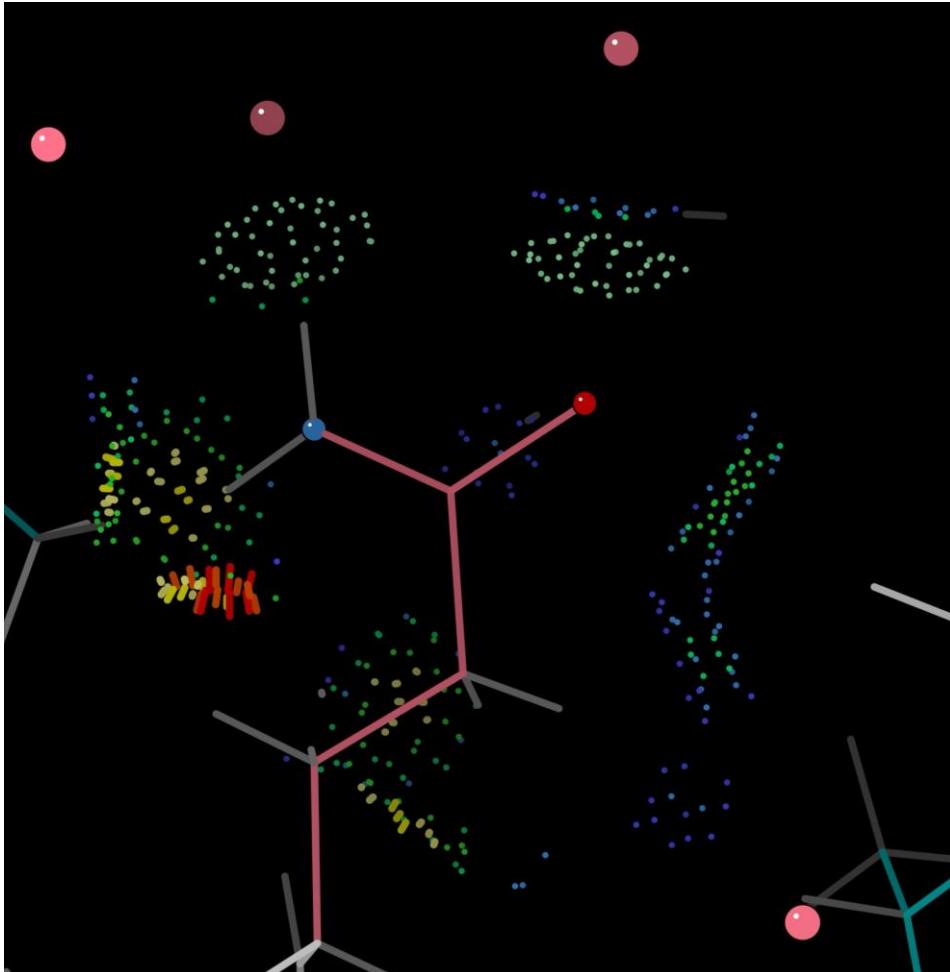
rebuilt: mmm



## Other outliers

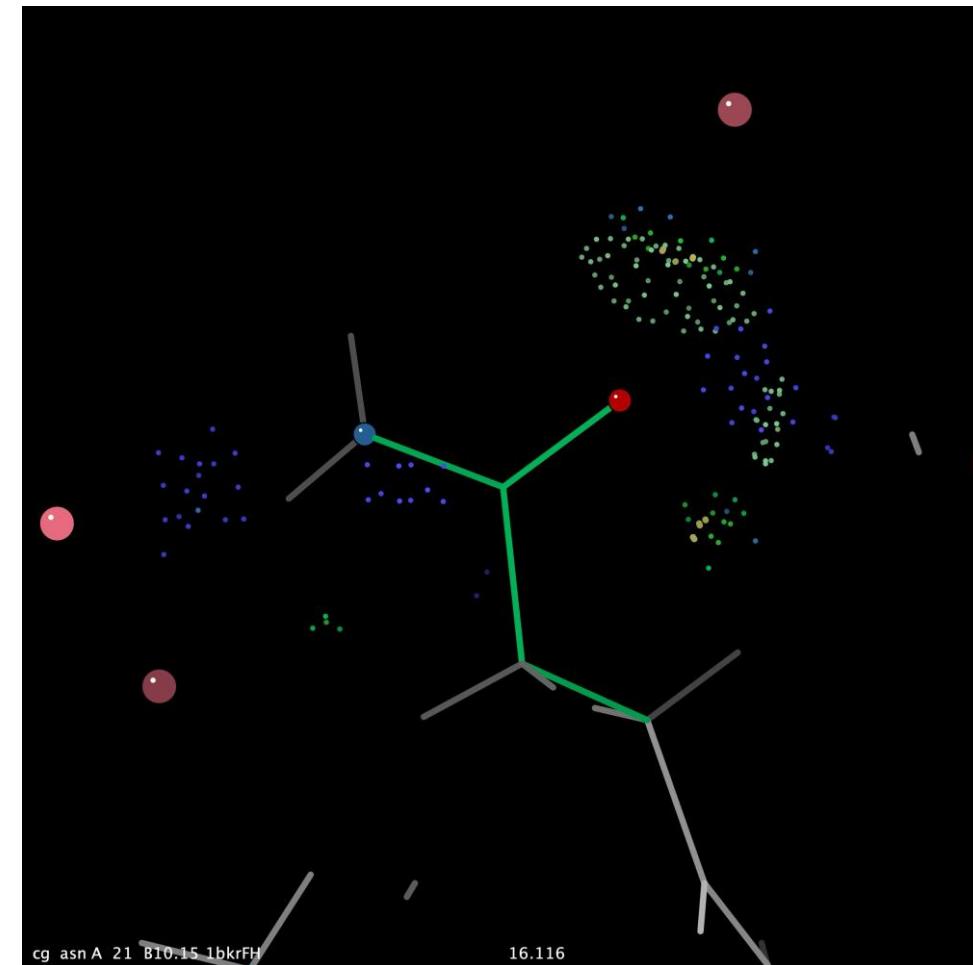
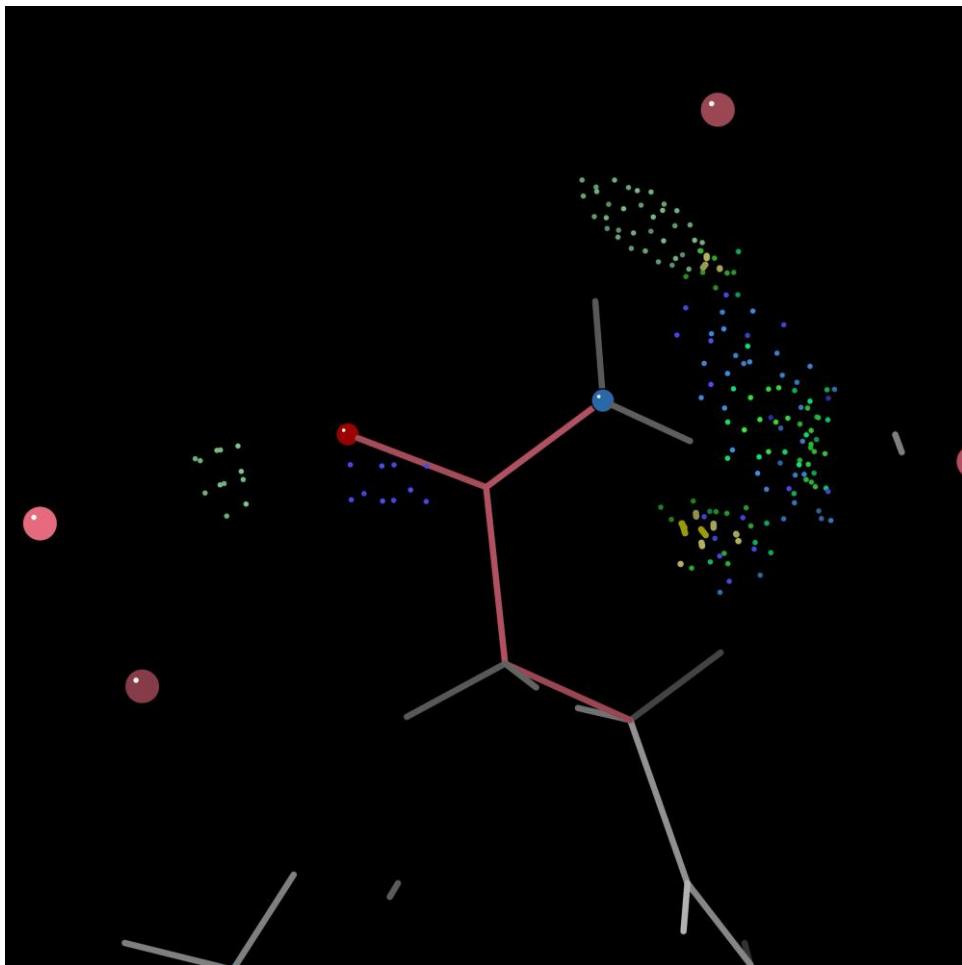
- Clashes usually occur alongside other outliers
- Emphasize modeling errors
  - *Real* rare features are less likely to have clashes
- Can imply direction for fixups

# All-Atom Contacts and Clashes: Asn/Gln/His Flip corrections



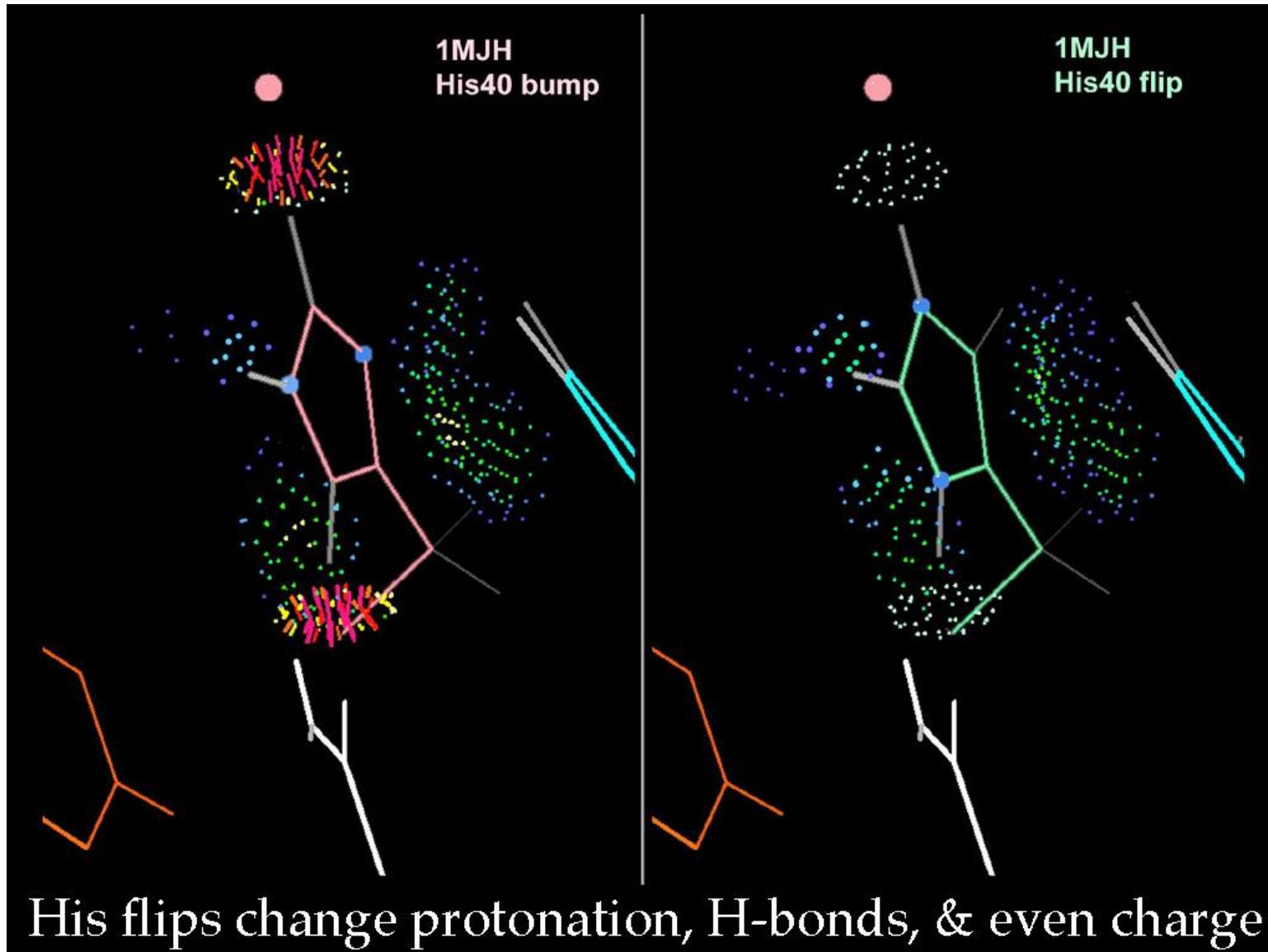
Which Gln is correct?

# All-Atom Contacts and Clashes: Asn/Gln/His Flip corrections



Which Asn is correct?

# All-Atom Contacts and Clashes: Probable causes

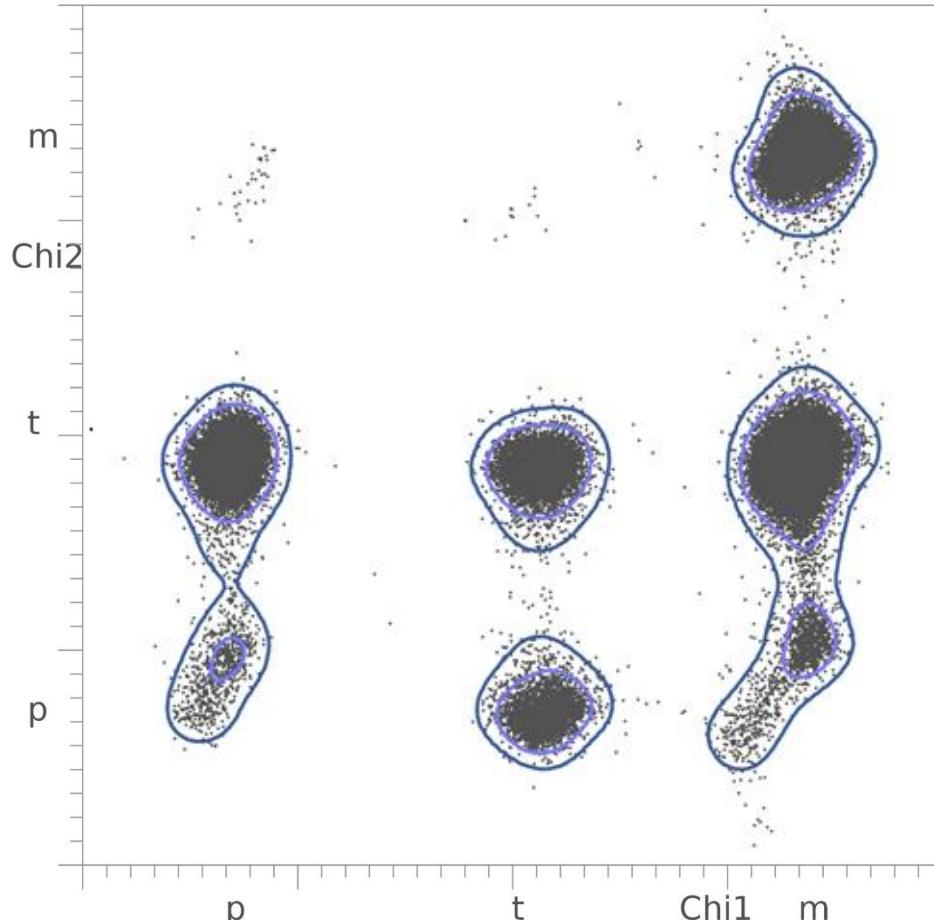


## Sidechain flips

- Asparagine, Glutamine, and Histidine (N/Q/H) are pseudo-symmetric
- Wrong orientation can produce clashes without other error markup
- Fix with Reduce or Coot tools, then re-refine.

# Sidechain Rotamers

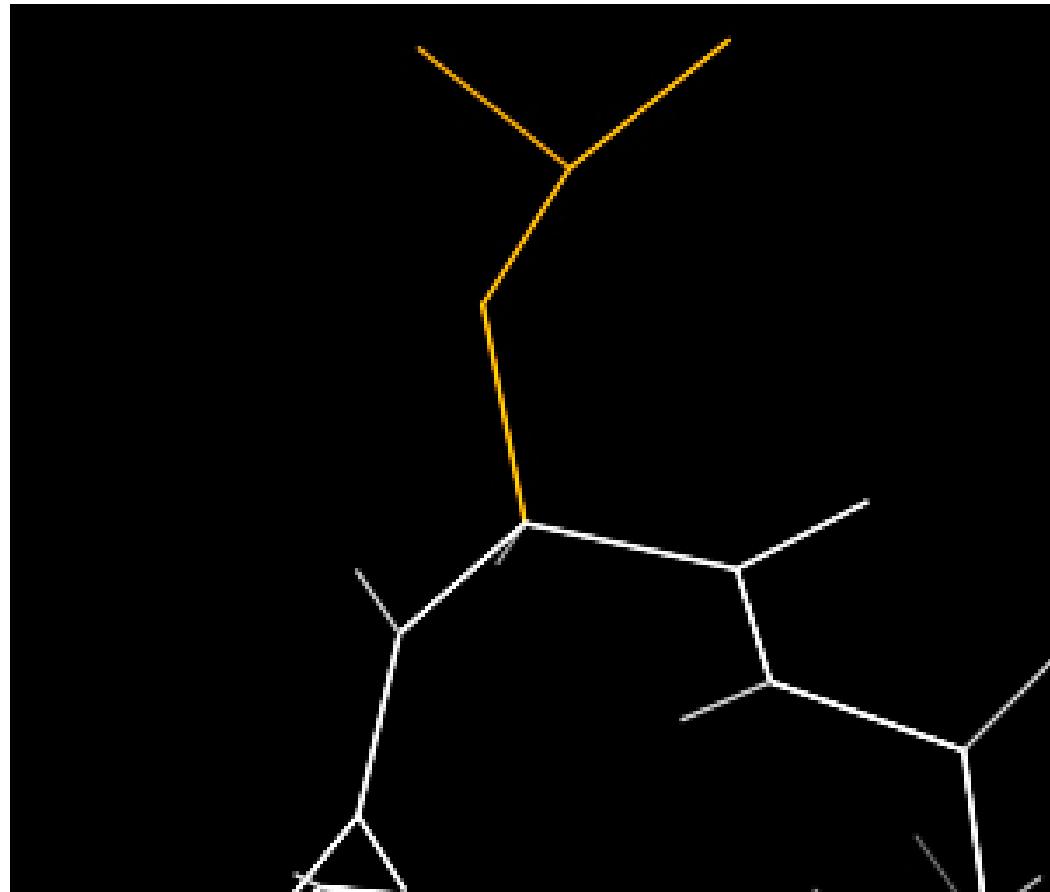
# Sidechain Rotamers: Method



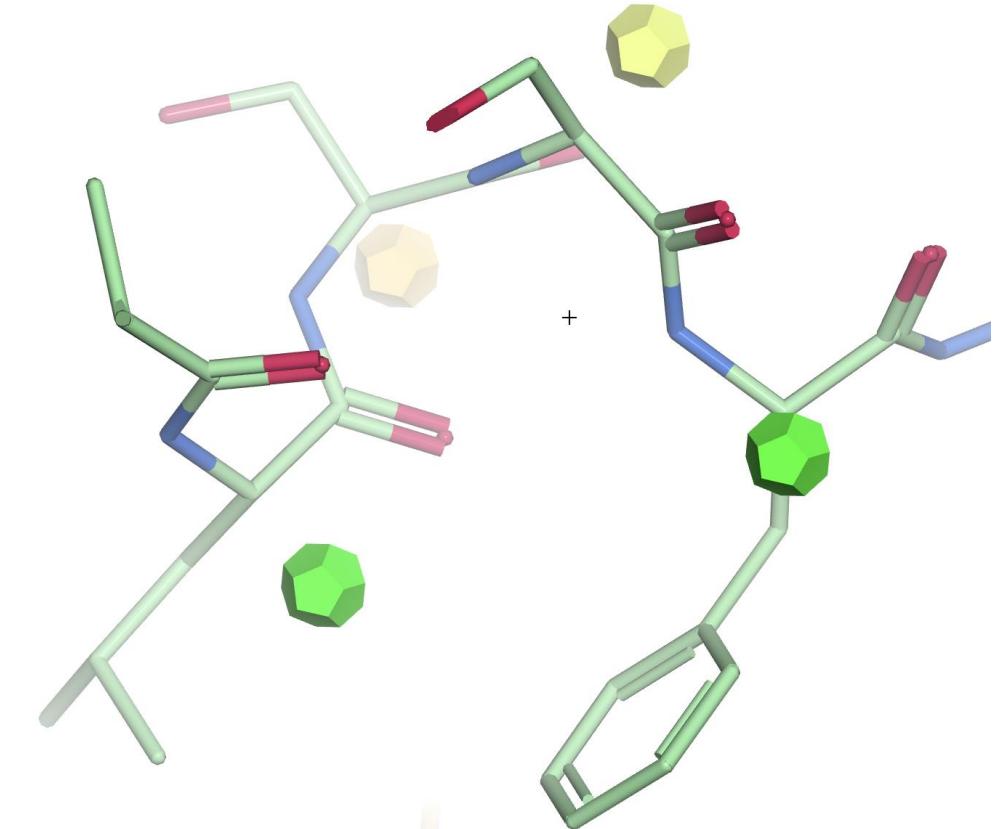
Rotamer distribution for Isoleucine in  $\chi_1/\chi_2$  space

- Sidechain conformations are described by a series of  $\chi$  (Chi) torsions
- Rotamers are statistically expected combinations of  $\chi$  values
- For tetrahedral atom centers, this means staggered
  - p +60°
  - t 180°
  - m -60°
- For planar atom centers, rotamers are much more continuous
  - Rotamers are named with a central value
  - e.g m90 or p-80 for Histidine
- Updated in 2016:
  - Favored (98% of data) Allowed (99.7% of data)

# Sidechain Rotamers: Visualization

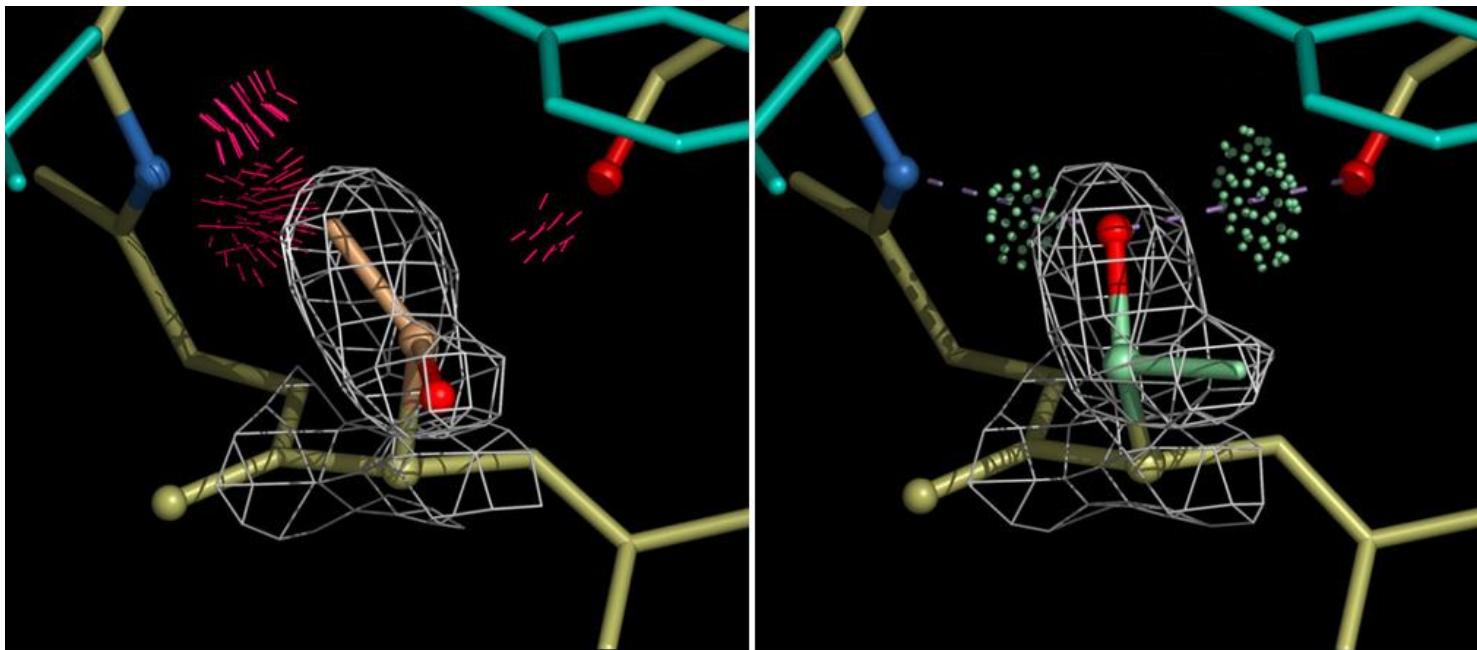


In KiNG, Rotamer outliers are traced in gold over the modeled sidechain



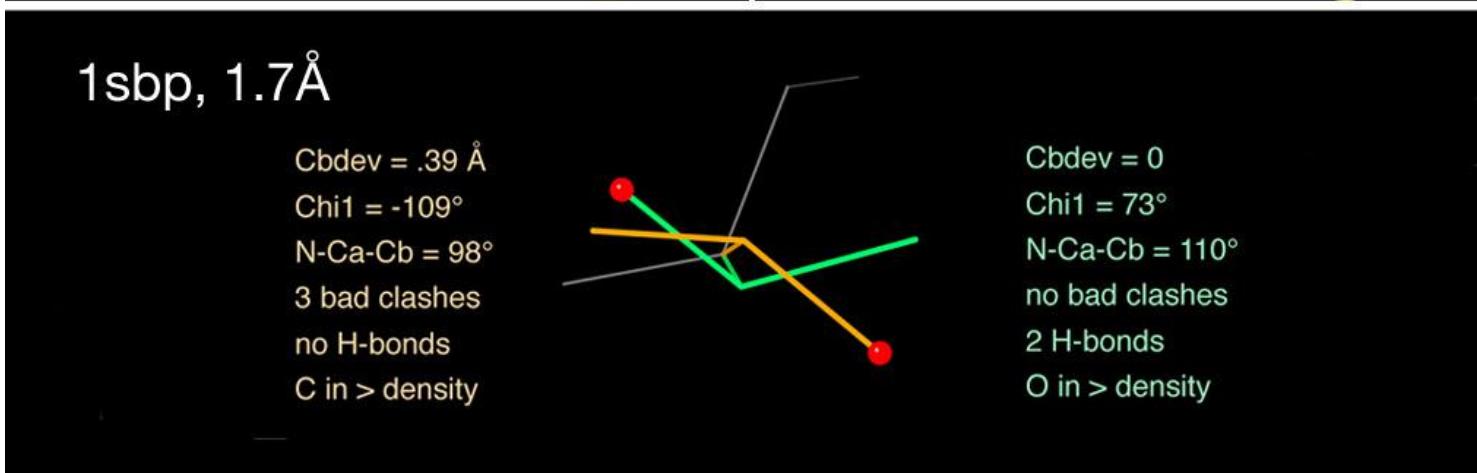
In Coot/Moorhen, Rotamers are marked with a colored dodecahedron

# Sidechain Rotamers: Probable causes

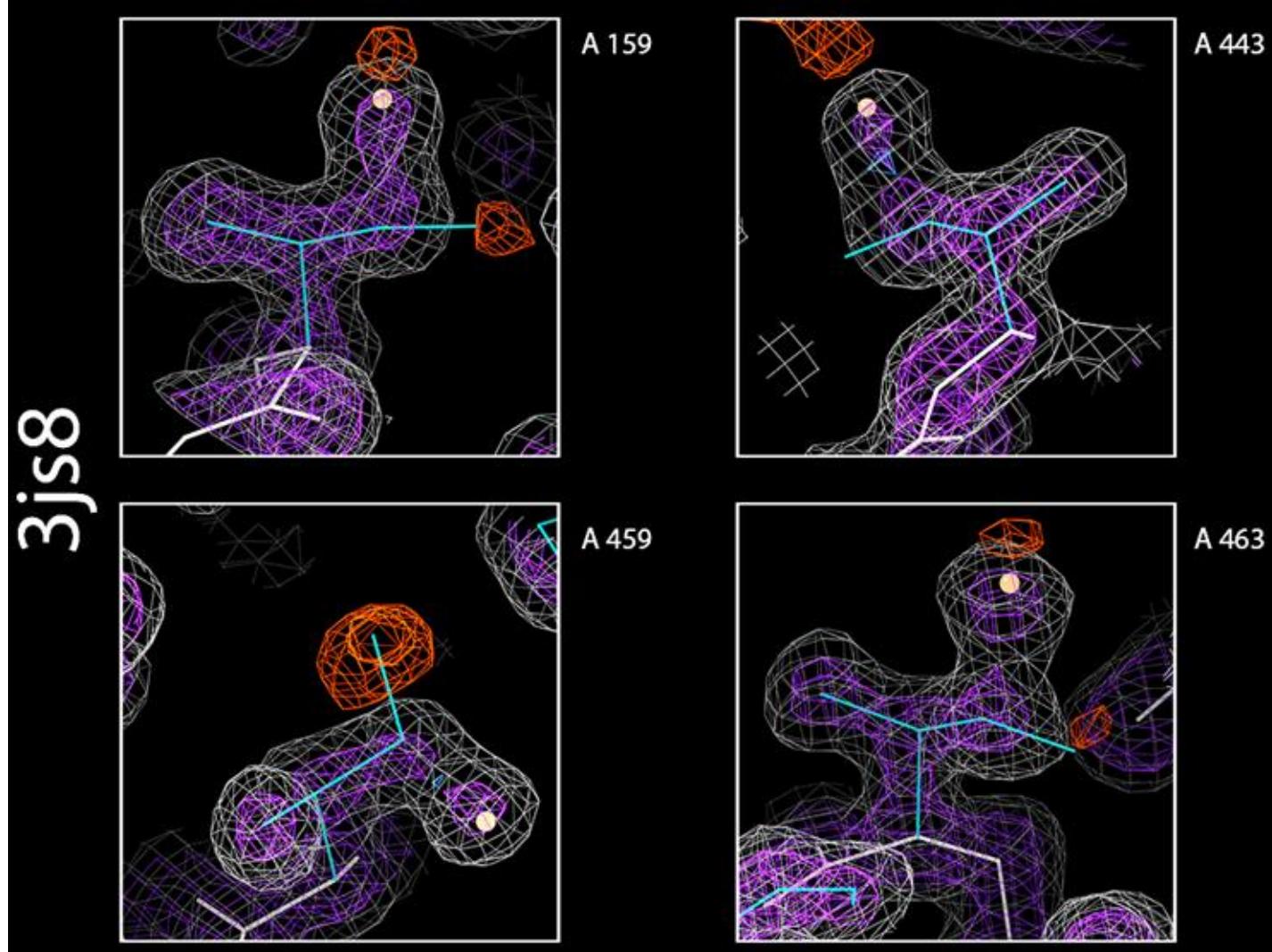


## Backwards Valine, Leucine, Threonine

- May find terminal atoms fit into density at the expense of the branch atom
- Simple to fix with a flip (then re-refinement)



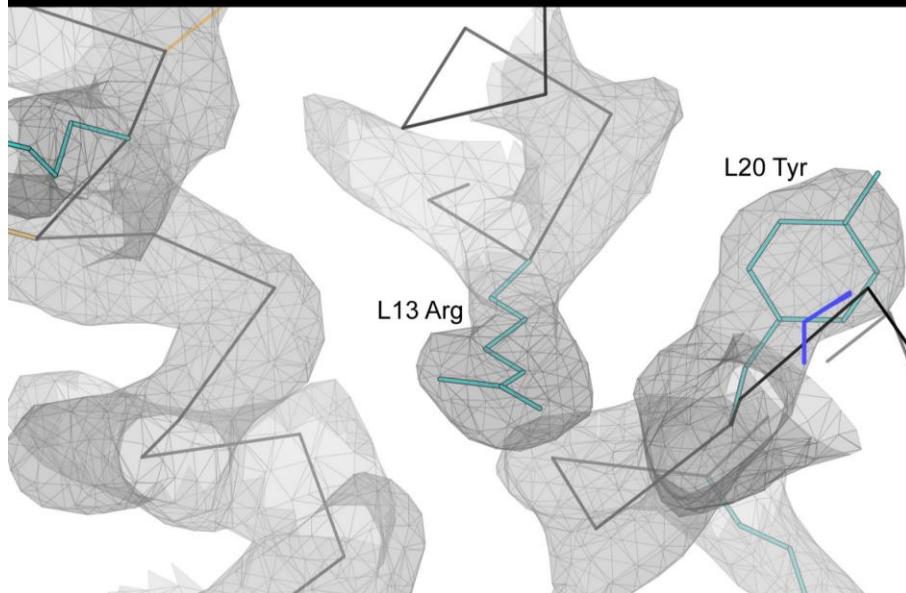
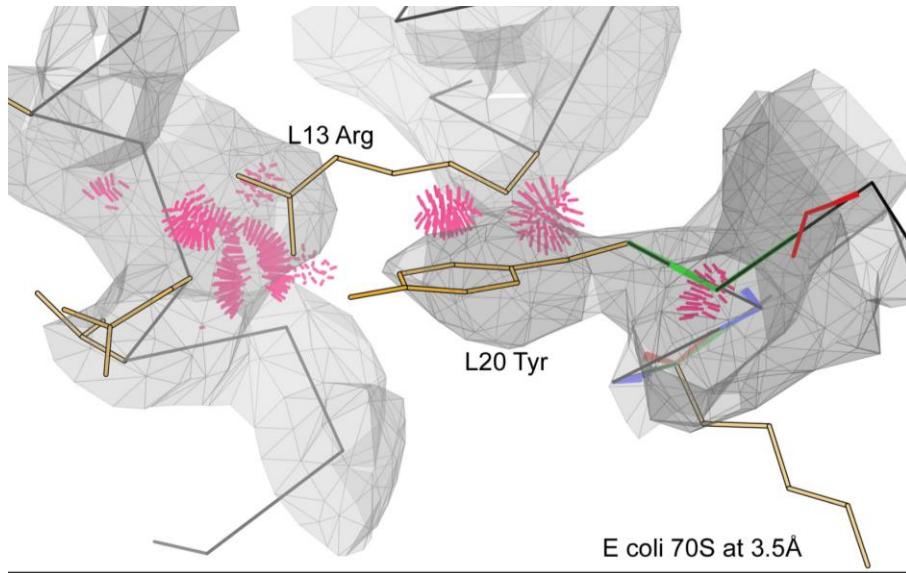
# Sidechain Rotamers: Probable causes



## Water problems

- Modeled water may co-opt sidechain density and create a rotamer outlier
- Isoleucine CD1 is especially vulnerable
- Delete water, rebuild sidechain

# Sidechain Rotamers: Probable causes



## Sidechains in wrong density

- Sidechains can get stuck in the density for other features
  - Other sidechains
  - Ligands
  - Backbone in  $\sim 3\text{\AA}$  maps
- Have to fix the whole network of misplacements

# Protein Backbone Validation

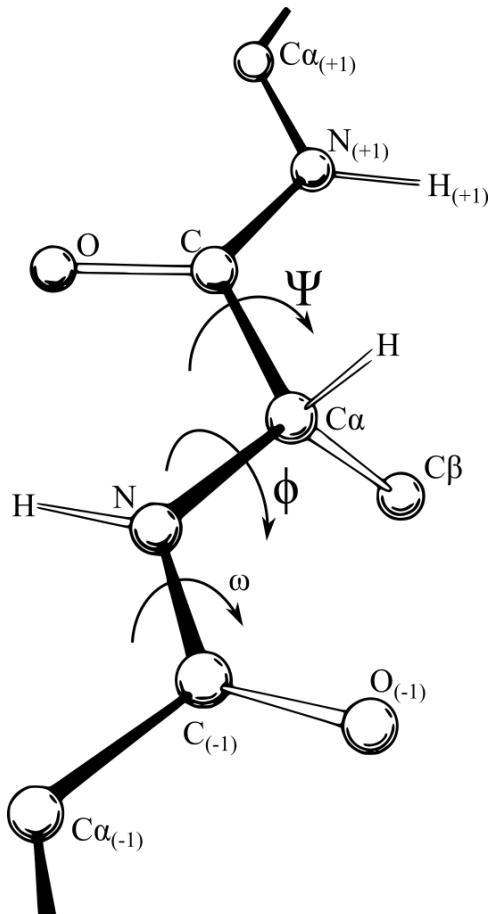
Ramachandran

CaBLAM

Rama-Z

Ramachandran

# Ramachandran: Method



- Phi and Psi torsions describe local protein backbone conformation
- Phi  $\Phi = C_{i-1}-N-CA-C$
- Psi  $\Psi = N-CA-C-N_{i+1}$
- Each residue's  $\Phi/\Psi$  pair is converted into cartesian coordinates and checked against contours of expected behavior

# Ramachandran: Visualization

Ramachandran plots shows location of each residue relative to contours of expected behavior

Different residue categories have very different expectations!

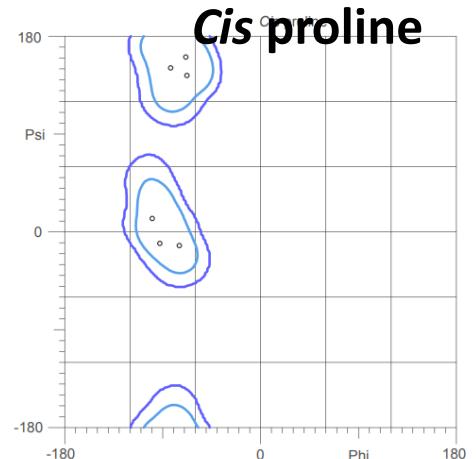
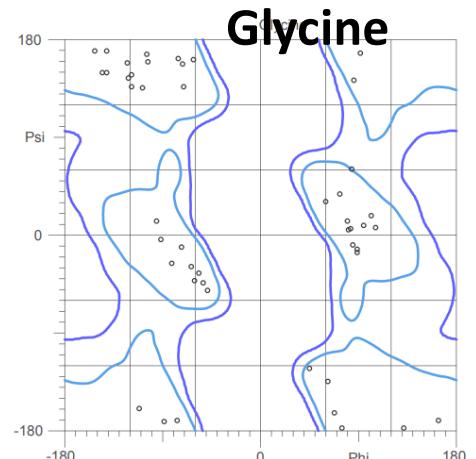
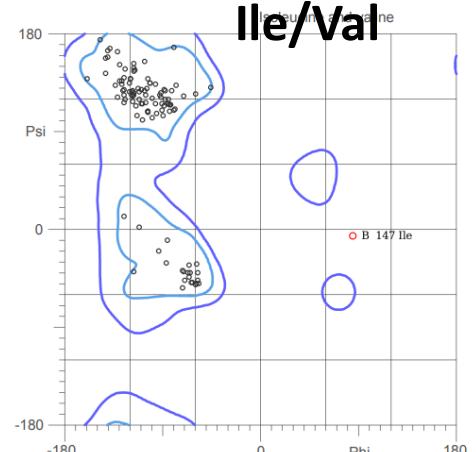
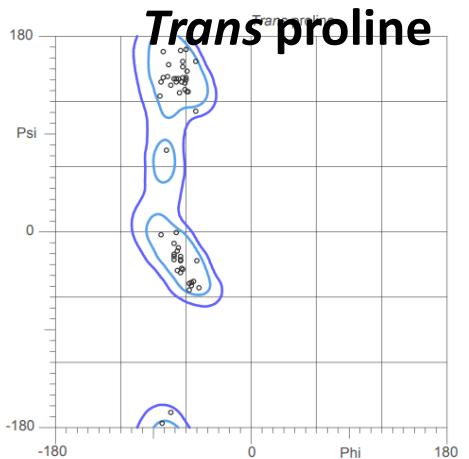
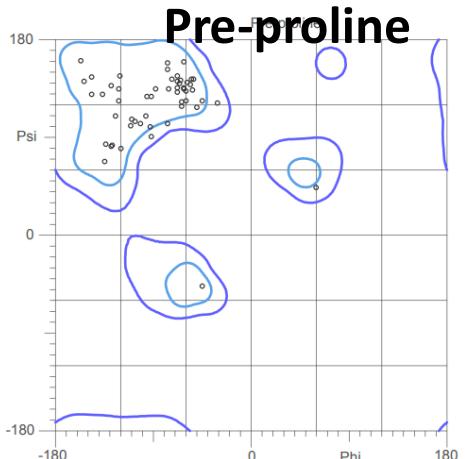
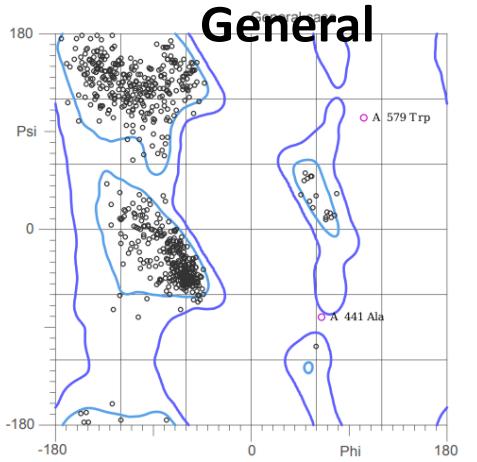
Glycine is permissive and symmetrical

Proline is restrictive

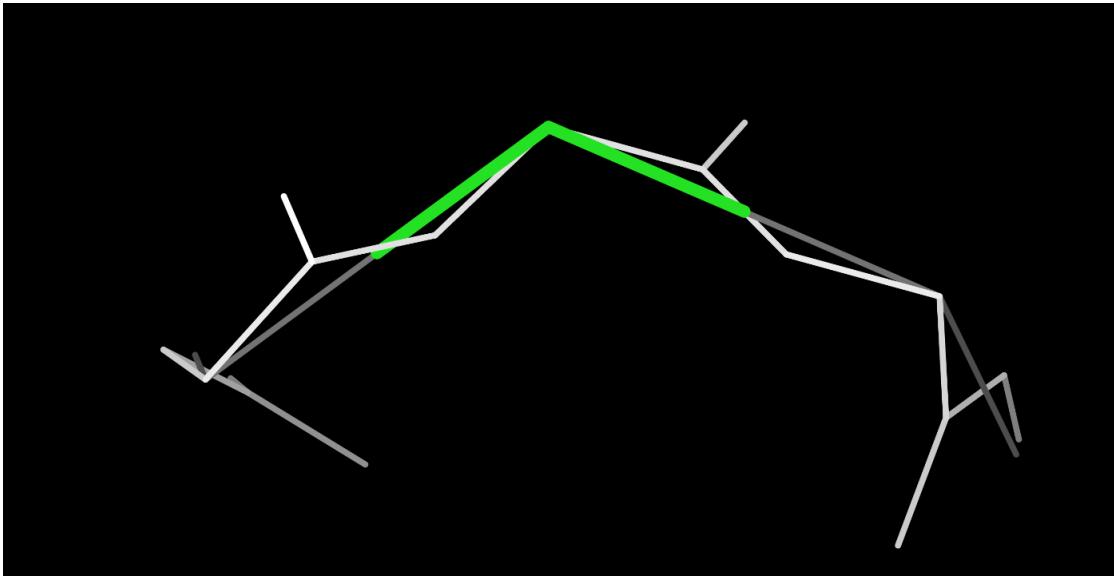
Branched C-Beta sidechain (Ile,Val) affect distribution

Favored (98% of data)

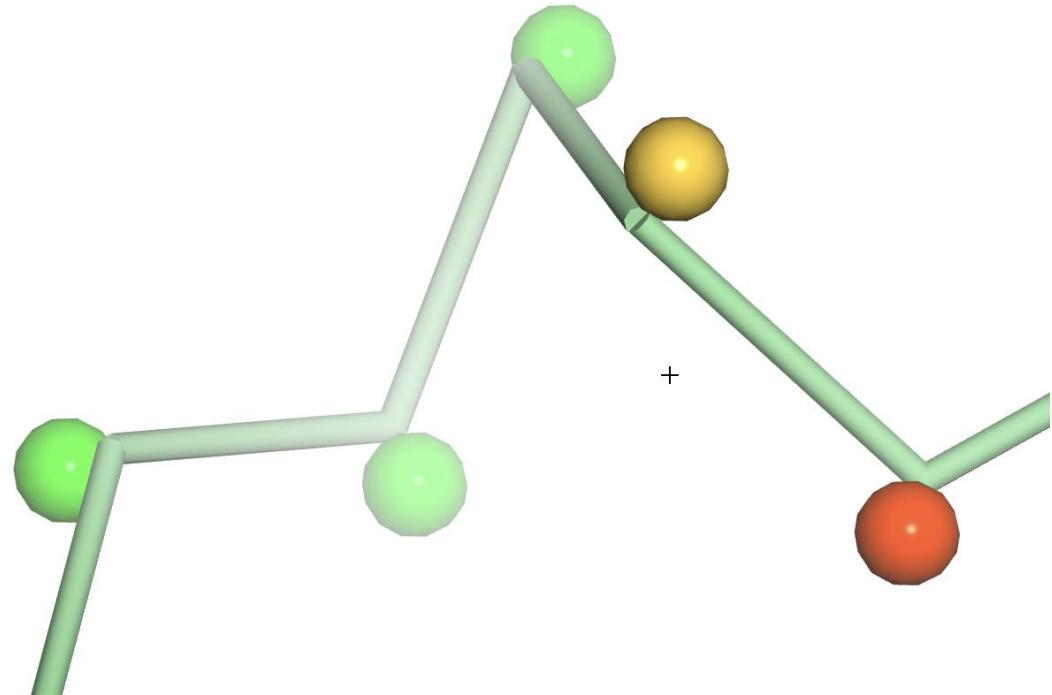
Allowed (99.5% of data)



# Ramachandran: Visualization



KiNG markup highlights an outlier residue's CA in green, and extends to the peptide bonds on either side, along the CA-CA-trace

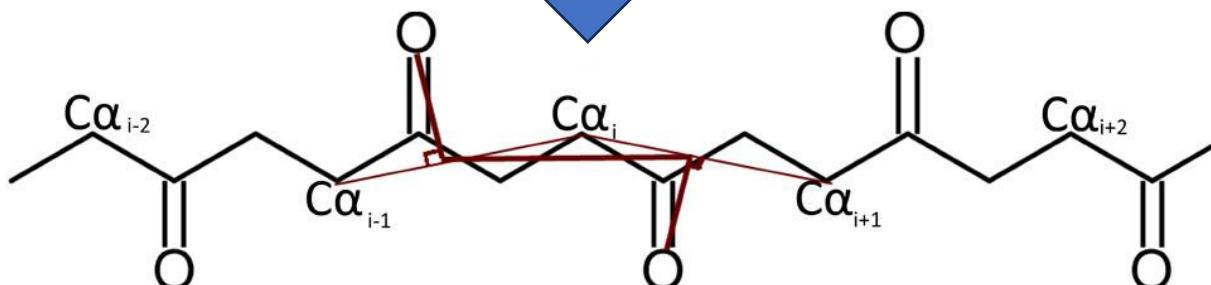
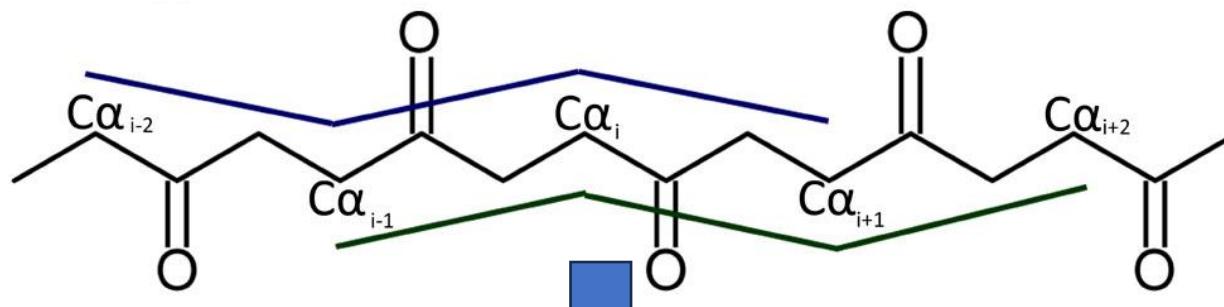


Coot/Moorhen markup places a ball at each CA, color-coded by Ramachandran favorability.

CaBLAM

# CaBLAM: Method

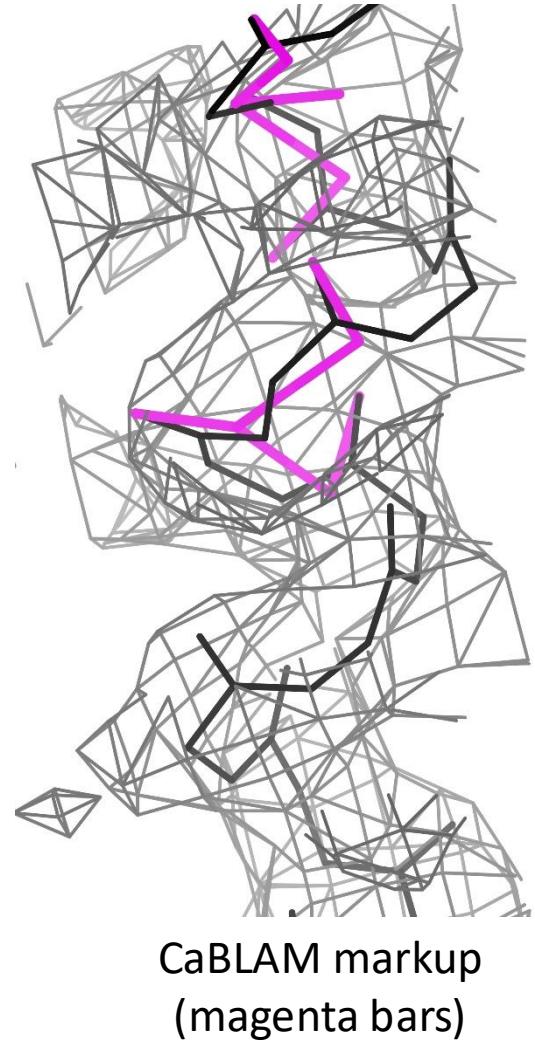
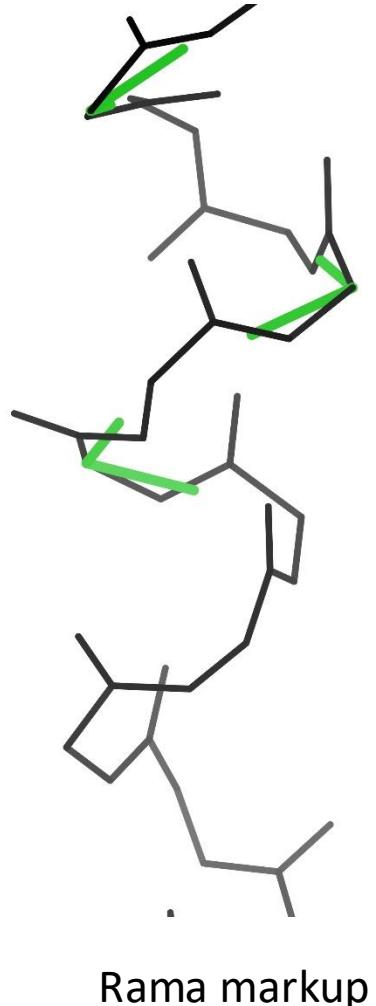
CA-pseudodihedrals capture model “intent”



Peptide-peptide-pseudodihedral captures  
common model errors

- At low resolution, the backbone CA trace is modeled better than the backbone details
- Common model errors involve wrong peptide plane orientation
- CaBLAM uses modeled CA trace geometry to predict likely peptide plane orientation, and marks the discrepancies

# Rama/CaBLAM: Probable causes

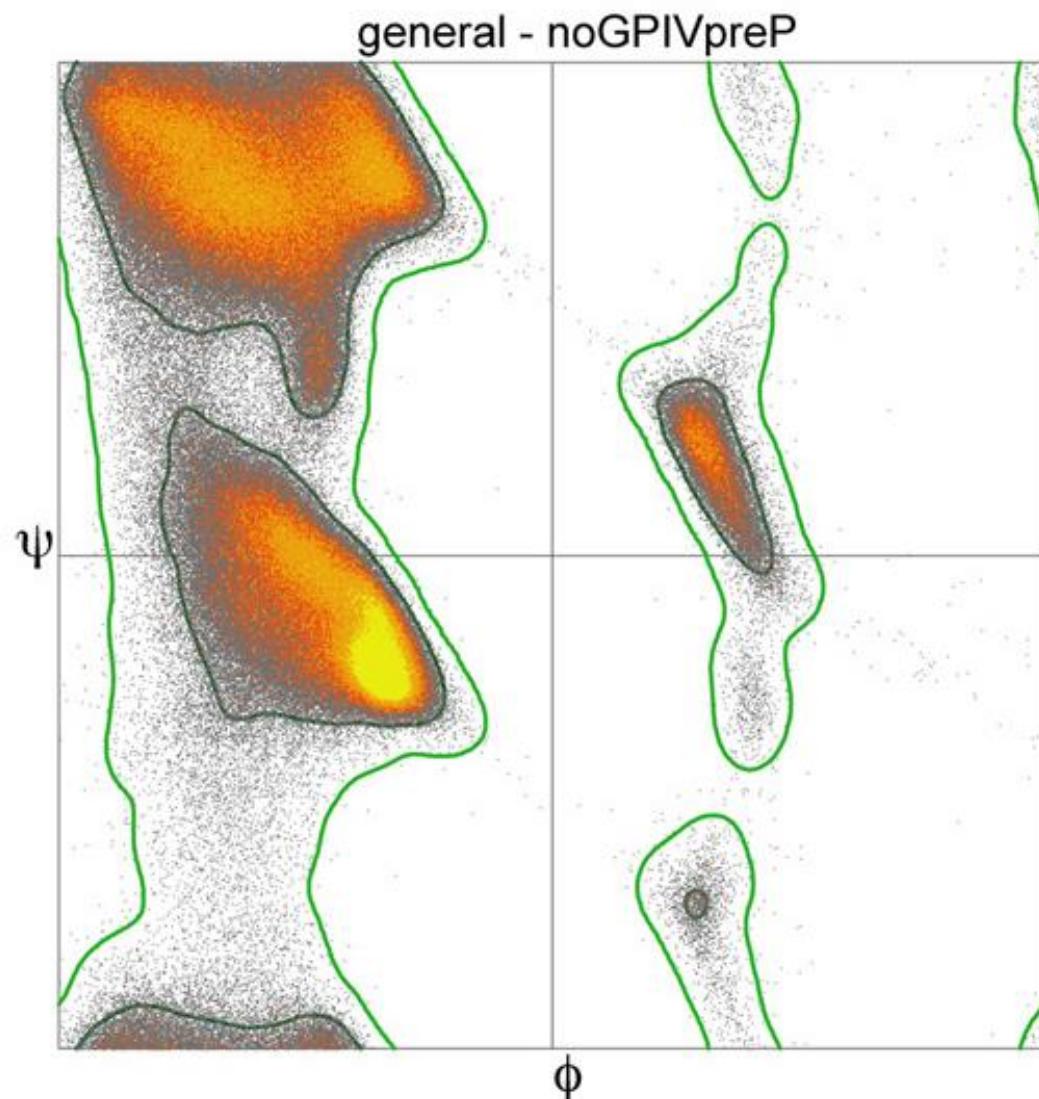


## Misplaced carbonyl oxygens

- At resolutions worse than  $\sim 2.5\text{\AA}$ , carbonyl oxygen density disappears
  - O may be fit in arbitrary orientation
- Low-resolution density envelope allows multiple models
  - Not everything that fits is protein-like
  - Data doesn't have enough information to choose among models

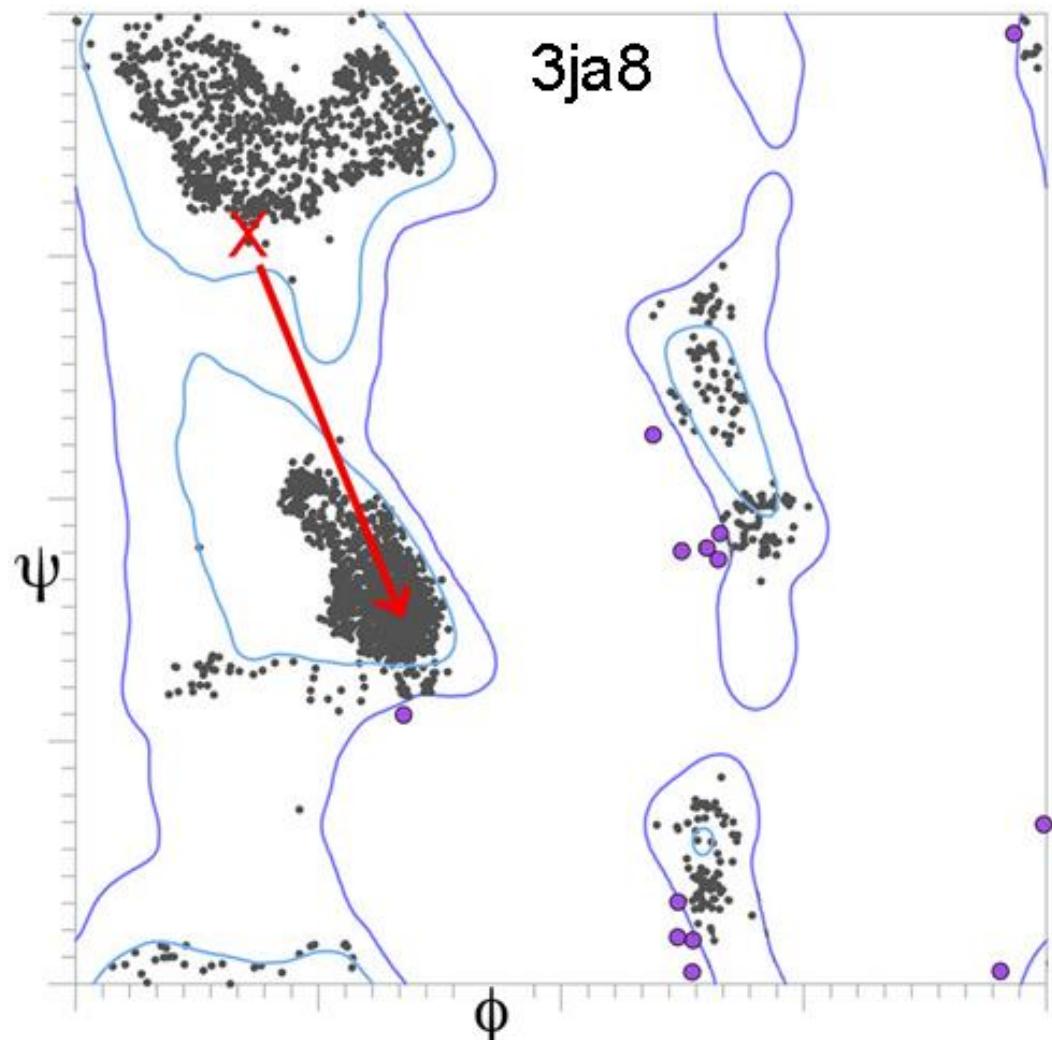
# Ramachandran Z-score

# Ramachandran Z-score: Method



- Compare observed Ramachandran distribution against expected distribution (shown)
- Assign statistical Z-score based on distance from expectation
- $|Z\text{-score}| \leq 2$  indicates a realistic distribution
- $|Z\text{-score}| > 3$  indicates a highly unrealistic distribution

# Ramachandran: Probable causes



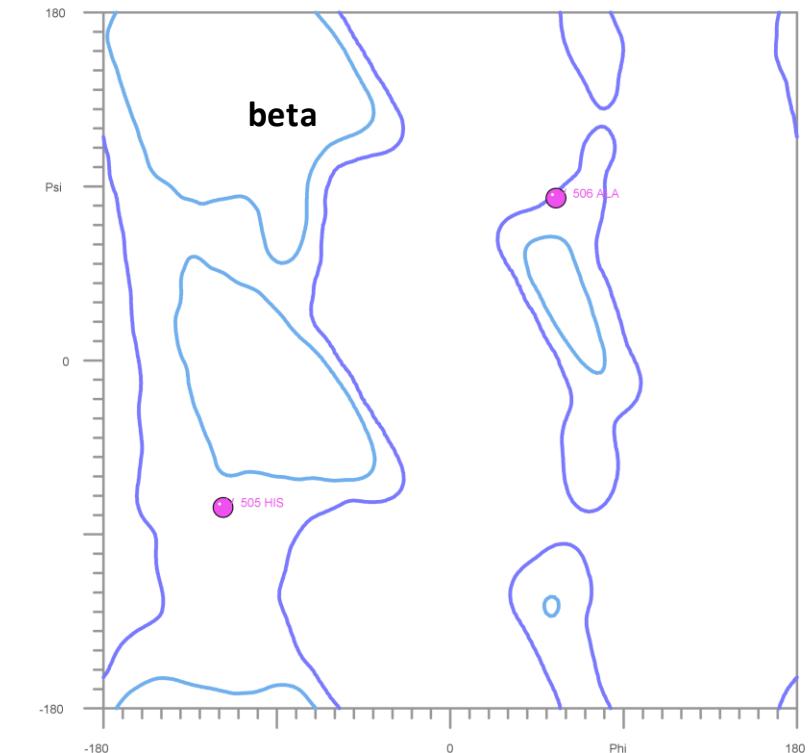
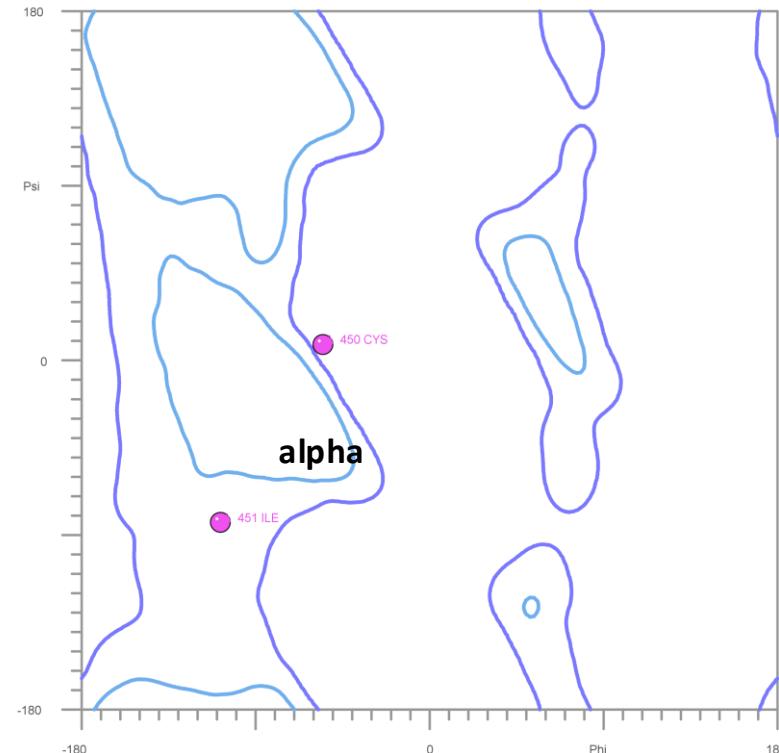
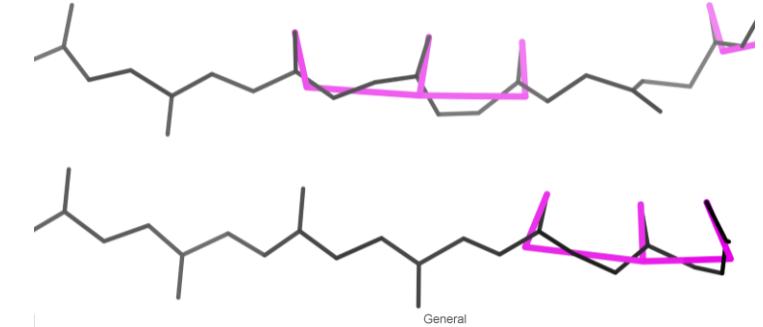
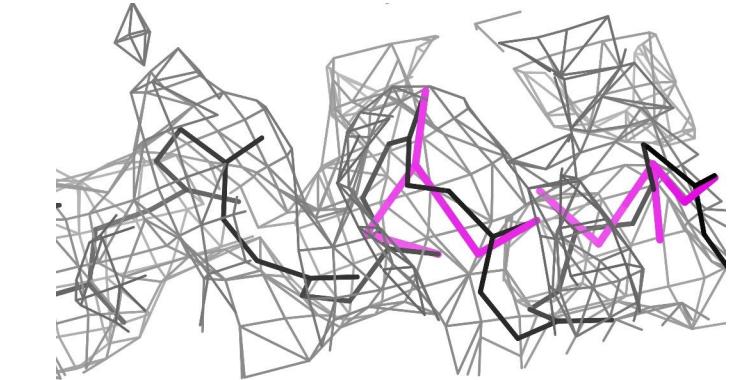
## Overfitting to Rama criteria

- Some programs allow refinement of the Ramachandran plot
  - Hides/compounds rather than fixes errors, if used carelessly
  - Artificially improves Ramachandran and MolProbit scores
- Over-idealized distribution may be detectible by Rama Z-Score
  - Use other methods to fix model errors
  - Then (maybe) Rama restraints to hold good structure in place

# Rama/CaBLAM: Probable causes

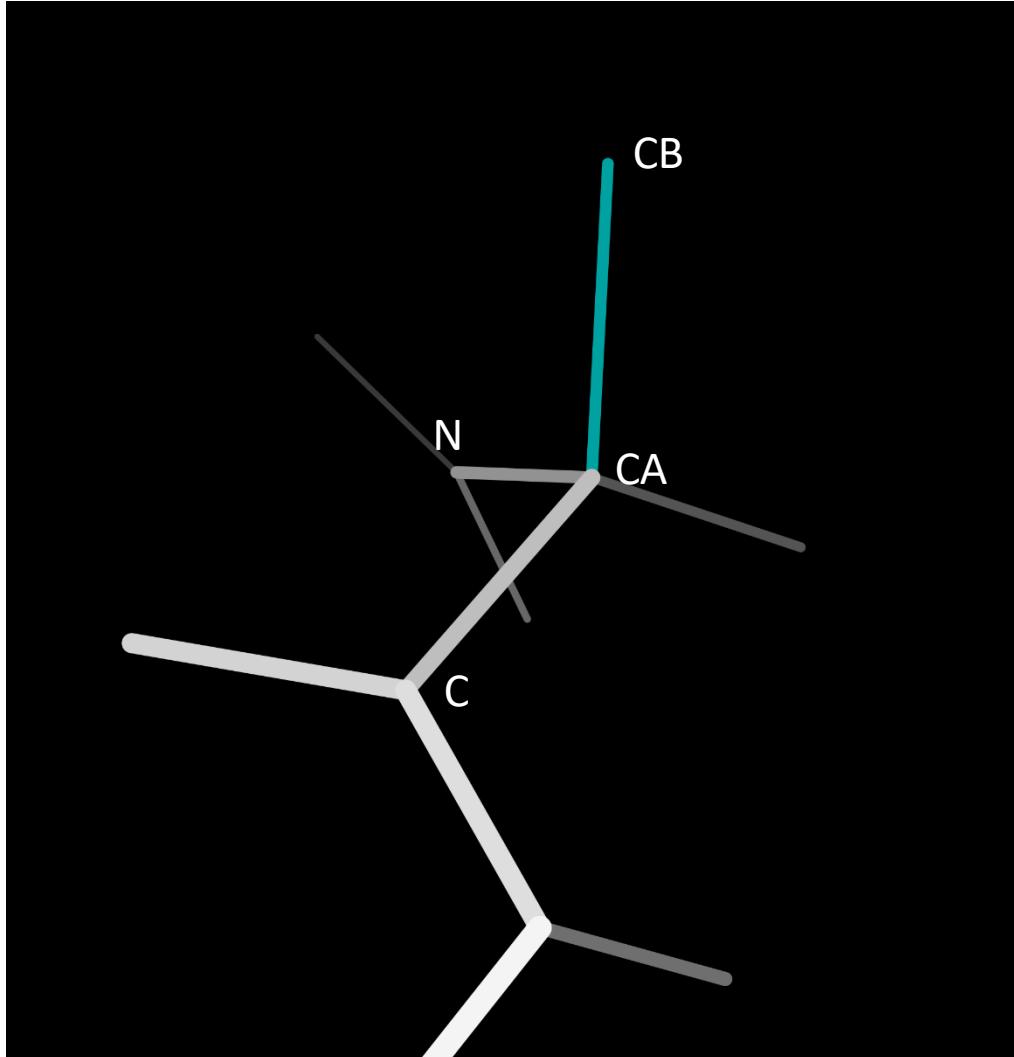
Current Rama position  
does not predict  
*Correct* Rama position

- If model errors are large, points in Rama space are displaced far from their intended regions
- 90° or even 180° peptide orientation errors are possible in low-resolution maps!



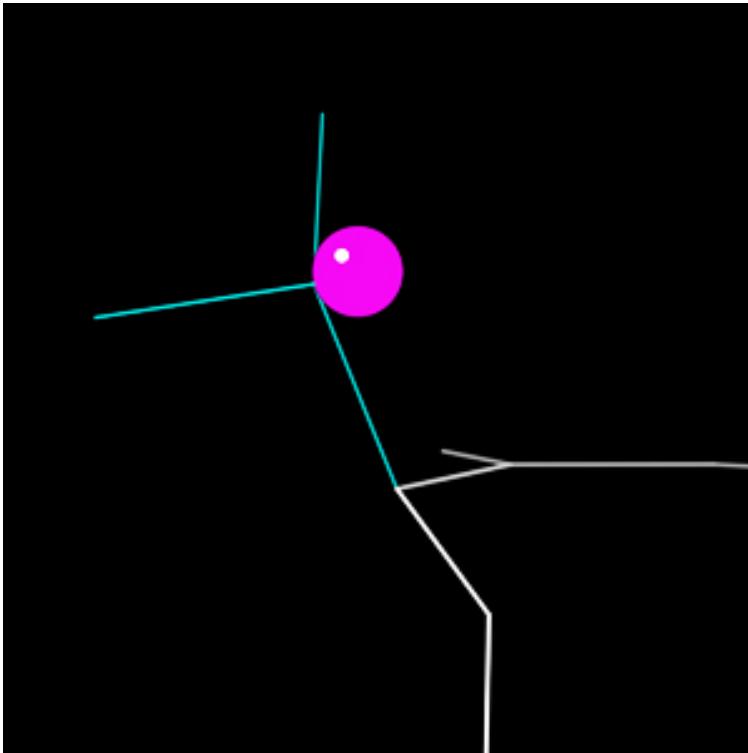
# C-Beta Deviation

# C-Beta Deviation: Method

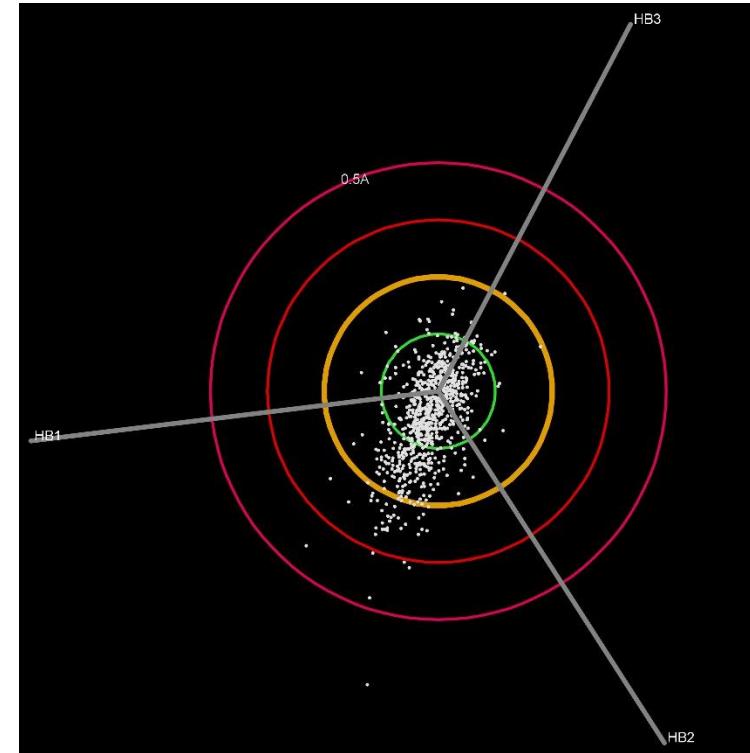


- Ideal CB position is defined by backbone geometry
- Calculate ideal position using average of two torsions
  - N-C-CA-CB
  - C-N-CA-CB
- CBs modeled  $>0.25\text{\AA}$  from ideal position are outliers

# C-Beta Deviation: Visualization



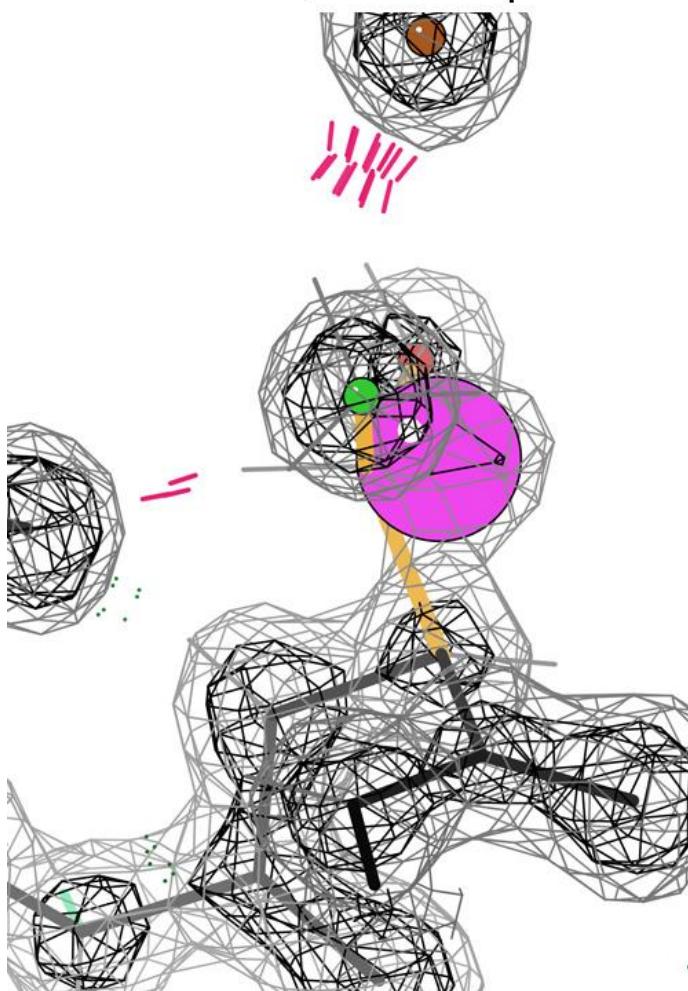
- In KiNG, a magenta sphere is drawn
  - Center at ideal CB position
  - Edge tangent to modeled position
  - Size of sphere proportional to severity of outlier



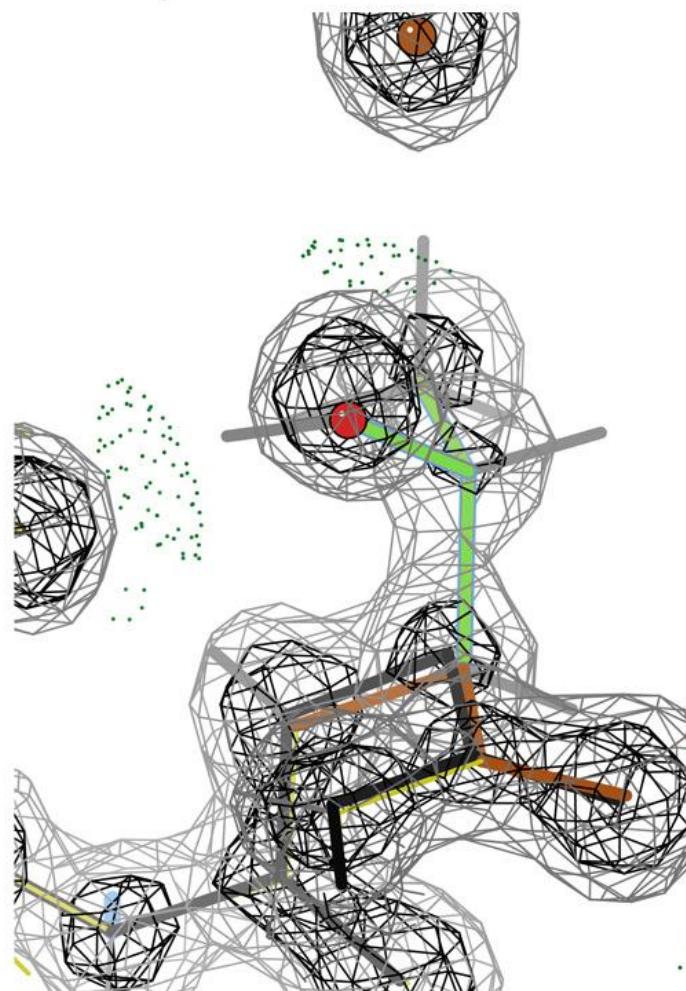
- Bullseye kinemage shows distribution and direction of all CB positions.
- Yellow circle is 0.25 Å outlier cutoff

# C-Beta Deviation: Probable causes

1bkr Thr101, 0.63Å C $\beta$ dev



refit, clashes now H-bonds



## Misplaced sidechains

- CB deviation outliers are usually caused by misplaced sidechains
  - Especially branched sidechains fit backwards, like this Thr

## Chirality errors

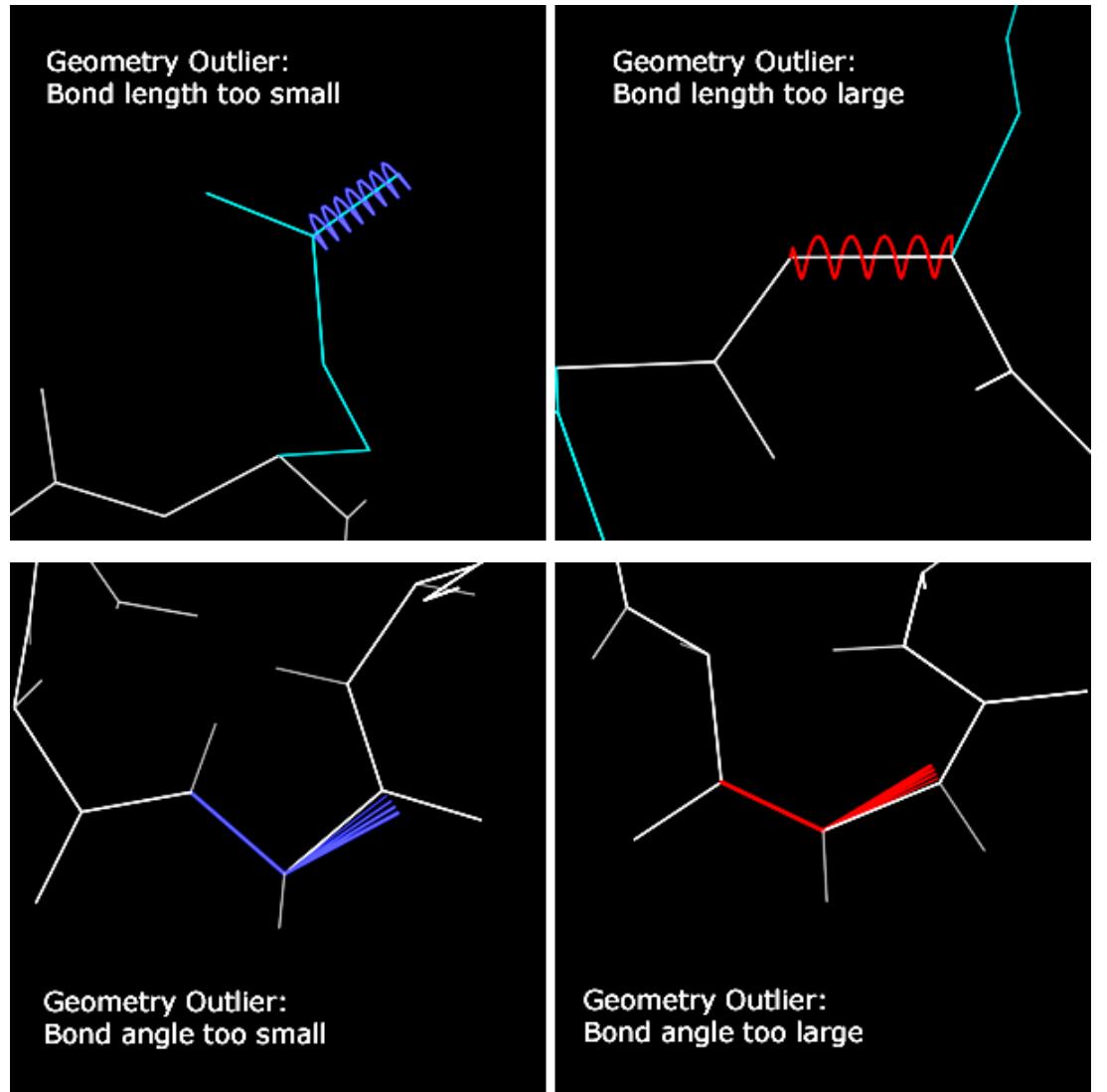
- If D amino acids are misnamed as L amino acids (e.g. ALA for DAL), or vice versa, very large Cbdevs result

# Covalent Bond Geometry

# Bond Geometry: Method

- Measure bond lengths and angles
- Check against a library of expected values
  - $>4\sigma$  deviation from expected = outlier
- Standard reference library has 1 value per bond or angle
- Derived from Engh and Huber
  - <https://doi.org/10.1107/S0108767391001071>
- Conformation-Dependent Library (CDL) has values that depend on local Ramachandran conformation
- Phenix default
- Derived from Karplus et al.
  - <https://doi.org/10.1107/S2059798315022408>

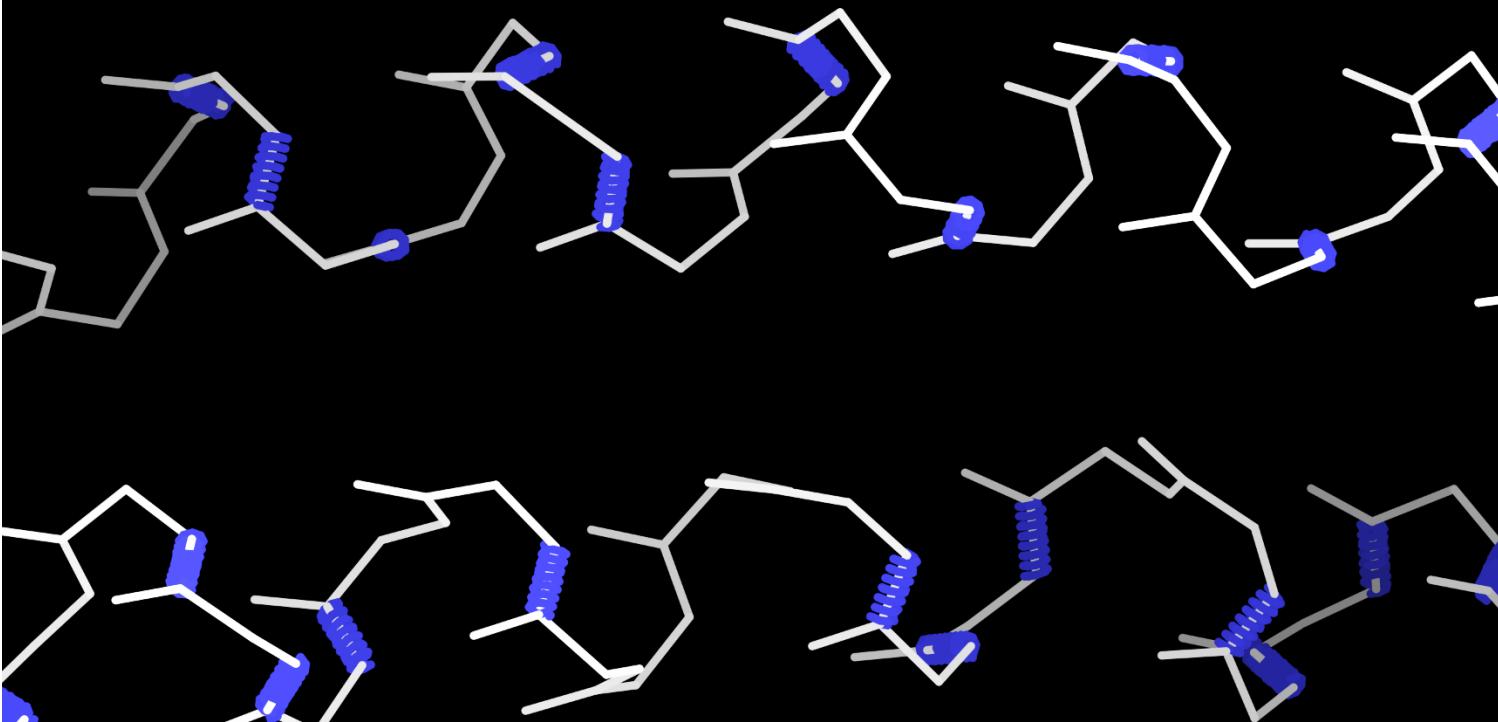
# Bond Geometry: Visualization



- Bond length outliers are drawn as springs
- Bond angle outliers are drawn as fans
- Color-coded
  - Red-shift = too far
  - Blue-shift = too close

# Bond Geometry: Probable causes

C-N peptide bond distances are systematically shortened

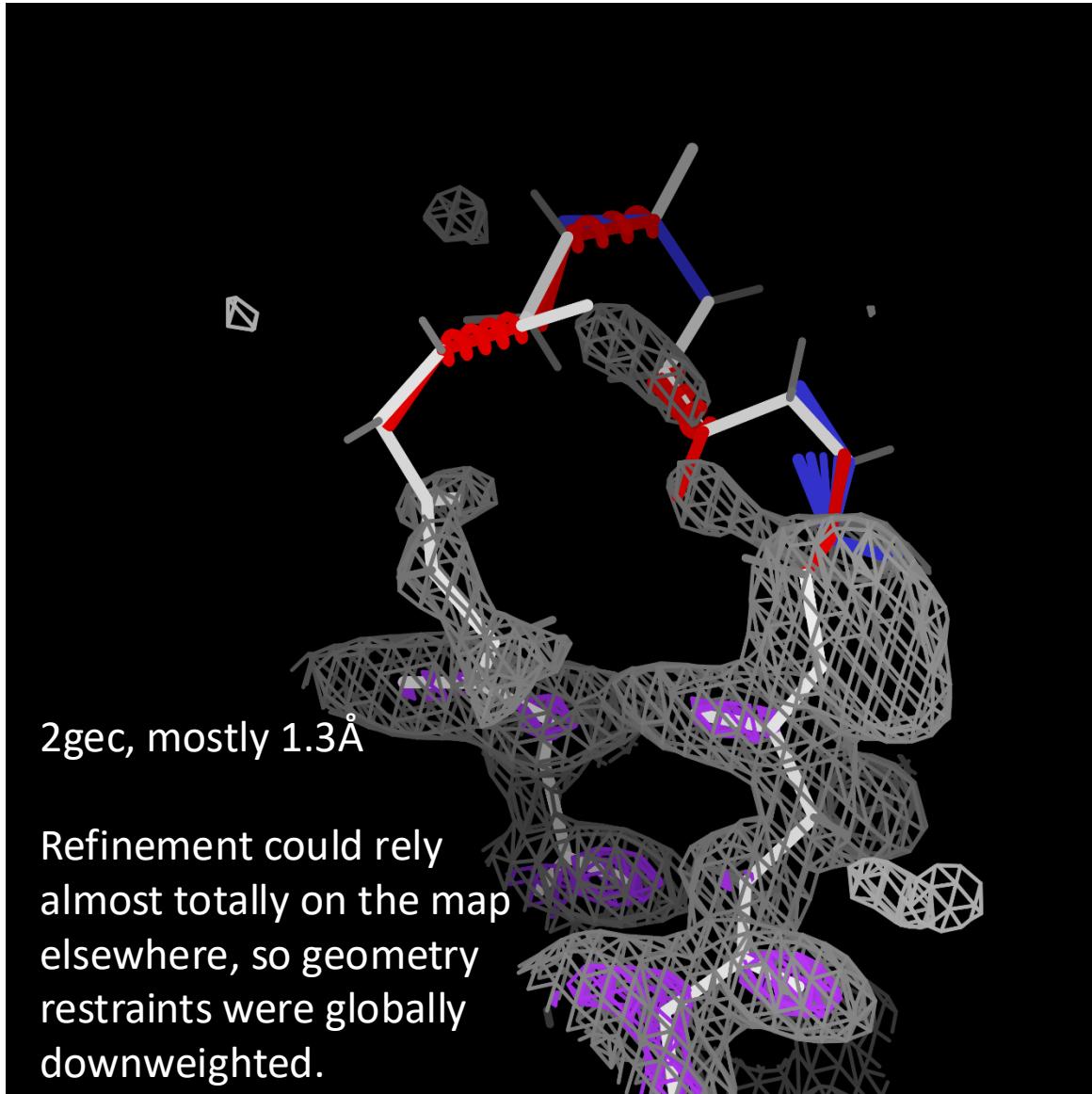


OmegaFold prediction for p81313, as of Sept 2022

## Systematic

- Systematic geometry errors occur in programs with different libraries or expectations
- Be aware of what you import
- Do geometry minimization and/or re-refine.

# Bond Geometry: Probable causes

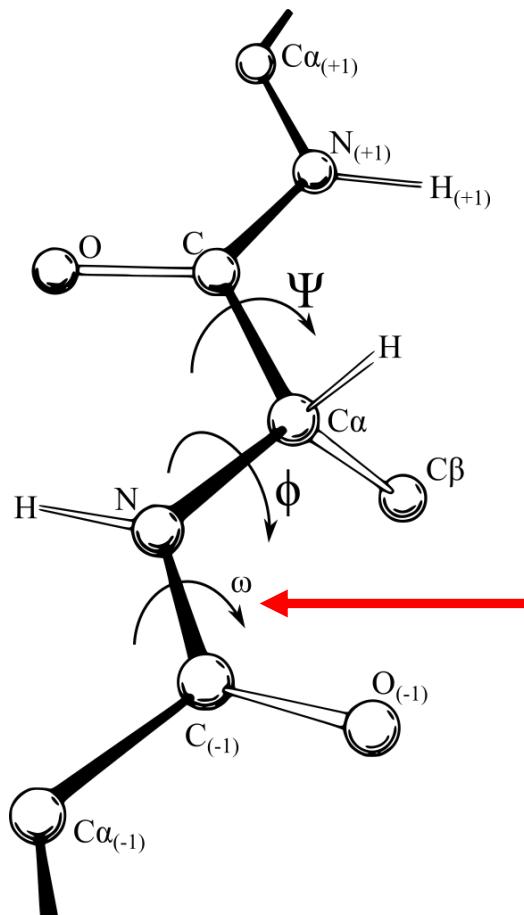


## Localized

- Localized geometry outliers result from conformational strain and/or missing density
- Fix the source of strain
- Manually apply more restraints to low-data regions
- Leave it unmodeled if a good solution is impossible

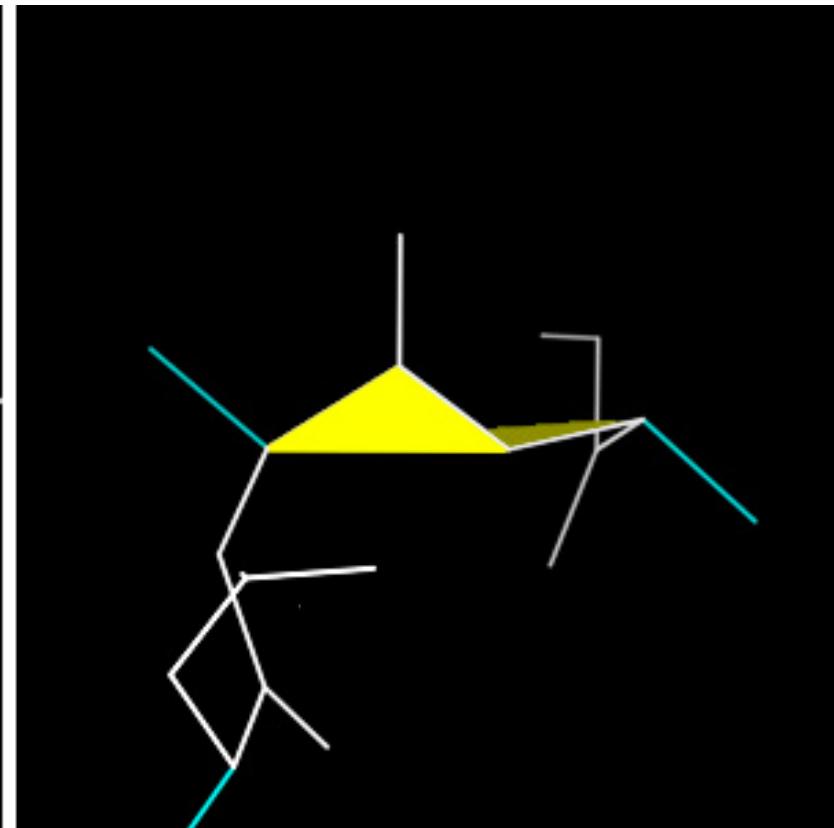
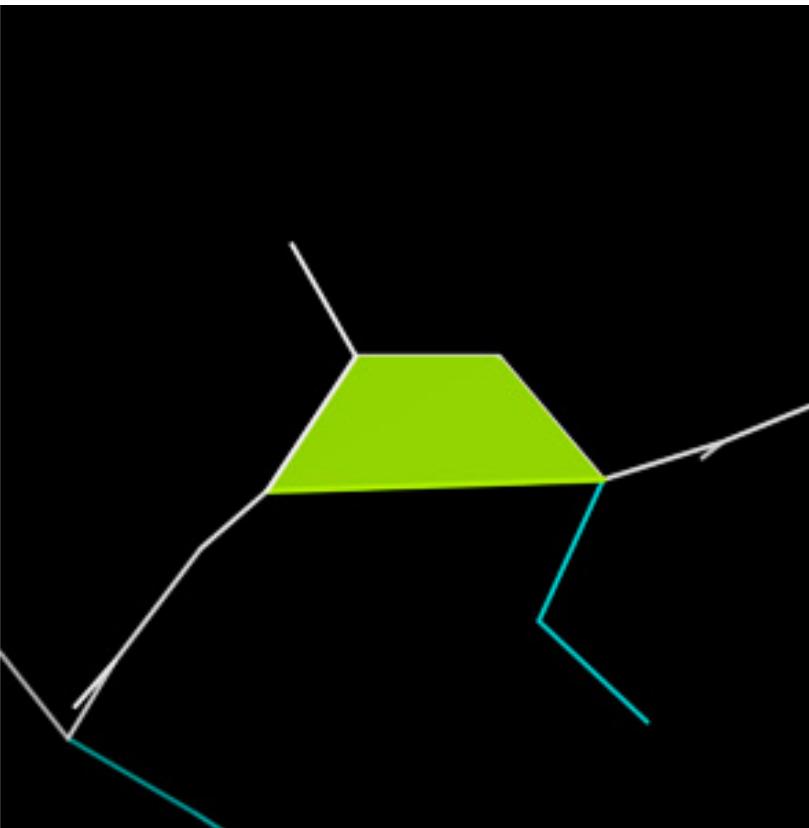
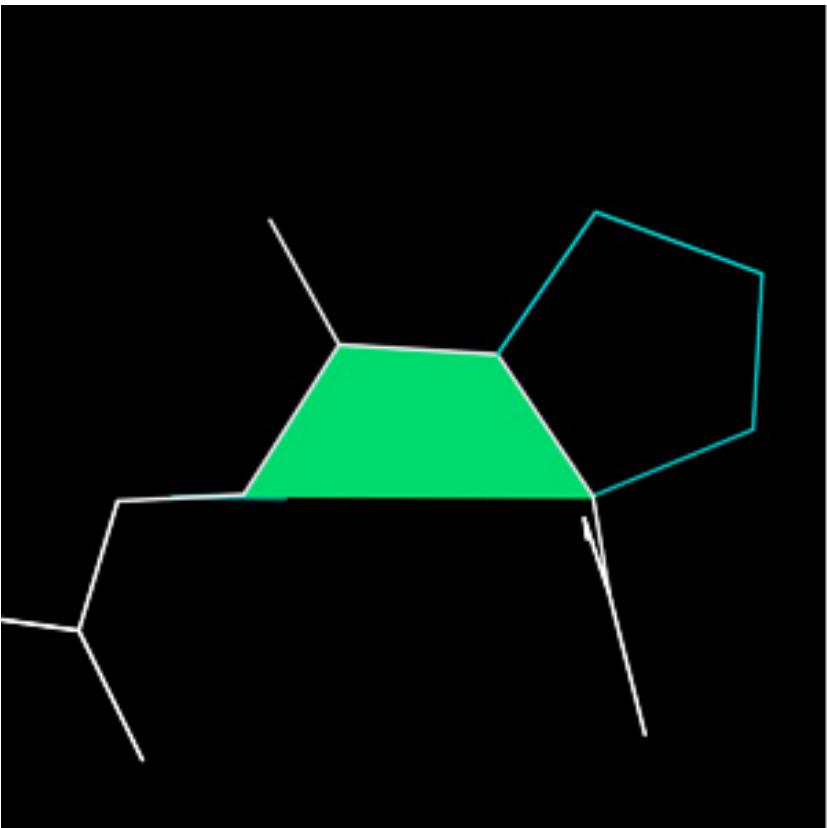
*Cis* Peptides

# Cis Peptides: Method



- The peptide bond that joins amino acids has partial double bond character and does not rotate freely
- CA-C-N-CA torsion
  - “Omega”
- Usually *trans* (CA on opposite sides)
- Rarely *cis* (both CA on same side)

# Cis Peptides: Visualization (KiNG)

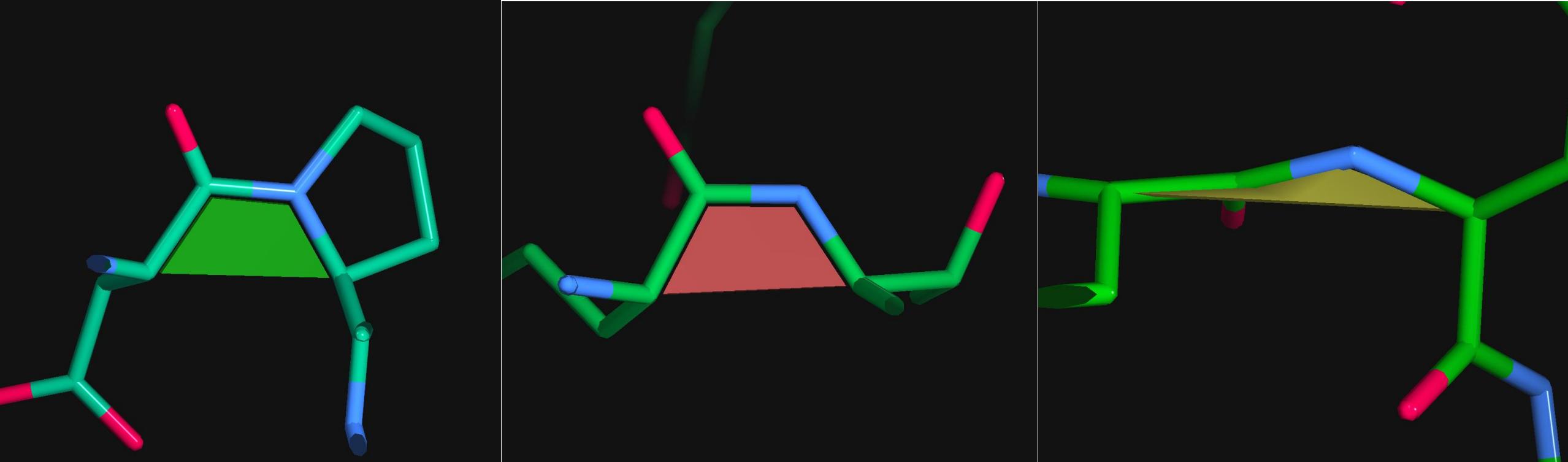


- *Cis* peptide bond is much more common preceding Proline
  - ~5% of **Proline**
- Gentle green trapezoid fills the characteristic CA-CA space

- *Cis* peptide bond is extremely rare preceding other residues
  - ~0.03% of **non-Proline**
- Unpleasantly lime trapezoid fills the characteristic CA-CA space

- Peptides **twisted** >30° from planar are severe geometry distortions
- Space is filled with yellow, angle between component planes approximates severity

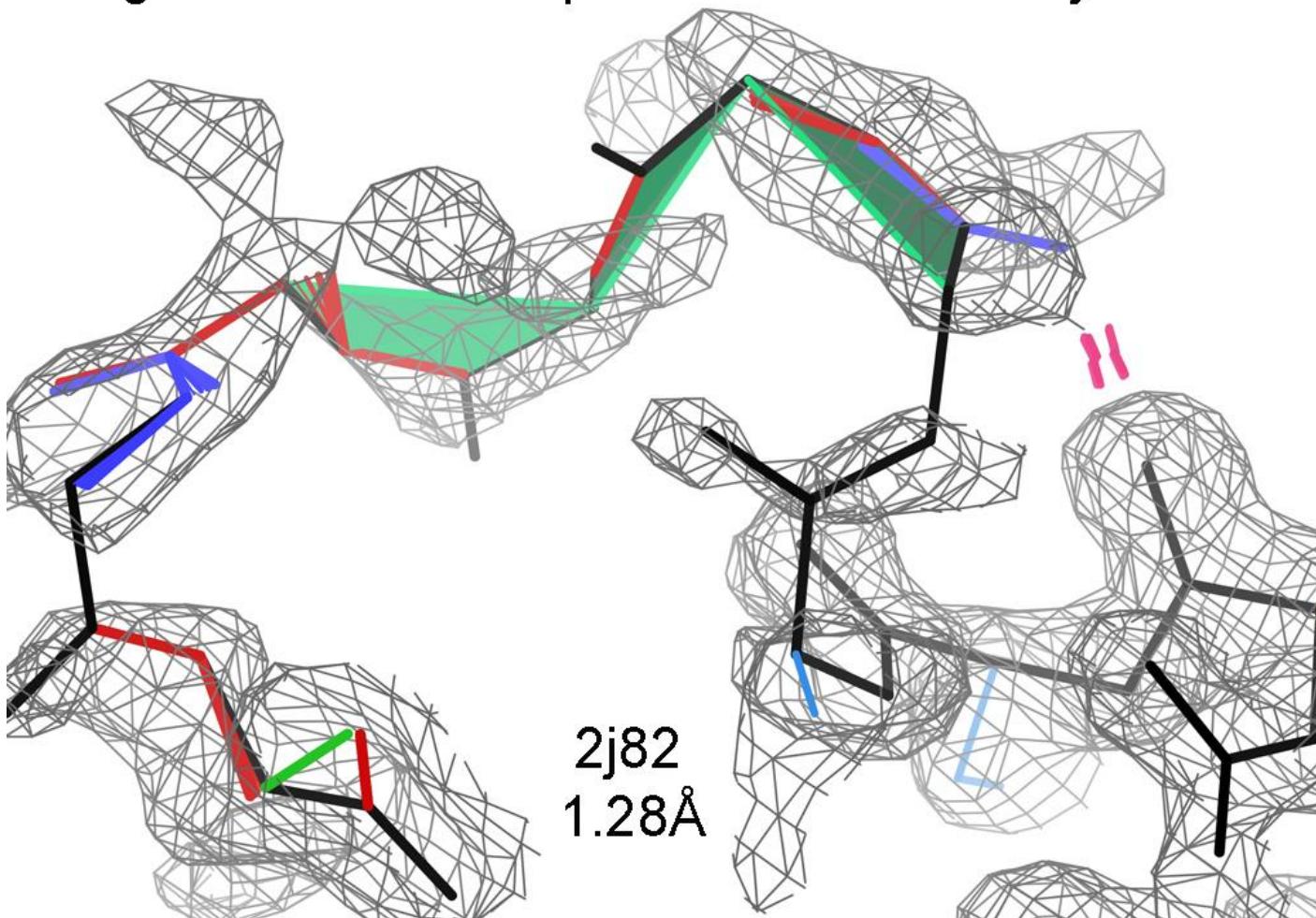
# Cis Peptides: Visualization (Coot)



- *Cis* peptide bond is much more common preceding Proline
  - ~5% of **Proline**
- Gentle green trapezoid fills the characteristic CA-CA space
- *Cis* peptide bond is extremely rare preceding other residues
  - ~0.03% of **non-Proline**
- Warning red trapezoid fills the characteristic CA-CA space
- Peptides **twisted** >30° from planar are severe geometry distortions
- Space is filled with yellow, angle between component planes approximates severity

# Cis Peptides: Probable causes

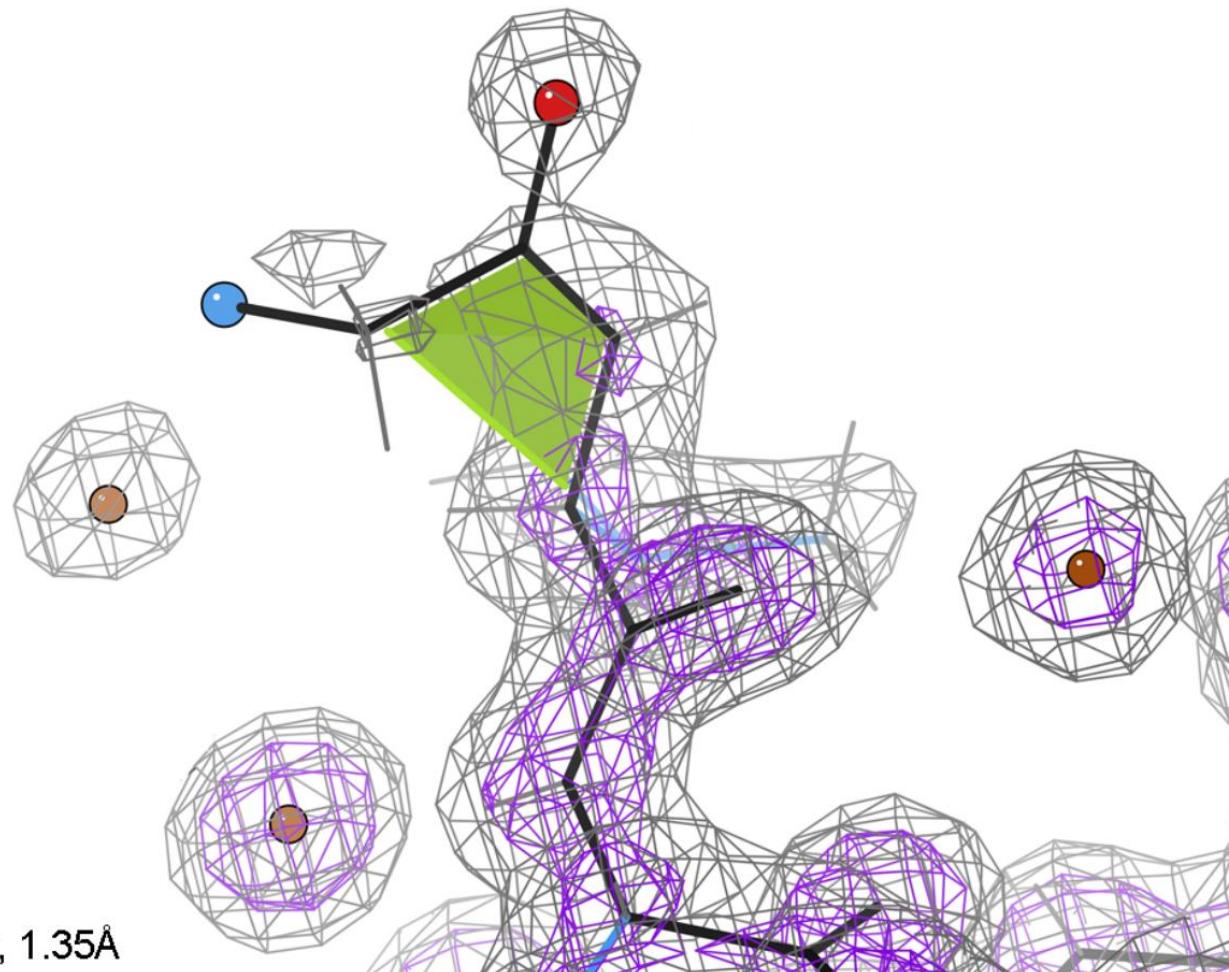
Arg-Gln-Asn-Ser triple *cis*-nonPro -- unjustified



## Fit to small density

- The *cis* CA-CA distance is shorter and **seems** to fit better into fragmented density
- A conformation this rare requires more justification than a marginally better fit
- Flip it to *trans* unless density, chemistry, homology, or another source gives you clear support

# Cis Peptides: Probable causes



## Chain termini

- Non-Pro *cis* peptides at chain ends are always wrong
- Limited density and lack of other constraints *allows* them to be modeled
- But that same lack of constraints means there's nothing to hold an unusual conformation in place

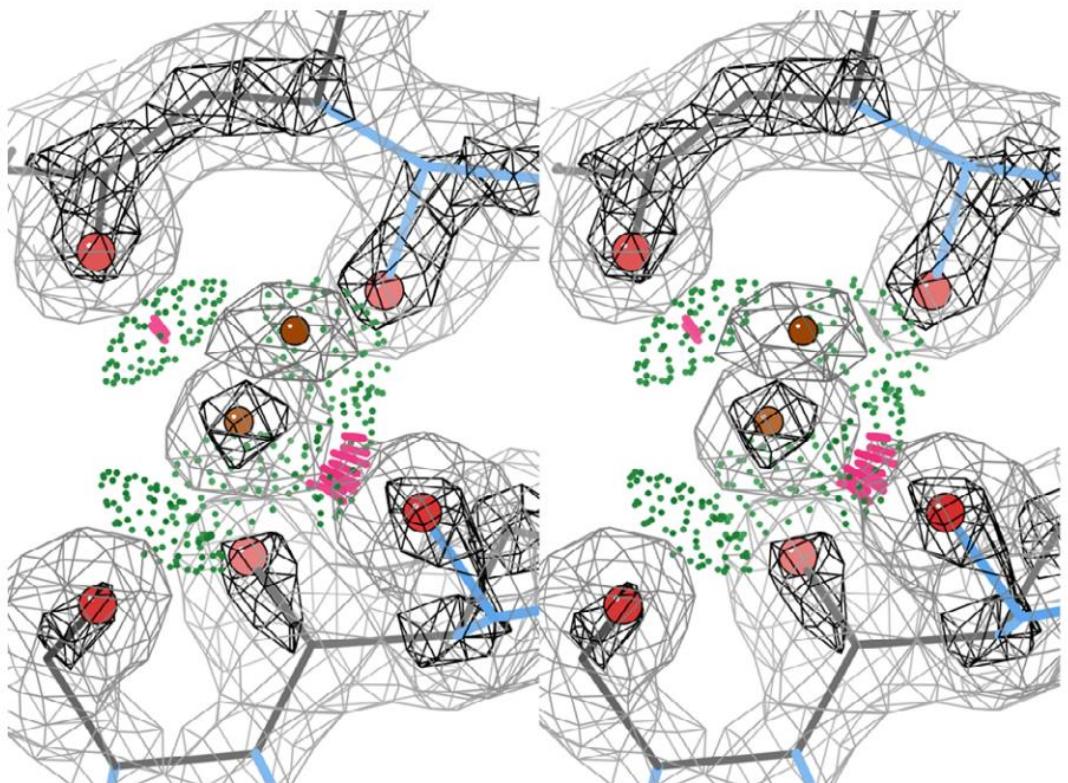
RNA Validations  
Rotameric backbone suites  
Ribose sugar puckers

(see extras for details)

molprobit.rna\_validate  
molprobit.suitename

# Water Validation

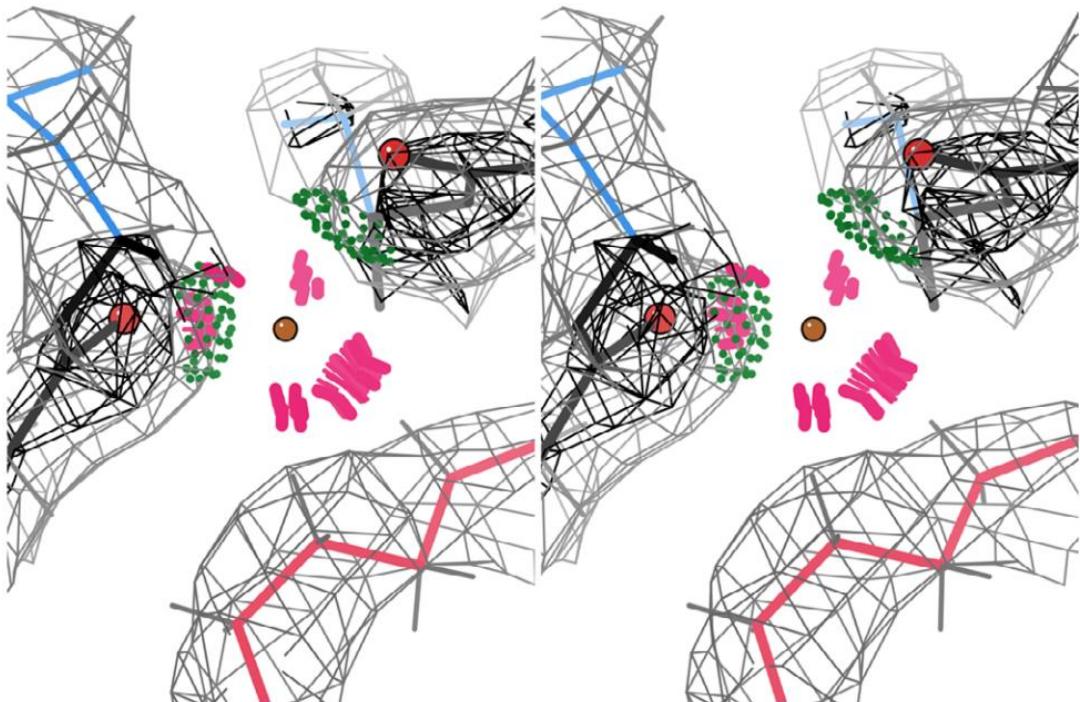
# A water that should be an ion



(Stereo image)  
HOH 606 from 6hhm, 1.23 Å

- Very strong density peak
- Octahedral contact geometry
  - (water is tetrahedral)
- Contacts are all polar groups ( $\delta-$ )
- This is actually a  $+$  ion, probably  $\text{Na}^+$

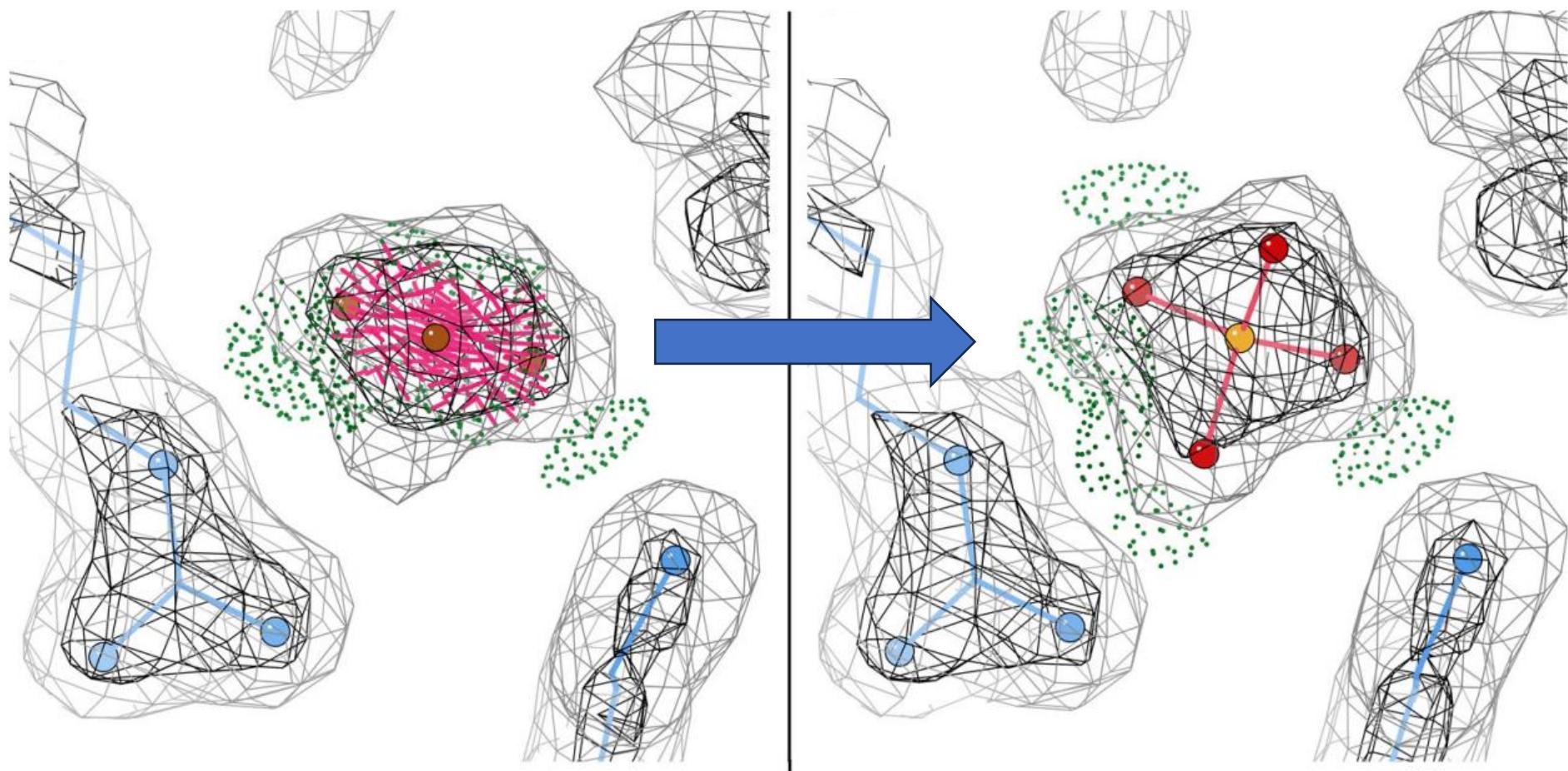
# A water that should not be



(Stereo image)  
HOH 504 from 5onu, 2.22 Å

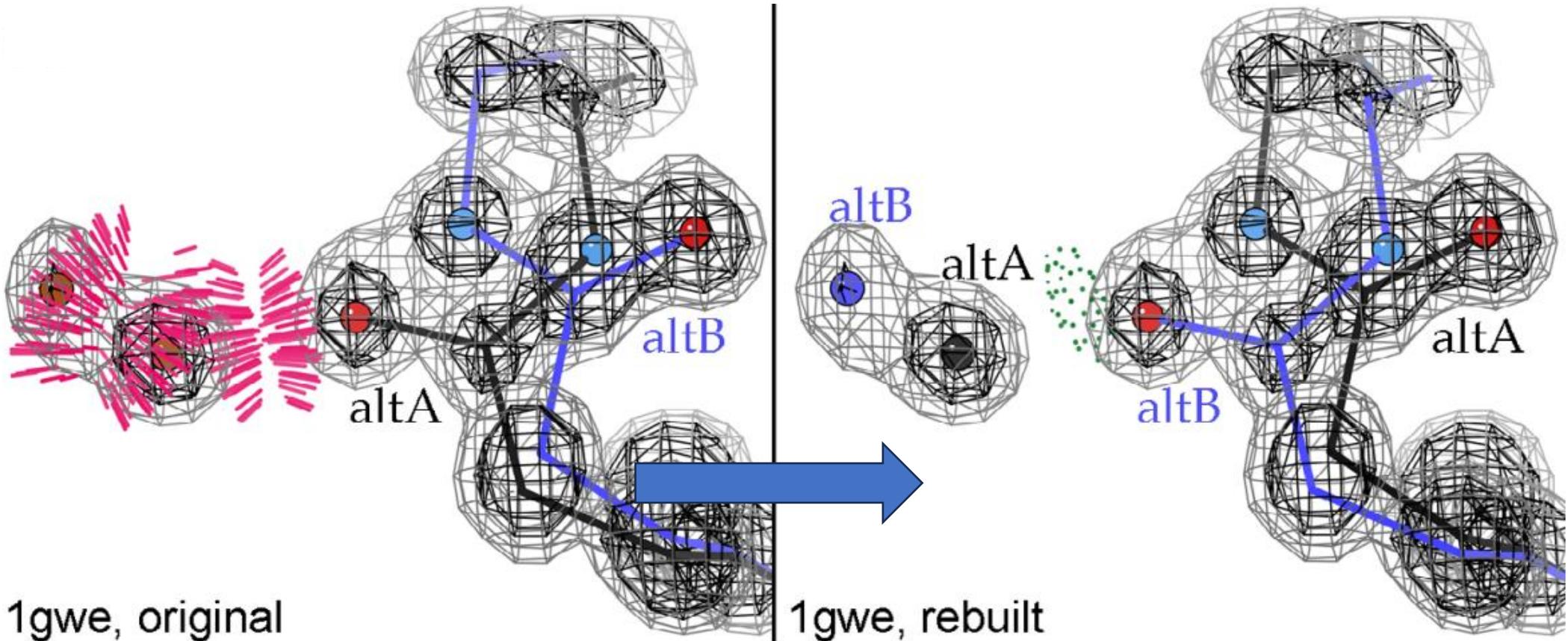
- No density peak
- Mix of polar and non-polar contacts
  - So unlikely to be a coordinated ion
- This water doesn't really exist

# Waters that should be ligands



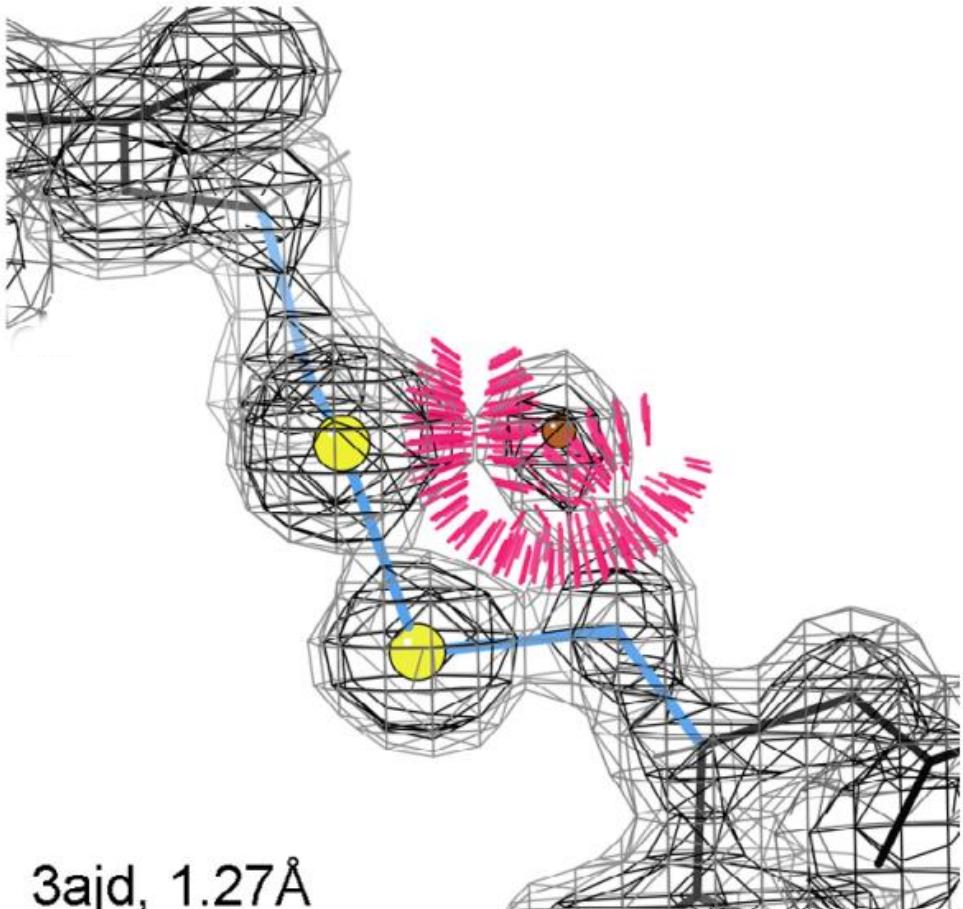
- Densely-clashing waters may actually be a ligand

# Waters that should be partial occupancy



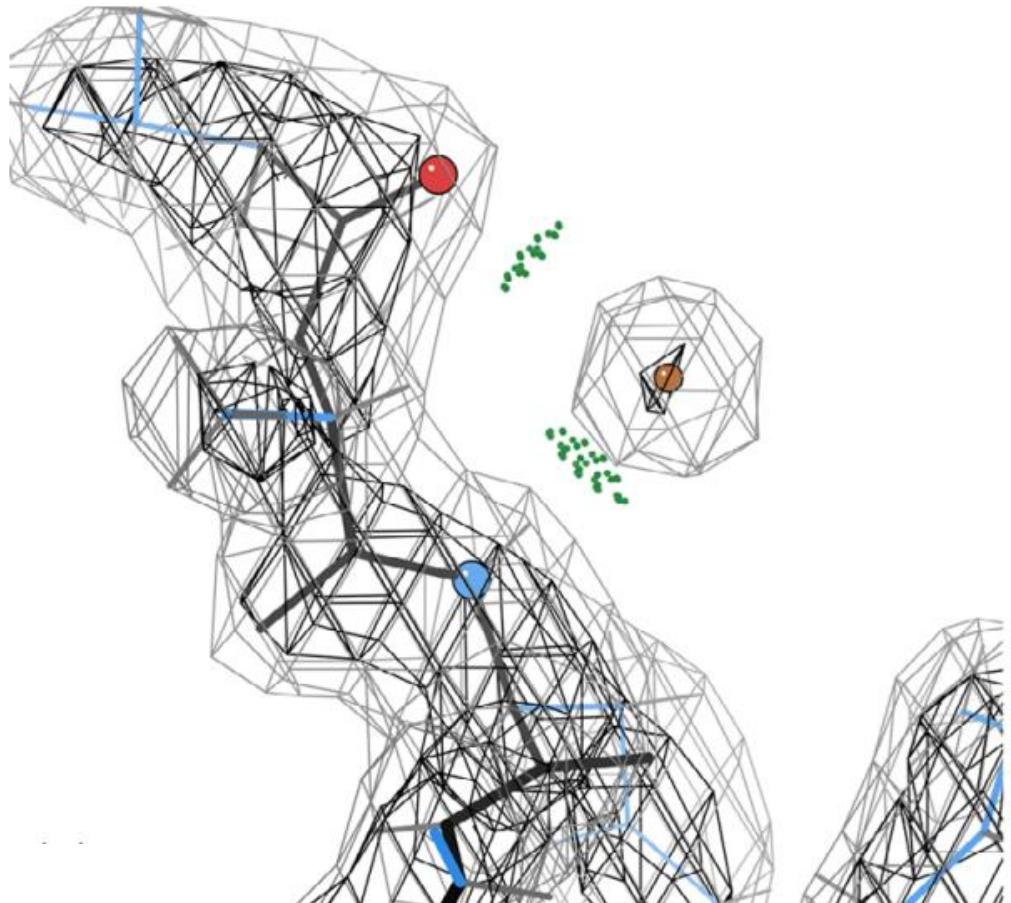
- Densely-clashing waters may actually be part of an alternate conformation network

# Waters that replace alternates



- Very close contacts
  - (Covalent bond distance)
- Clash with non-terminal sidechain atoms
- Could be an unmodeled alternate conformation

# Waters can be real, too!



- Clear density peak
  - Weaker than macromolecule density is fine
- Hydrogen bonds
- Contacts with both  $\delta+$  and  $\delta-$  polar partners, so an ion is unlikely

# MolProbity Score

# MolProbity Score

- The MolProbity Score combines validations and scales the result to look like a resolution
  - Clashscore
  - Ramachandran
  - Rotamers
- MolProbity better than model resolution is good
- MolProbity worse than model resolution is bad

# MolProbity Score

**A single statistic cannot explain a whole structure's quality!**

**Don't rely on it!**

**Especially at low resolution!**

**You now know enough to look at the other statistics**

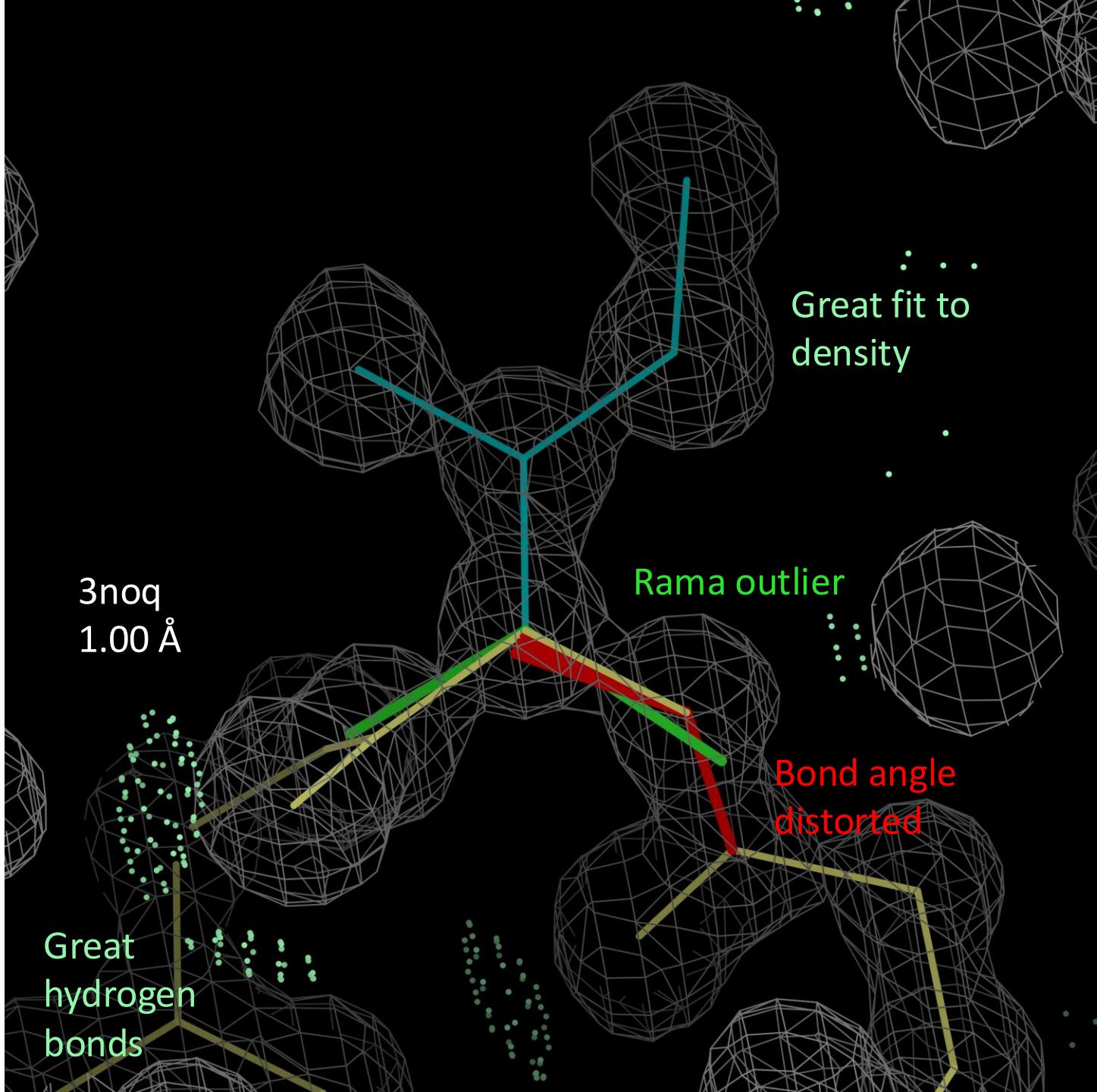
**You now know enough to look at your model and the markup in detail**

# When do you stop?

- Realistically? Do as much as you can.
  - Ideally stop when you – and refinement – can't make the structure better
- Zero outliers is not the goal!
  - Some outliers are justified
  - Some outliers are not justified, but can't be fixed
- If you can't obtain a physically-reasonable solution, consider deleting the region.

# Outliers can be real

- Zero outliers should **not** be the goal.
- Rama outlier, supported by data and environment.



# AlphaFold validation

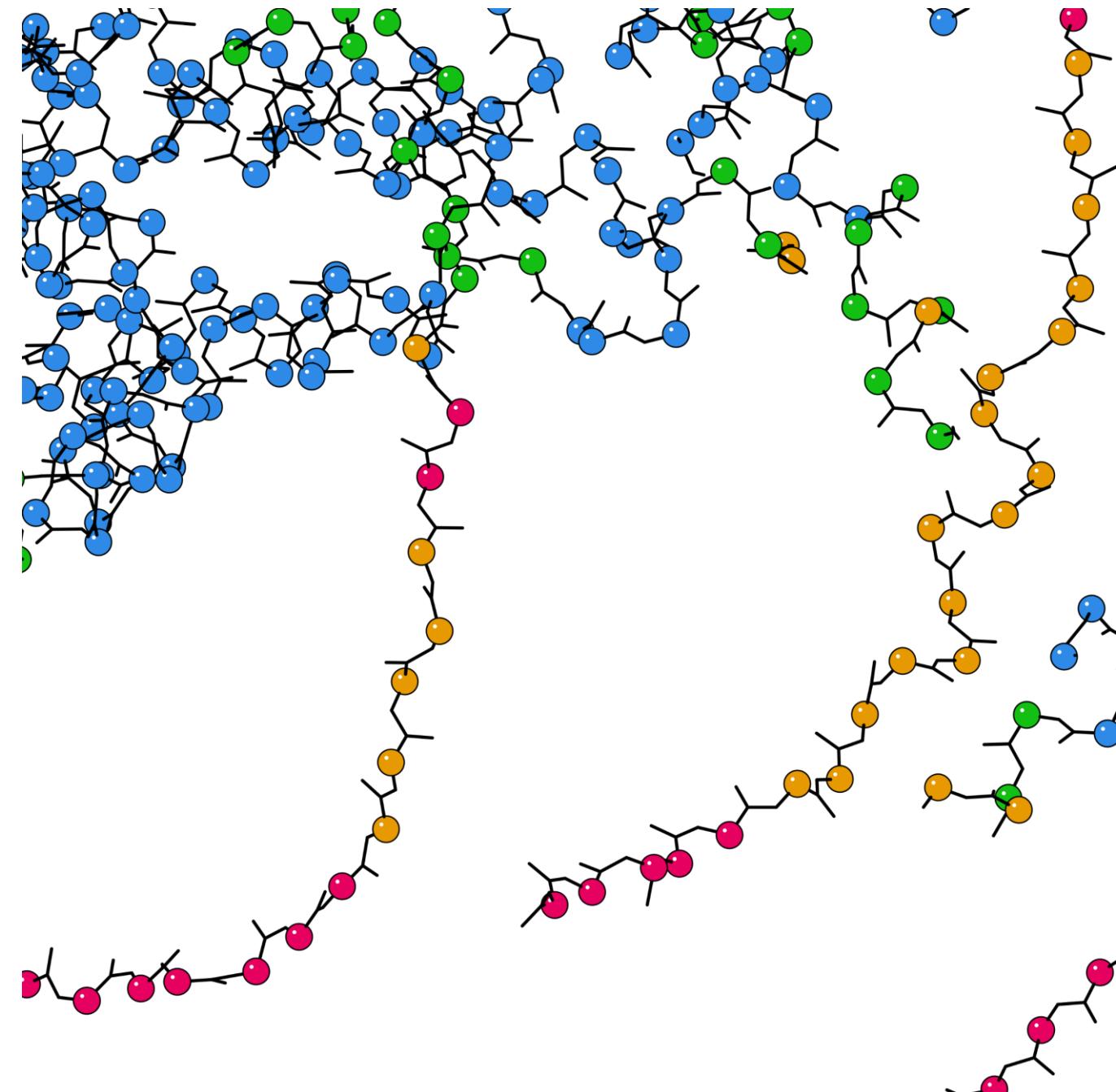
phenix.barbed\_wire\_analysis output.type=kin  
(under development)

# Validation tool

- Predictive (blue)
- Unpacked high pLDDT (gray)
- Near-predictive (green)
- Pseudostructure (gold)
- Barbed wire (hot pink)

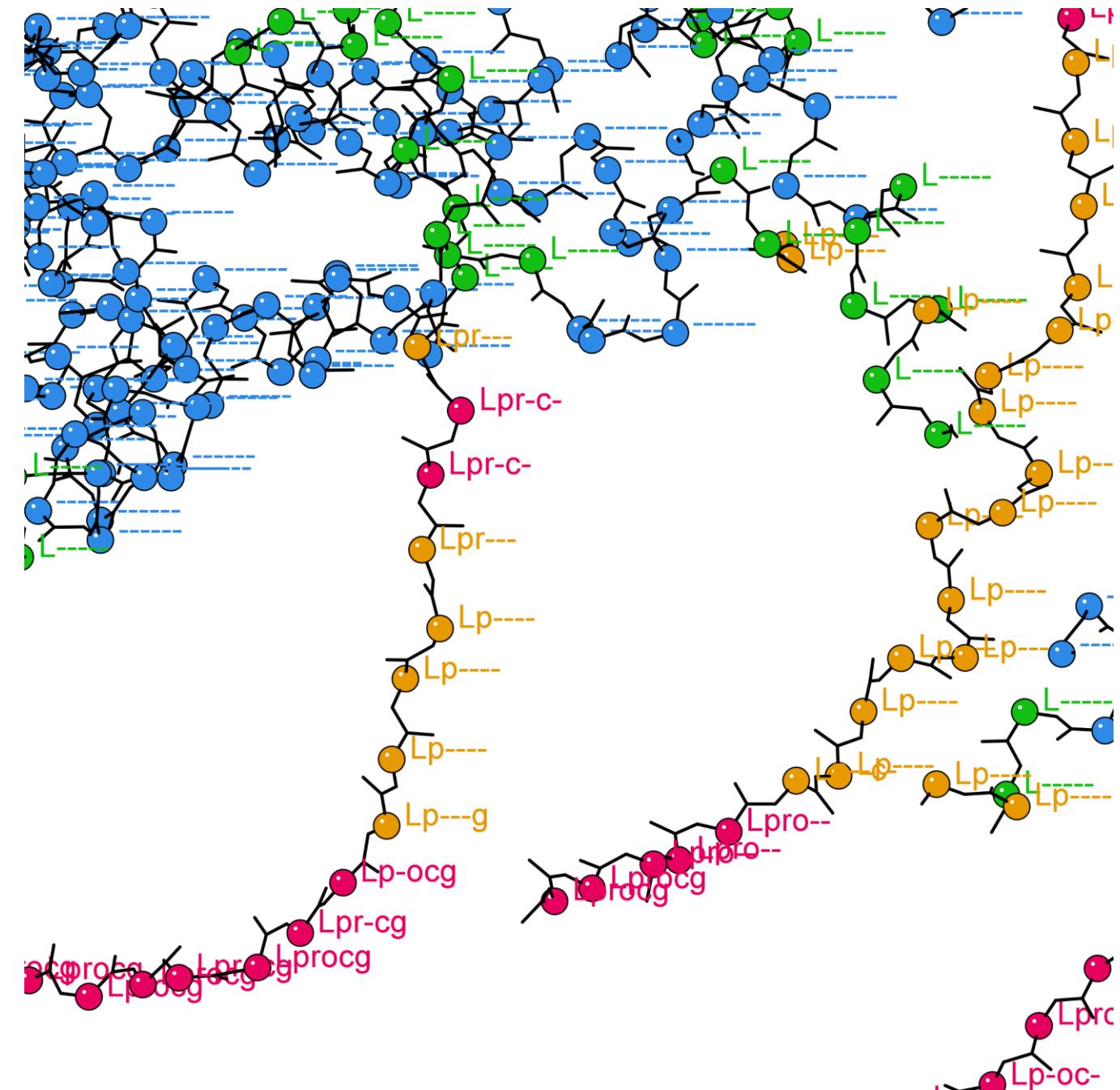
  

- Note barbed wire/unpacked possible transitions



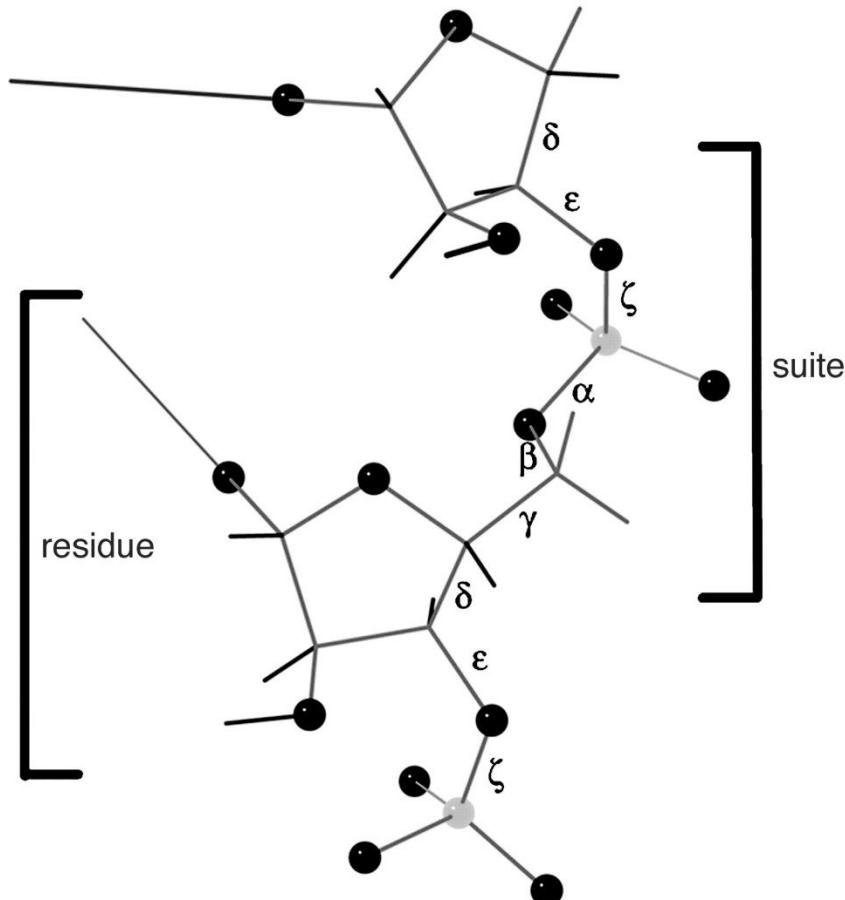
# Validation tool

- Letter codes show assessment of each residue
- More letters = more barbed-wire-like
  - L = low pLDDT
  - p = low packing
  - r = bad Rama
  - o = bad omega (cis)
  - c = bad CaBLAM
  - g = bad bond geometry



# RNA Suites

# RNA Suites: Method



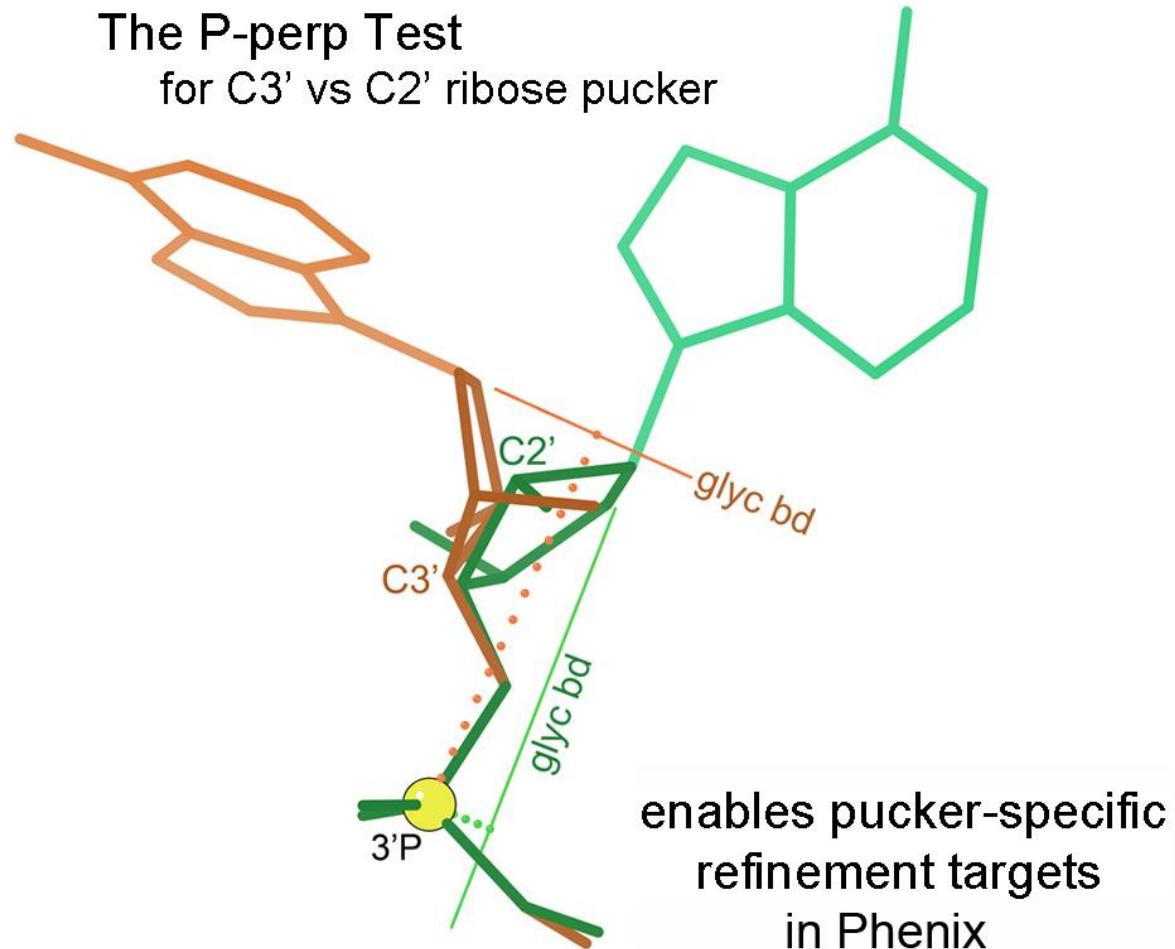
- Useful RNA backbone division is sugar-to-sugar suite, not P-to-P residue
- Suite conformation names are a combination of a number and a letter/character
  - e.g. 1A is the most common A-form helix conformation
- Outliers are named as !!
  - Pronounced “bang, bang”
  - Many !!’s are real, rare conformations

# RNA Ribose Puckers

# RNA Ribose Puckers: Method

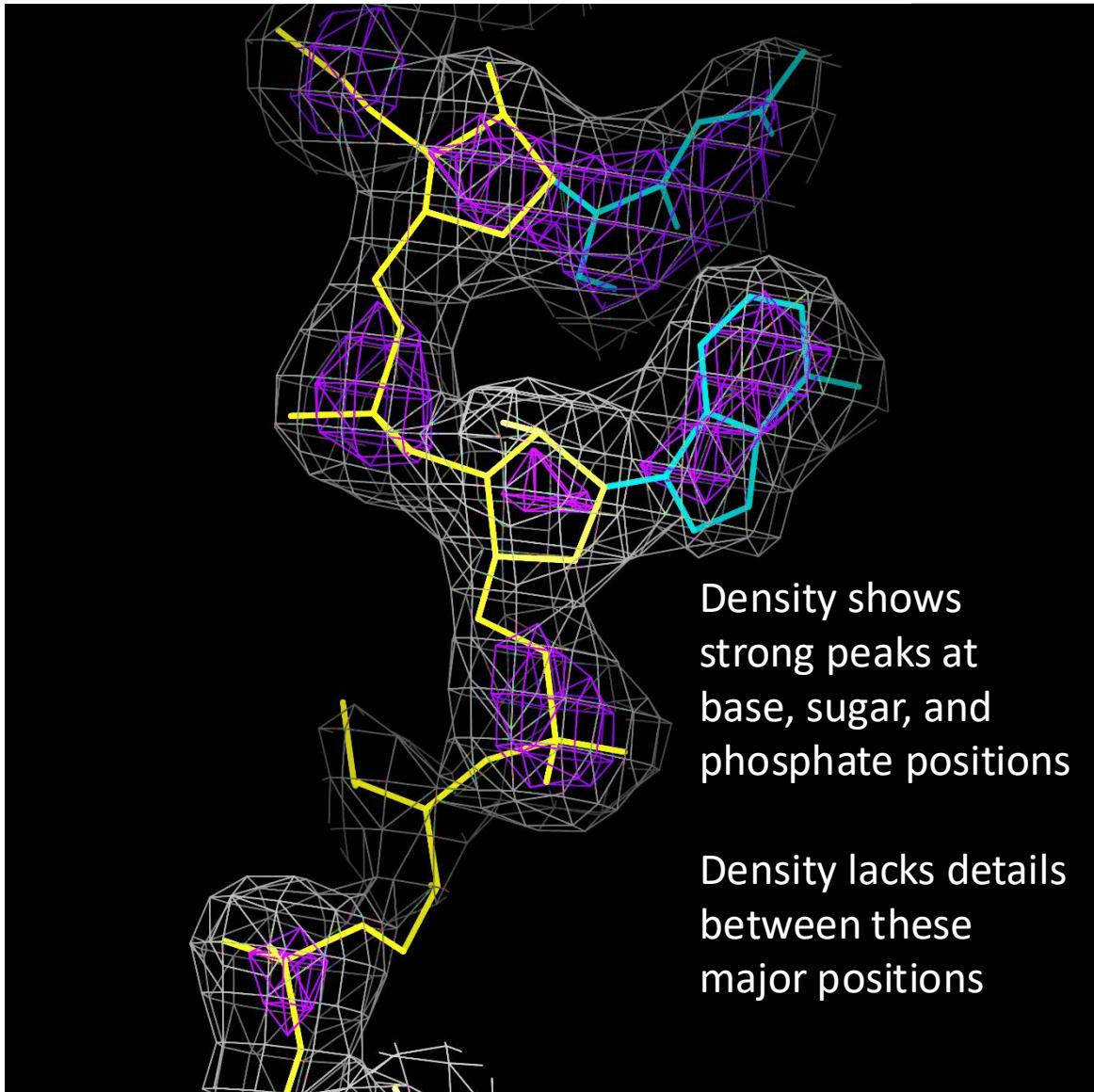
## The P-perp Test

for C3' vs C2' ribose pucker



- The backbone ribose in RNA can have one of two pucker states
  - C2' endo
  - C3' endo
- Ribose pucker correlates very strongly with perpendicular distance from the 3'phosphate to the glycosidic bond vector
  - Glycosidic bond joins ribose sugar to nucleobase
- At low resolution, perpendicular distance is easy to see, ribose pucker is hard to see
- If there's a mismatch, the pucker is probably wrong

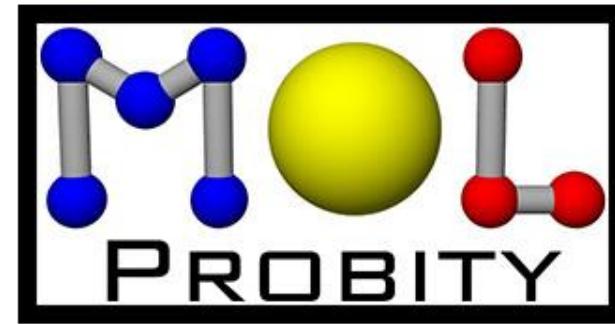
# RNA Errors: Probable Causes



- RNA backbone has many degrees of freedom
- Electron density often leaves RNA backbone underdetermined
  - Even when bases are better resolved
- More tools to help with this are in development

# Resolution and the Limits of Validation

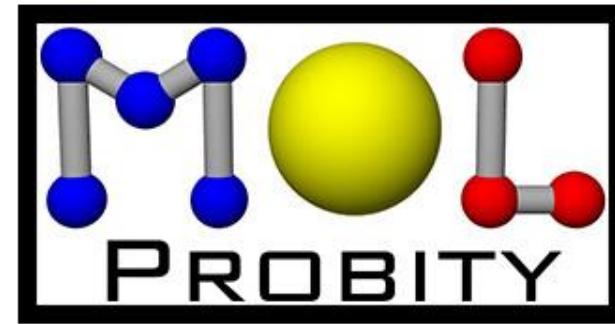
At 1.5 $\text{\AA}$  to 2.5 $\text{\AA}$



MolProbity is still very effective.

The density contains enough specific information  
that where your model fits the density,  
the simple validations (geometry, Rama, rotamers),  
and the explicit-H all-atom contacts

then it's pretty sure to be accurate !

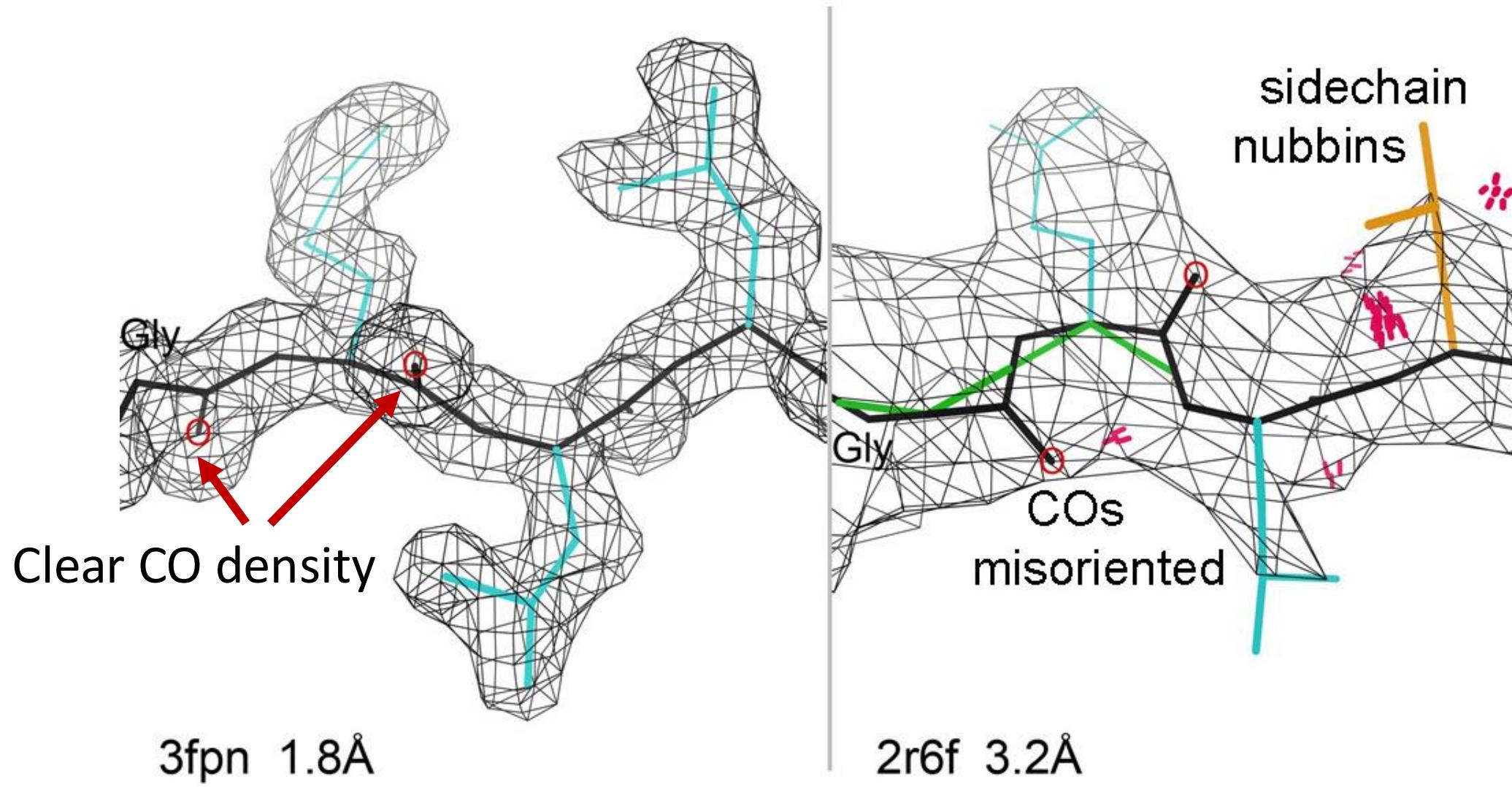


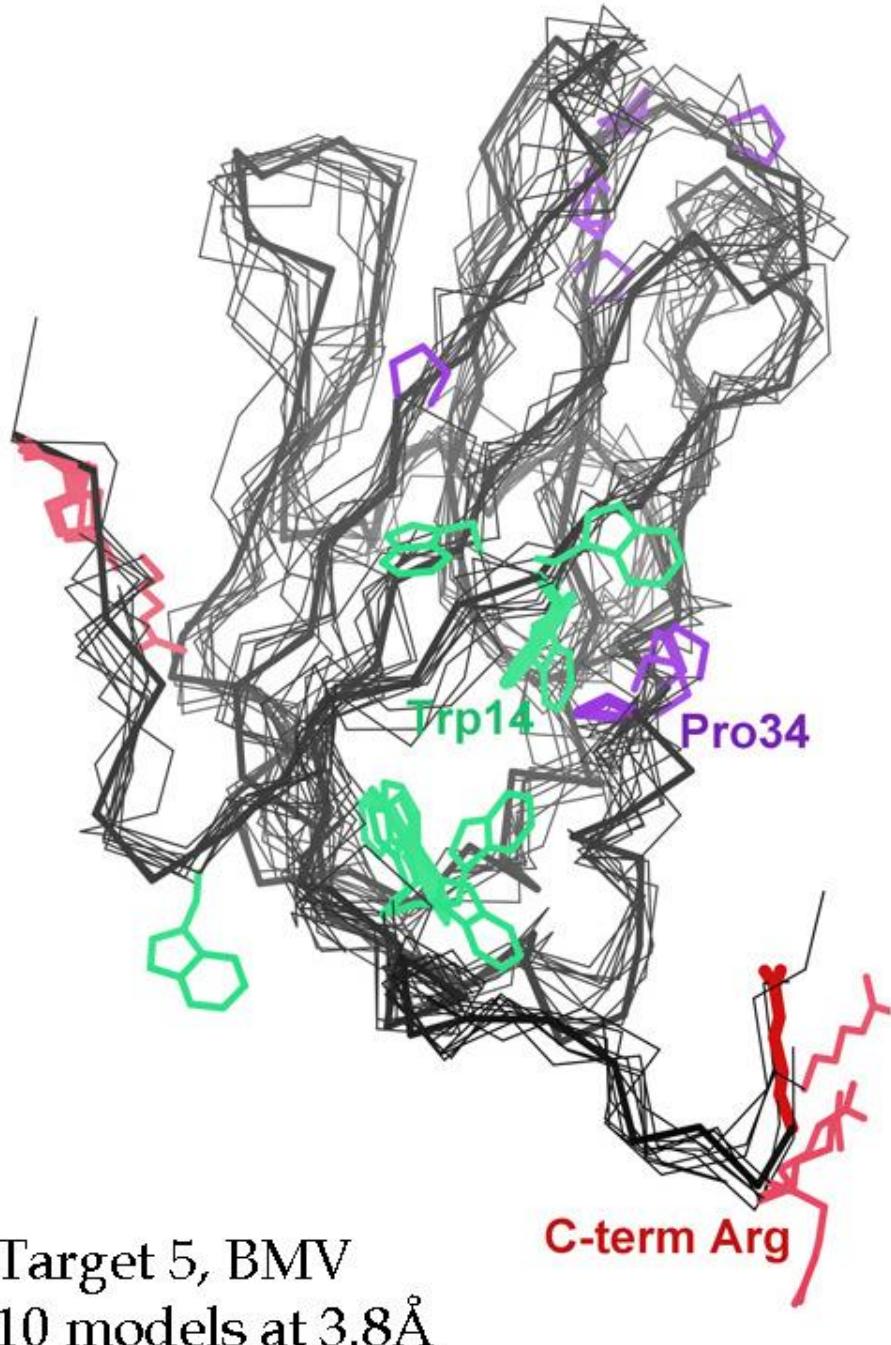
But that's not true at 3 to 4 Å !!

Why does this happen ?

What are we doing about it ?

Tackling lower resolution (2.5 to 4Å)  
Very challenging both for x-ray and for cryoEM





At 3-4 Å,  
many distinct  
models are equally  
compatible with  
the broad density

Much other information  
is needed, which can  
lead to overfitting  
and systematic errors